

RESEARCH ARTICLE

Altered biochemical composition in testis of rabbits during aflatoxicosis

Chaudhari SB

Department of Zoology, Institute of Science Mumbai

Email: sushilkumarbc@gmail.com

Manuscript details:

Received: 24.10.2016
Accepted: 24.12.2016
Published : 31.12.2016

Cite this article as:

Chaudhari SB (2016) Altered biochemical composition in testis of rabbits during aflatoxicosis *International J. of Life Sciences*, 4 (4): 630-634.

Copyright: © 2016 | Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

ABSTRACT

Feeding Aflatoxin-contaminated diet (7.5mg/kg) to young adult New Zealand strain of rabbits for 90 days caused a significant reduction in body weight and weight of testis. Reduction in glycogen ($P < 0.02$) and protein contents ($P < 0.05$). were followed with elevation in lipid ($P < 0.001$) while sodium, Potassium, magnesium and inorganic Phosphorus contents reduced, calcium concentration was increased in the the testis of aflatoxin fed rabbits. Percent ash content increased in aflatoxin fed rabbits indicating accumulation of inorganic constituents. Among monovalent cations, sodium concentration was comparatively higher than potassium ($Na^+/K^+ = 1.62$). Ingestion of aflatoxin contaminated diet for 90 days caused increase in sodium and reduction in potassium contents, as a result the ratio between sodium and potassium was increased (2.78). Glycogenolysis and water loss can be accounted for alterations in K^+ and Na^+ concentrations respectively. It indicates a situation of dehydration. Among divalent cations, calcium concentration was comparatively much less than magnesium ($Ca^{+2}/Mg^{+2} = 0.29$). Feeding aflatoxin contaminated diet to rabbits caused an increase in both calcium and magnesium concentrations, resulting in increased calcium and magnesium ratio (0.30). Some of the key enzyme activities were estimated 1n testis of rabbits during aflatoxicosis. Activity of SDH showed a decrease suggesting decreased operation of mitochondrial TCA cycle. LDH and G-5-pase activities showed significant increase ($P < 0.001$).

Keywords: Aflatoxin, Aflatoxicosis, Testis.

INTRODUCTION

Aflatoxins are secondary toxic fungal metabolites produced by *Aspergillus flavus*. Ingestion of aflatoxin contaminated food/feed stuffs causes occurrence of a serious toxic disease called aflatoxicosis. aflatoxin is potent hepatotoxin (Busby and Wogan,1984) and hepatocarcinogenesis to several animals PC and suspected to be linked to liver cancer in men (Groopman et. al.,1988; Stoloff,1977; Verma and Raval,1992a; Kolhe and Ingale, 2011) in addition dietary aflatoxicosis also caused biochemical changes in liver, heart and kidney (Verma and Kolhe,1997; Kolhe *et al.*, 2006) causing hyperglycemia, hypercalcaemia

(Verma and Chaudhari, 1995) as well as reduced RBC count hemoglobin contained PCV (Verma and Raval, 1992b). Marvan and his colleagues (1983) studied distribution patterns of AFBI residues in goslings and chicken. Results revealed that highest concentrations of AFB₁ residues are present in gonads. Testis is one of the important reproductive organs which perform functions like spermatogenesis. Male sexual act and regulation of male reproductive functions by various hormones. Hence the present investigation was undertaken.

MATERIAL AND METHOD

Aspergillus parasiticus (NRRL3240 obtained from the Indian Agricultural Research Institute, New Delhi, India) was grown on sucrose, magnesium sulfate - potassium nitrate - yeast extract (SMKY) liquid medium at 28 ± 2 °C for 10 days (Diener and Davis, 1966). Culture filtrate, were pooled and extracted with analytical grade chloroform (1:2. v/v) and passed through the bed of anhydrous sodium sulfate. The chloroform extract was evaporated to dryness and the residue dissolved in fresh chloroform, transferred to vials and labelled. For quantitative analysis, 100 µl aflatoxin extract was spotted on silica gel G coated TLC plates along with an aflatoxin standard (a gift from the International Agency for Research on Cancer, Lyon, France) and developed in a solvent comprising toluene: iso- amyl alcohol: methanol (90:32:2. v/v) (Reddy *et al.*, 1970) The air-dried plates were observed under long -wave UV light (360 nm) for aflatoxins. Aflatoxins were confirmed chemically by spraying trifluoro acetic acid and 25 % sulfuric acid. Each spot was scraped separately, dissolved in chilled methanol and subjected to spectrophotometric analysis at 360 nm using a Shimadzu UV- 160A spectrophotometer (Nabney and Nesbitt, 1965). Crude aflatoxin concentrate in chloroform was thoroughly mixed with feed to obtain a concentration of 7.5 mg/kg. This toxin mixed feed was left overnight for complete evaporation of the chloroform. Food for control animals was similarly treated with chloroform alone and analysed for the absence of toxins.

Young inbred New Zealand strain rabbits (*Oryctolagus cuniculus*) weighing approximately 1.2 kg were fed with food and water *ad libitum* and were maintained under laboratory conditions. 10 such rabbits five in

each group) were fed with aflatoxin-contaminated meal (7.5 mg/kg) for 90 days. Group 2 animals received non -toxic feed and served as the controls.

On completion of the treatment, rabbits were sacrificed by cervical dislocation; testis was isolated weighed and processed for biochemical analysis of glycogen (Seifter *et al.*, 1950), protein (Lowry *et al.*, 1951), lipid (Folch *et al.*, 1957) and water (by drying the tissue in anhydric air oven) contents.

A known amount of nonfat dry tissue (NFDI) was ashed in a muffle furnace at 600 degree centigrade for overnight and ash content measured gravimetrically samples were digested in 1 N HCL and analyzed for their Sodium and potassium contents by flame photometry calcium and magnesium by titration (Jackson, 1973) and phosphorus by photo colorimetric method (Jackson, 1973).

Student's t-test was used for statistical analysis of data.

RESULTS

Rabbits fed with aflatoxin contaminated diet showed behavioral alterations like. reduced food and water consumption and weight loss. The rabbits became dull and lethargic with the signs of respiratory disorders. staggering. ataxia of nervous system. Falling of hairs; ear twitching and grinding of teeth were also noted reduced body weight was noted. Results revealed a significant decrease in body weight from 30 days onwards after the treatment. Relative increase in weight also showed a decrease.

The data presented in table biochemical profile of testis indicates altered during aflatoxicosis. Glycogen. protein and water contents were significantly decreased after feeding aflatoxin contaminated diet to rabbits ($P < 0.02$. $P < 0.05$. $P < 0.01$ respectively). Lipid content was significantly increased ($P < 0.001$). The concentration of various inorganic components in testis. It is apparent that the ash content showed an increase in aflatoxin fed rabbits Indicating an accumulation of inorganic constituents.

Among monovalent cations Na⁺ concentration was comparatively higher than K⁺ ($Na^+/K^+ = 1.62$).

Table 1: Effect of feeding aflatoxin contaminated diet on the testis of rabbits

Parameters		Control	Treated
Initial body weight (gm)		1168.80 ± 34.12	1175.60 ± 35.09
Final body weight (gm)		1768.10 ± 21.56	1383.60 ^a ± 23.29
Relative Increase in body weight		51.27	17.69
Testis weight (gm)		7.0950 ± 0.2320	4.9560 ^a ± 0.2638
Testis weight (gm/100gm body wt)		0.4390 ± 0.1381	0.3225 ± 0.2147
Biochemical composition	Glycogen ¹	0.6421 ± 0.0458	0.4110 ^c ± 0.0632
	Protein ¹	7.1743 ± 0.2120	5.9572 ^d ± 0.4210
	Lipid ²	4.9300 ± 0.0934	6.8100 ^a ± 0.1270
	Water ²	79.9500 ± 0.9620	74.8900 ^b ± 0.7390
Inorganic constituents	Ash ³	8.34 ± 0.09	8.97 ± 0.13
	Sodium ⁴	76.08 ± 2.85	121.74 ^a ± 1.95
	Potassium ⁴	46.85 ± 0.57	43.67 ^c ± 0.66
	Sodium/Potassium	1.62	2.78
	Calcium ⁴	153.67 ± 1.47	159.00 ± 3.34
	Magnesium ⁴	514.02 ± 2.21	522.38 ^b ± 0.66
	Calcium/Magnesium	0.29	0.30
	Inorganic phosphorus ⁴	826.67 ± 12.88	826.53 ± 10.16

Values are mean ± S.E.M.; n = 5.

Significant at: a = P < 0.001; b = P < 0.01; c = P < 0.02; d = P < 0.05

Values expressed as: 1. Mg % fresh weight; 2. Mg % dry weight; 3. Mg % non-fat dry tissue, 4. Meq/1000 gm non-fat dry tissue

Induced aflatoxicosis caused an increase in sodium and reduction in potassium contents, as a result the ratio between sodium and potassium was increased (2.78). Glycogenolysis and water loss can be accounted for alterations in K⁺ and Na⁺ concentrations, respectively. It indicates a situation of dehydration with loss of water along with salts. Among divalent cations, calcium concentration was comparatively much less than magnesium (Ca²⁺/Mg²⁺ = 0.29). Feeding aflatoxin contaminated feed to rabbits caused an increase in both calcium and magnesium concentrations, resulting in increased calcium and magnesium ratio (Ca²⁺/Mg²⁺ = 0.30). Feeding aflatoxin contaminated diet caused slight decrease in inorganic phosphorus. Table also shows activities of enzymes such as SDH, LDH and G-6-pase in the testis of the control and aflatoxin fed rabbits. SDH activity showed a decrease suggesting a decreased operation of mitochondrial TCA cycle. LDH and G-6-pase activities showed significant (P < 0.001) increase.

DISCUSSION

Significantly decreased glycogen concentration during aflatoxicosis in rabbits could be due to the effect of aflatoxin on synthetic enzymes, inhibition of

glycogenesis or increased glycogenolysis. Reduction in glycogen content coincides with decreased K⁺ and increased Ca²⁺ concentrations: later is known to enhance glycogen phosphorylase activity (Mc Gilvery, 1979). Intraperitoneal administration of aflatoxin to day-old chicks reduced UDP-glucose, glycogen transglucosylase activity (Shankaran *et al.* 1970). Reduction in protein concentration can occur due to alteration in the mechanism of gene expression and changes in DNA dependent RNA synthesis (Clifford and Rees, 1967; Newberne and Butler, 1969; Edds, 1973). and protein synthesis (Sarasin and Moule, 1975; 1976; White and Rees, 1980; Jeffery *et al.* 1984). Decreased K⁺ can be correlated with reduced protein synthesis (Harper, 1963, Verma *et al.* 1998). This reduction in protein content may be due to reduced synthesis as mentioned above and/or its increased utilization in gluconeogenesis in liver and kidney (Kolhe, 1994). Increased accumulation of lipid content in aflatoxin fed rabbits might be due to fatty infiltration in testis. Accumulation of lipid in the liver of chicks and ducklings fed with aflatoxin contaminated diet have been reported (Shank and Wogan, 1966; Carnaghan *et al.* 1966; Newberne and Butler, 1969). Reason for this accumulation is not clearly understood. Decreased water content may be due to dehydration.

Feeding aflatoxin contaminated meal caused alteration in Na⁺/K⁺ ratio (Verma *et al.*, 1998) which indicates permeability alterations. It could be due to a decreased activity of sodium pump, responsible for maintaining higher concentration of K⁺ intracellularly with expulsion of Na⁺. It is an energy requiring step. Reduced SDH activity during aflatoxicosis might be causing reduction in stored energy and hence activity of sodium pumps. Further, peroxidation or membrane lipids during aflatoxicosis initiates loss of membrane integrity and membrane bound enzyme activities (Younes and Seigers, 1984; Pasquali-Rouchetti *et al.*, 1979) which in turn brought about a disturbance in cellular homeostasis. Increased Na⁺ concentration could be correlated with increased accumulation of water intracellularly. Also reduced K⁺ concentration in the cells could be correlated with increased glycogenolysis (Williams *et al.*, 1971; Luly *et al.*, 1972). Among the divalent cations, Ca⁺² concentration was comparatively low than Mg⁺² (Ca⁺² /Mg⁺² = 0.29). During aflatoxicosis calcium concentration registered a little increase while magnesium showed significant increase. Increased concentration of Ca⁺² in cytosol, mitochondria and microsomal fraction as a result of AFB₁ treatment have been reported by Toskulkao and Glinsukon (1988). Increased accumulation inside the mitochondria causes mitochondrial dysfunction and reduction in ATP content.

SDH is a key enzyme of Krebs cycle which is primarily concerned with aerobic oxidation of Acetyl Co-A to produce energy. Activity of SDH showed decrease during aflatoxicosis in rabbits. It could be due to reduction in oxygen transport to the tissues. Aflatoxin induced alterations in mitochondrial activity (Toskulkao and Glinsukon, 1988) and mitochondrial swelling has been reported (Roy, 1968).

It is concluded that aflatoxin contaminated diet caused alterations in testis during aflatoxicosis in rabbits.

Acknowledgement

The authors are grateful to Dr M.D. Friesen of the International Agency for Research on Cancer, Lyon, France, for providing samples of pure aflatoxins. And to Prof. R. J. Verma, Dept of Zoology, Gujrat University, Ahmedabad for guidance for this study.

Conflicts of interest: The authors stated that no conflicts of interest.

REFERENCES

- Busby WF and Wagon GN (1984) Alatoxins in chemical carcinogens (Ed. Searle, S.E) ACS Manograph.182, American chemical society, Washington D.C. PP. 945 - 1136.
- Butler WH, Greenblatt M and Lijinsky W (1969) Carcinogenesis in cats by aflatoxins B₁,G₁ and B₂. Cancer Res. 29: 2206-2211.
- Carnaghan, RBA, Lewis G, Patterson DSP and Allcroft R (1966) Biochemical and pathological effects of groundnut poisoning in chicken. Pathol. Vet. (Base 1). 3: 601-615.
- Clifford J and Rees KR (1967) The action of aflatoxin B₁ on the rat liver. *Biochem. J.*,102: 65-75.
- Diener UL and Davis ND (1966) Aflatoxin Production by isolates of *Aspergillus flavus*. *Phytopathology*56:1390 - 1393.
- Folch-Pij, Less M and Sloane Stanley GH (1957) A simple method for the isolation and purification of total lipid from animal tissues. *J Biol Chem.* 226:497-509.
- Gropman JD, Cain LG and Kensler TW (1988). Aflatoxin exposure in human population: measurement of relationship to cancer. *CRC critical Rev. Toxicol.*19: 113 - 145.
- Harper AE (1963). Glucose-6-Phosphatase. In: Methods of enzymic analysis. Bergmeyer. Hans-Ulrich (ed), Academic Press. New York and London. pp. 788-792.
- Jackson ML (1973) Soil chemical analysis, New Delhi : Hall of India Pvt. Ltd;
- Jeffery FH, Marton JG and Miller JK (1984) Effect of some clinically significant mycotoxins on the incorporation of DNA. RNA and protein precursors in cultured mammalia cells. Res. Vet. Sci. 37:30-38.
- Kolhe AS and Ingale SR (2011) Effect of Mycotoxins in Human and Animal System. *Journal of Research & Development*, 1(2): 22-28.
- Kolhe AS, Verma RJ, Chaudhari SB and Bhole RV (2006) Effects of Dietary Aflatoxicosis of Biochemical composition of kidney. *Research Link*25, IV (8): 134-136.
- Lowry DH, Rosenbrough NI, Farr A and Randall J (1951) Protein measurement with the folin phenol reagent. *J. Biol. Chem.* 193 :265-275.
- Luly P, Barnabei C. and Tria E (1972) Hormonal control in vitro of plasma membrane bound (Na⁺ -K⁺) ATPase of rat liver *Biochem. Biophys. Acta.*282: 447-452.
- Marvan F, Vernerova E Samek M, Relsnerova H, Nemej J and Martakova R (1983) Aflatoxin B₁ (AFB₁) residues in the organs of young poultry. *Biol. Chem. Vet. (Praha)*, 24: 85-92.
- McGilvery RW (1979) *Biochemistry, ~ functional approach.* 2nd edition, W.B. Saunders Company, Philadelphia.
- Nabney J and Nesbitt, BF (1965) A. spectrophotometric, Thin layer chromatography of aflatoxin. *Anal. Biochem.*38: 568 - 571.

- Newberne PM and Butler WH (1969) Acute and chronic effects of aflatoxin on the liver of domestic and laboratory animals - A review. *Cancer Res.* 29: 236-250.
- Pasquali-Rouchetti I, Bini. A., Botti. B, DeAlo]sl G, Fornicri. C and Vannini V (1979) Ultrastructural and biochemical changes induced by progressive lipid peroxidation on isolated microsomes and rat liver endoplasmic reticulum. *Lab. Invest.*, 42: 457-466.
- Reddy TV, Vishwanathan, L, and enkitasubramanian, T A.(1970) Thin layer chromatography of aflatoxins. *Anal Biochem.*38 :568-571.
- Roy AK (1968) Effect of aflatoxin on polysomal profiles and RNA synthesis in rat liver. *Biochem. Biophys Acta.* 169: 206-211.
- Sarasin A and Moule Y (1975) Translational step inhibited in vivo by aflatoxin B1 in rat-liver polysomes. *Eur.J.Biochem.*54: 329-340.
- Sarasin A and Moule Y (1976) Helical polysomes induced by aflatoxin B1 in vivo A new hypothesis for helix formation by chemicals and carcinogen. *Experimental Cell Research.* 97: 346-358.
- Seifter S, Dayton S, Novic B and Muntwyler V (1950) The estimation of glycogen with anthrone reagent. *Arch. Biochem.* 25: 191-200.
- Shank RC and Wogan GN (1966) Acute effects of aflatoxin B1 on liver composition and metabolism in the rat and duckling. *Toxicol. Appl. Pharmacol.* 9: 468-476.
- Shankaran R, Raj HG, Venkitasubramanian TA (1970) Effect of aflatoxin on carbohydrate metabolism in chick liver. *Enzymologia.* 39, 371-378.
- Stoloff L (1977) Aflatoxin - an overview. In *Mycotoxins in Human and Animal Health.* Rodricks. J .C.,Hesseltine. C.W., Mehlman. M.A. (eds.). Pathotox Publishers, Park Forest South I. pp. 16-28.
- Toskulkao C and Glinsukon T (1988) Hepatic lipid peroxidation and intracellular calcium accumulation in ethanol potentiated aflatoxin B1 toxicity. *J.Pharmacobio. Dyn.*11: 191-197.
- Verma RJ and Chaudhari SB (1995) Hypercalcaemia during aflatoxicosis. *Med. Sci. Res.* 23, 703-704.
- Verma RJ, Chaudhari SB, Kolhe AS and Nair Anita (1998) Hyperkalaemia during aflatoxicosis . *Med. Sci. Res.* 26, 733-35
- Verma RJ and Rawal PJ (1992b) Alteration in erythrocytes during induced chronic aflatoxicosis in rabbits., *Bull. Environ. Contam.Toxicol.*;49 : 861-865.
- Verma RJ and Rawal PJ (1992a) Impact of Aflatoxin on Human beings and Animals, *Indian Rev Life Sci.*;12 : 235.
- Verma RJ and Kolhe AS (1997) Effect of dietary aflatoxicosis on biochemical composition of heart. *Proc. Nat. Acad. Sci. India.*,67(B) III & IV: 239-242.
- Williams TF, Exton JA, Friedmarm N and Park CR (1971) Effects of insulin and adenosine-3'5'-monophosphate on K flux and glucose output in perfused rat liver. *Am.J. Physiol.*,221: 1645-1651.
- Younes M and Seigers CP (1984) Interrelation between lipid peroxidation and other hepatotoxic event. *Biochem. Pharmacol.*, 33: 2001-2003.