

## RESEARCH ARTICLE

# Incidence of mycotoxins in poultry feeds and feed ingredients used in Warangal (TS), India

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**ABSTRACT**

Incidence of mycotoxins is a major problem encountered by poultry industry as in most cases low grade feed ingredients are used in the formulation of feeds. Investigations carried out to assess the incidence of mycotoxins in poultry feeds revealed that aflatoxin followed by ochratoxin A were most common and contaminated large number of poultry feeds and feed ingredients. Most of the samples were found to be contaminated with more than one mycotoxin. Farm mixed feed samples were recorded with highest frequencies of mycotoxin contamination than the commercial brands,

**Keywords:** Poultry feeds, mycotoxins, aflatoxin, ochratoxin A, broilers, layers.

**INTRODUCTION**

Contamination of food and agricultural commodities by various types of toxigenic molds is a serious nonetheless a widely neglected problem. Regardless of decades of extensive research, mold infestation still remains a challenging problem (Munkvold, 2003). It has been estimated by Food and Agriculture Organization (FAO) that worldwide, approximately 25% of the crops get contaminated by moulds and are affected by mycotoxins (Richard *et al.*, 1989; Rice and Ross, 1994), and the estimated loss extends to billions of dollars (Trail *et al.*, 1995). Poor harvesting practices, improper drying, handling, packaging, storage, and transport conditions contribute to fungal growth and increase the risk of mycotoxin production. Contamination of agricultural commodities by fungi results not only in the downgrading nutrient content but also cause a health hazard for human, livestock and poultry birds. The most notable importance of contamination of agricultural commodities by toxigenic fungi is buildup of mycotoxin concentrations to injurious levels in food and feeds (Perrone *et al.*, 2007). Poultry feed are formulated from a mixture of ingredients that include cereal grains, cereal byproducts, fats, plant protein sources, animal byproducts, vitamins, mineral supplements, crystalline amino acids and feed additives. The composition of the feed varies according to type of poultry bird (broiler or layer) and also age of the bird.

Since poultry feeds are rich in all types of nutrients easily to prone to microbial growth at appropriate conditions. Defects in formulations, faulty storage conditions and unseasonal rains promote the infestation by microorganisms. Of the potential contaminants, mycotoxigenic fungi are the most widespread and notorious particularly in hot, humid conditions. In view of the importance of mycotoxins in the health of poultry birds and a range of poultry feeds used, it was thought worthwhile to investigate the natural incidence of mycotoxins in different poultry feeds used in Warangal District, which is a major center for poultry industry in Telangana State.

## MATERIALS & METHODS

### Isolation and enumeration of mycotoxigenic fungi associated with poultry feed, feed ingredients

#### Study region

Samples of commercial poultry feeds, feed ingredients were collected from poultry farms located in Warangal district and the respective feed manufacturers located in other places also. In case of farm mixed poultry feeds, individual feed ingredients along with mixed feeds were also collected.

#### Sampling criteria

Samples of commercially prepared poultry feeds, farm mixed poultry feeds and feed Ingredients were collected through systematic random sampling from grain markets, feed ingredient suppliers, poultry farms and feed manufacturers. For each sample, three initial samples of three kg each were collected from different places. After thorough mixing, a composite sample of 1 kg placed in a sterile polythene bag was brought to the laboratory and they were refrigerated until further use. Conditions of the sample at the collection time were recorded.

#### Natural incidence of mycotoxins in poultry feeds.

Natural contamination of commercial poultry feeds was carried out by "rapid TLC method for multi-mycotoxin analysis" as suggested by Devegowda and Raju (2005).

Different mycotoxins produced by species of *Aspergillus*, *Fusarium* and *Penicillium* were detected by standard methods. Aflatoxins (Stack and Pohland,

1975); ochratoxin-A (Gimeno, 1979); sterigmatocystine (Adey and Mateles, 1964); penicillic acid (Gorst-Allman and Steyn, 1979); deoxynivalenol and nivalenol (Ramakrishna and Bhat, 1987); moniliformin, T-2 toxin and zearalenone (Kamimura *et al.*, 1981)

## RESULTS AND DISCUSSIONS

### 1. Broiler feeds

The mould infested broiler feed samples collected from different broiler poultry farms of Warangal district were analyzed for the presence of mycotoxins and the results are presented in Table 1. From table it is evident that the contamination of different broiler feeds with mycotoxins is a common feature. Out of 35 broiler concentrate feed samples tested for the presence of mycotoxins, 7 samples were positive for aflatoxins, 3 samples for ochratoxin A, 1 sample for zearalenone and 7 samples for T- 2 toxin. Sterigmatocystin was not found in broiler concentrate feed samples. Similarly, 28 samples of broiler concentrate feed with maize were positive for aflatoxins and 13 for ochratoxin A, 2 for zearalenone and 8 for T-2 toxin, out of 35 samples were screened. Sterigmatocystin was not detected in broiler concentrate feed with maize.

Out of 35 broiler pellet feed samples screened for presence of different mycotoxins, 9 samples were positive for aflatoxins, 2 for ochratoxin A and 1 for T-2 toxin. Zearalenone and sterigmatocystin were not found in the broiler pellet feed samples tested. Similarly, out of 35 broiler pre starter crumbs samples screened for mycotoxin analysis, 3, 2, 1 and 1 samples were positive for aflatoxins, ochratoxin A, zearalenone and for sterigmatocystin respectively. However T-2 toxin was not detected in the samples screened. No toxin was reported in broiler starter feed samples.

### 2. Layer feeds

Different contaminated layer feed samples were collected from different layers poultry farms of Warangal district and screened for presence of different mycotoxins and the results are presented in Table 2. From the table it is evident that out of the different layer feed samples screened for presence of mycotoxin contamination, homemade feed (mixed at the farm) samples were highly contaminated with

different mycotoxins. Screening of 35 mixed layer feed samples indicated that 25 samples were positive for

**Table 1 : Natural contamination of different broiler feeds with mycotoxins**

Name of the broiler feed sample tested	Number of feed samples tested	Number of positive feed samples	% of incidence	Toxin detected
Broiler concentrate feed	35	7	20.0	Aflatoxins
		3	8.6	Ochratoxin A
		1	2.9	Zearalenone
		7	20.0	T-2 toxin
		ND	--	Sterigmatocystin
Broiler concentrated feed with maize	35	28	80.0	Aflatoxins
		13	37.1	Ochratoxin A
		2	5.7	Zearalenone
		8	22.9	T-2 toxin
		ND	--	Sterigmatocystin
Broiler pellet feed	35	9	25.7	Aflatoxins
		2	5.7	Ochratoxin A
		ND	--	Zearalenone
		1	2.9	T-2 toxin
		ND	--	Sterigmatocystin
Broiler pre starter crumbs	35	3	8.6	Aflatoxins
		2	5.7	Ochratoxin A
		1	2.9	Zearalenone
		ND	--	T-2 toxin
		1	2.9	Sterigmatocystin

ND = Not detected

**Table 2 : Natural contamination of different layer feeds with mycotoxins**

Name of the layer feed sample tested	Number of feed samples tested	Number of positive feed samples	% of incidence	Toxin detected
Home made feed (mixed feed)	35	25	71.4	Aflatoxins
		13	37.1	Ochratoxin A
		2	5.7	Zearalenone
		5	14.3	T-2 toxin
		3	8.6	Sterigmatocystin
Chick feed	35	5	14.3	Aflatoxins
		2	5.7	Ochratoxin A
		ND	--	Zearalenone
		1	2.9	T-2 toxin
		ND	--	Sterigmatocystin
Developer feed	35	8	22.9	Aflatoxins
		3	8.6	Ochratoxin A
		1	2.9	Zearalenone
		2	5.7	T-2 toxin
		ND	--	Sterigmatocystin
Grower feed	35	11	31.4	Aflatoxins
		3	8.6	Ochratoxin A
		1	2.9	Zearalenone
		--	--	T-2 toxin
		1	2.9	Sterigmatocystin
Perlay feed	35	9	25.7	Aflatoxins
		ND	--	Ochratoxin A
		ND	--	Zearalenone
		3	8.6	T-2 toxin
		ND	--	Sterigmatocystin

**Table 3 : Natural contamination of different poultry feed ingredients with mycotoxins**

Name of the feed ingredient tested	Nummber of samples tested	Number of positive samples	% of incidence	Toxin detected
Maize seed	50	34	68.0	Aflatoxins
		13	26.0	Ochratoxin A
		2	4.0	Zearalenone
		8	16.0	T-2 toxin
		9	18.0	Sterigmatocystin
Sunflower meal	50	22	44.0	Aflatoxins
		3	6.0	Ochratoxin A
		1	2.0	Zearalenone
		4	8.0	T-2 toxin
		ND	--	Sterigmatocystin
Groundnut cake	50	28	56.0	Aflatoxins
		13	26.0	Ochratoxin A
		3	6.0	Zearalenone
		2	4.0	T-2 toxin
		ND	--	Sterigmatocystin
Deoiled rice bran	50	14	28.0	Aflatoxins
		8	16.0	Ochratoxin A
		6	12.0	Zearalenone
		4	8.0	T-2 toxin
		1	2.0	Sterigmatocystin
Fish meal	50	3	6.0	Aflatoxins
		ND	--	Ochratoxin A
		ND	--	Zearalenone
		ND	--	T-2 toxin
		1	2.0	Sterigmatocystin
Soybean meal	50	9	18.0	Aflatoxins
		6	12.0	Ochratoxin A
		5	10.0	Zearalenone
		4	8.0	T-2 toxin
		ND	--	Sterigmatocystin

aflatoxins and 13 samples for ochratoxin A, 2 for zearalenone, 5 for T-2 toxin and 3 for sterigmatocystin. This can be attributed for the usage of damaged feed ingredients by the layer farmers to reduce the feed cost.

When 35 chick feed samples were screened for their natural contamination of different mycotoxins 5, 2 and 1 sample were positive for aflatoxins, ochratoxin A and T-2 toxin respectively. Zearalenone and sterigmatocystin were not traced in the chick feed samples screened. Similarly, 35 developer feed samples screened for presence of mycotoxin, contamination 8 samples were positive for aflatoxins, 3 for ochratoxin A, 1 for zearalenone and 2 for T-2

toxin. Sterigmatocystin was absent in developer feed samples.

Screening of 35 grower feed samples revealed that 11 samples were positive for aflatoxins, similarly 3 for ochratoxin A, 1 for zearalenone and 1 for sterigmatocystin. T-2 toxin was not detected in grower feed samples screened. Out of of 35 prelay feed samples tested 9, 3 samples were positive for aflatoxins, T-2 toxin respectively. Interestingly, ochratoxin A, zearalenone and sterigmatocystin were not detected in the samples screened.

### 3.Feed ingredients

Feed ingredients are the key resources for contamination of feeds with mycotoxins. Hence, it was felt necessary to screen the different feed ingredients

used in Warangal district, and the results are shown in the Table 3. From the table it is evident that most of the feed ingredients screened were positive for different mycotoxins. Out of 50 samples of maize screened, 34 samples are positive for aflatoxins and 13 for ochratoxin A, 2 for zearalenone and 8 for T-2 toxin, and 9 for sterigmatocystin.

Out of 5 samples of sunflower meal screened for presence of mycotoxins, 22 samples were positive for aflatoxins, and 3 for ochratoxin A, 1 for zearalenone and 4 for T-2 toxin. No sample of sunflower meal was positive for sterigmatocystin. Similarly, out of 50 groundnut cake samples screened for presence of mycotoxins, 28, 13, 3, 2 samples were positive for aflatoxins, ochratoxin A, zearalenone, T-2 toxin respectively. Sterigmatocystin was not recorded in the samples screened.

Screening of 50 deoiled rice bran samples for their natural contamination of mycotoxins indicated that 14 samples were positive for aflatoxins and 8 for ochratoxin A, 6 for zearalenone, 4 for T-2 toxin and 1 for sterigmatocystin. Similarly, in 50 fish meal samples tested only 1 sample was positive for sterigmatocystin, 3 samples were positive for aflatoxins, ochratoxin A, zearalenone and T-2 toxins were not traced out in fishmeal sample. Similarly, when 50 soybean meal samples screened for presence of mycotoxins, 9 samples were positive for aflatoxins, and 6 for ochratoxin A, 5 for zearalenone and 4 for T-2 toxin. No sample was positive for sterigmatocystin contamination.

Studies on natural incidence of mycotoxins in poultry feeds revealed that aflatoxin followed by ochratoxin A are most common and contaminated large number of poultry feeds and feed ingredients. Most of the samples were contaminated with more than one mycotoxin. Farm mixed feed samples were recorded with highest frequencies of mycotoxin contamination than commercial brands. These findings are in line with the worldwide natural occurrence of different mycotoxins in poultry feeds and feed ingredients (Dalcero *et al.*, 2002; Labuda *et al.*, 2005; Oliveira *et al.*, 2006; Zinedine *et al.*, 2006; Rodrigues *et al.*, 2011; Rodrigues and Naehrer, 2012). Widespread occurrence of mycotoxins in all types of feeds and their significance in poultry industry calls for imposition of quality control regulations for commercial feeds or

home-brew methods for detection of mycotoxins in the feeds prepared at farm level.

**Conflicts of interest:** The authors stated that no conflicts of interest.

## REFERENCES

- Adey J and Mateles RC (1964) Incorporation of labeled compounds into aflatoxins. *Biochimica et Biophysica Acta* 86, 418-420.
- Dalcero A, Magnoli C, Hallak C, Chiacchiera S, Palacio G and Rosa CDR (2002) Detection of ochratoxin A in animal feeds and capacity to produce this mycotoxin by *Aspergillus* section *Nigri* in Argentina. *Food Additives and Contaminants* 19, 1065-1072.
- Devegowda G and Raju MVLN (2005) Manual on mycotoxin quantification in foods and feeds. Veterinary College, University of Agricultural Sciences, Karnataka University, India.
- Gimeno A (1979) Thin layer chromatographic determination of aflatoxins, ochratoxins, sterigmatocystin, zearalenone, citrinin, T-2 toxin, diacetoxyscirpenol, penicillic acid, patulin and penitrem A. *Journal-Association of Official Analytical Chemists* 62, 579-585.
- Gorst-Allman CP and Steyn PS (1979) Screening methods for the detection of thirteen common mycotoxins. *Journal of Chromatography A* 175, 325-331.
- Kamimura H, Nishijima M, Yasuda K, Saito K, Ibe A, Nagayama T, Oshiyama H and Naoi Y (1981) Simultaneous detection of fusarial toxins. *Journal-Association of Official Analytical Chemists* 64, 1067.
- Labuda R, Parich A, Vekiru E and Tancinova D (2005) Incidence of fumonisins, moniliformin and *Fusarium* species in poultry feed from Slovakia. *Annals of Agricultural and Environmental Medicine* 12, 81-86.
- Munkvold GP (2003) Cultural and genetic approaches to managing mycotoxins in maize. *Annual Review of Phytopathology* 41, 99-116.
- Oliveira GR, Ribeiro JM, Fraga ME, Cavaglieri LR, Direito GM, Keller K, Dalcero AM and Rosa CA (2006) Mycobiota in poultry feeds and natural occurrence of aflatoxins, fumonisins and zearalenone in the Rio de Janeiro State, Brazil. *Mycopathologia* 162, 355-362.
- Perrone G, Susca A, Cozzi G, Ehrlich K, Varga J, Frisvad JC, Meijer M, Noonim P, Mahakarnchanakul W and Samson RA (2007) Biodiversity of *Aspergillus* species in some important agricultural products. *Studies in Mycology* 59, 53-66.
- Ramakrishna Y and Bhat V (1987) Comparison of different spray reagents for identification of trichothecenes. *Curr. Sci.* 56, 524-526.
- Rice LG and Ross PF (1994) Methods for detection and quantitation of fumonisins in corn, cereal products and animal excreta. *Journal of Food Protection* 57, 536-540.
- Richard JL Cole RJ and Archibald SO (1989) Mycotoxins: economic and health risks. Council for Agricultural

Science and Technology, Task Force Report Ames Iowa 116, 1-99.

Rodrigues I and Naehrer K (2012) A three-year survey on the worldwide occurrence of mycotoxins in feedstuffs and feed. *Toxins* 4, 663-675.

Rodrigues I, Handl J and Binder E (2011) Mycotoxin occurrence in commodities, feeds and feed ingredients sourced in the Middle East and Africa. *Food Additives and Contaminants, Part B* 4, 168-179.

Stack ME and Pohland AE (1975) Collaborative study of a method for chemical confirmation of the identity of aflatoxins. *Journal of the Association of Official Analytical Chemists* 58, 110-113.

Trail F, Mahanti N and Linz J (1995) Molecular biology of aflatoxin biosynthesis. *Microbiology* 141, 755-765.

Zinedine A Brera C Elakhdari S Catano C, Debegnach F, Angelini S, de Santis B, Faid M, Benlemlih M, Minardi V and Miraglia M (2006) Natural occurrence of mycotoxins in cereals and spices commercialized in Morocco. *Food Control* 17, 868-874.

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