

Effect of chemical pre-treatments (HCL AND H₂SO₄) on seed germinability in *Jatropha Curcas* (L.)

Gaikwad Rajesh Shrirangrao

Department of Botany, Swami Vivekanand Senior College, Mantha- 431504. Dist. Jalna (Maharashtra) India. Email: <u>drrsgaikwad@gmail.com</u>.

Manuscript details:

Available online on http://www.ijlsci.in

ISSN: 2320-964X (Online) ISSN: 2320-7817 (Print)

Cite this article as:

Gaikwad Rajesh Shrirangrao (2020) Effect of chemical pre-treatments (HCL AND H₂SO₄) on seed germinability in *Jatropha Curcas* (L.), *Int. J. of. Life Sciences*, Special Issue, A14: 34-38.

Article published in Special issue of National e-Conference on Recent Aspects in Biosciences-2020" organized by Departmentt of Botany Rashtramata Indira Gandhi College, Jalna, Maharashtra, India date, June 29, 2020.

Copyright: C Author,

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, distribution sharing, adaptation, and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third-party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a vqoo of this license, visit http://creativecommons.org/ licenses/by/4.0/

ABSTRACT

Seed propagation studies were carried out on percent germination, shoot length and root length of *Jatropha curcas*. Seeds were treated with different chemical treatments and effect of seed age. Seeds of *J. curcas* treated with HCL, H₂SO₄ and hot water showed significantly more germination and shoot length. Similarly the chemical treatment H₂SO₄ (50%) at different time period worked significantly better than other treatments.

Key words: Seed Propagation, chemical treatment, Jatropha curcas.

INTRODUCTION

Jatropha curcas L., known as Physic nut, is a drought resistant perennial plant belonging to family Euphorbiaceae (Kaushik and Roy 2007). The plant grows well on poor soils. It has a long history of cultivation in tropical and subtropical regions of the world. Jatropha curcas is a succulent that sheds its leaves during the dry season and well adapted to arid and semi-arid condition and often used for erosion control. It is truly a multipurpose tree species fit for agro forestry and other afforestation programme (Kaushik et.al 2006). Extract from the plant is known for their effect on the wide range of organisms including mouse, nematodes and insect pests, (Kumar et.al 2007). Most of the species of Jatropha are capable of producing seeds with high oil content, but only few species of Jatropha particularly Jatropha curcas found highly suitable for large scale cultivation because of its high oil content about 30-40 % which can be combusted as fuel without being refined. Jatropha curcas is mainly cultivated for the production of oil (27 - 40 %) that has fuel substitute (Achten et al 2007). The seeds of J. curcas have highly potentials as a fuel substitution. However, the seeds in general, are toxic to human and animals. Curcin is a toxic protein isolated from the seeds and also contains a high concentration of phorbol esters (Adolf et al., 1984; Makkar et al., 1997). Two new esterases (JEA and JEB) and a lipase (JL) were isolated from the seeds (Staubmann et al., 1999). In Egypt, the seed is used for the treatment of gout jaundice and arthritis (Khafagy et al., 1977). The seed of J. curcas

has also been used traditionally for the treatment of many ailments including inflammation, burns and convulsions (Osoniyi and Onajobi, 2003). The seed oil can be applied to treat skin diseases and eczema and to sooth rheumatic pain (Heller, 1996).

The oil is also used externally for the treatment of dropsy, sciatica and paralysis (Mujamdar *et al.*, 2000). The 36% linoleic acid (C18:2) content in *J. curcas* kernel oil is used for skin care. The oil has a strong purgative action and also is commonly used for skin diseases and to sooth pain such as pain caused by rheumatism. In South Sudan, the seeds as well as the fruits are used as a contraceptive or as abortifacient (List and Horhammer, 1979).

The problem of great concern regarding this plant is a number of seeds produced per plant is very low. Besides, the plant produces seeds after 2-3 years approximately, depending on environmental conditions and seeds have a limited viability (50%) within 15 months (Palanivelu et.al 2005). *Jatropha curcas* oil is a strong alternative for diesel replacement (Reddy and Ramesh, 2005). Among various bio-diesel plants listed above the species of *Jatropha* in general and *Jatropha curcas*. in particular have become popular for the cultivation in the region of Maharashtra.

Looking to its question of low seed viability, to improve the seed germination, have need of healthy, quick growing of seedlings in short span of time from their sowing time. Keeping this in view, the aim of this study was to investigate the effects of chemical pre-treatments on germination behavior of the seed.

MATERIALS AND METHODS.

1) Collection of seed material

In the present study, seeds of the germplasm JS – 12 of *Jatropha curcas* were obtained from Jain Irrigation System Ltd., Jain Agri Park, Jain Hills Jalgaon (M.S).

2) Sowing of treated seeds

The seeds were sown during the months of February to March and September to October. The seeds were sown in polybags. The preferred polybags having size of 22.5 x 12.5 cm were used for sowing. Polybags were filled with mixed soil and well decomposed Farm Yard Manure in equal proportion in the ratio (1:1:1). The drainage holes were provided at the bottom of polybags. The treated as well as untreated seeds were sown in each polybags at a

depth of 1.5 to 2.0 cm (Kaushik and Kumar, 2003), (Paramathma *et al.*, 2004) and (Gurunathan, 2006).

RESULTS AND DISCUSSION

In order to understand the effect of seed age on the germinability. The seeds of *Jatropha curcas* of different age were used for germinability in pot soil and results are given in table 1.It is clear from table 1 that the seeds with age 1-6 month shows maximum germinability in case of *Jatropha carcus*. The seeds germinability was constant up to 12 month in *J. curcas* Similarly after 6 to 24 months there was reduction in percent germinability in *J. curcas*.

Table 1:	Effect o	f seed	age	on	seed	germinability	in
Jatropha	curcas .						

	Species of Jatropha						
Seed age	Jatropha curcas						
(month)	Percent	Shoot	Root				
	Germination	length	length				
		(cm)	(cm)				
1-6 Month	60	22.88	15.72				
12 Month	60	21.32	13.42				
24 Month	40	18.96	11.69				
S.D.	11.55	1.97	2.02				
S.E. ±	6.67	1.14	1.17				
C.D.(P=0.05)	17.13	2.93	3.00				

In order to study the effect of HCL, H_2SO_4 and Hot water on seed germinability, the seeds of *J. curcas* were treated with 50% of both acid concentration and hot water at ($50^{\circ}c \pm 2^{\circ}c$), percent germinability, shoot length and root length in cm were observed. The results are mentioned in table 2. It is clear from result summarized in table 2 that seeds of *J. curcas* treated with 50% HCL, 50% H₂SO₄ for four minutes were proved favourable to express maximum germinability, while same treatment found highly effective for shoot and root length. Seed treated with hot water for 6, 10, 15 and 20 minutes showed equally more or less effective.

It is also interesting to mentioned that as there was increase in time of treatment of HCL and H_2SO_4 , reduction in percent germinability was seen. As the time of treatment of HCL and H_2SO_4 was maintained about 6 minute there was reduction in present germinability in *J. curcas.*

It is clear from table 3 that the seeds of J. curcas treated by HCL showed 05 different types of fungi namely Alternaria alternata, Aspergillus flavus, A. fumigatus, Aspergillus niger, Fusarium moniliforme, F. oxysporum, Mucur spp, Rhizopus stolonifer. The seeds of J. curcas treated by H₂SO₄ showed 06 different types of fungi namely Alternaria alternata, A. fumigatus, Aspergillus niger, F. oxysporum, Mucur spp, Rhizopus stolonifer similarly the incidence on seeds treated by Hot water showed 06 different types of fungi namely Alternaria alternata, Aspergillus flavus, Aspergillus niger, F. oxysporum, Mucur spp, Rhizopus stolonifer. Gaikwad and borkar (2018) studied and concluded that the seeds treated by KOH showed 09 different types of fungi namely Alternaria alternata, Alternaria tenuis , Aspergillus niger, A. fumigatus, Aspergillus flavus, Fusarium moniliforme, F. oxysporum, Mucur spp, Penicillium spp,. Rhizopus stolonifer and seeds treated by NaOH showed 08 different types of fungi namely Alternaria alternate, Aspergillus niger, A. fumigatus,

Aspergillus flavus, F. oxysporum, Mucur spp, Penicillium spp., Rhizopus stolonifer. Gaikwad Rajesh (2019) concluded that the seeds treated by IAA+ IBA for 600 ppm, showed 07 different types of fungi namely Alternaria alternata, Aspergillus niger, Aspergillus flavus, A. fumigatus, F. oxysporum, Mucur spp, and Rhizopus stolonifer. Seeds treated by IAA+ NAA for 400 ppm showed 07 different types of fungi namely Alternaria alternate, Aspergillus niger, A. fumigatus, Aspergillus flavus, F. oxysporum, Mucur spp, Rhizopus stolonifer. It is clear from literature that during the course of association of micro-organisms with seeds either in the field during storage the microbes utilize and deteriorate seeds contents in many cases as reported in case of Arachis hypogea (Lalitha Kumari et.al., 1970). Similar to oil seeds degradation of medicinally important chemicals from the seeds of medicinal plants have been reported to be degraded by the seed mycoflora as reported by (Sinniah et al 1983)., in case of Strychnos nux vomica seeds.

Table 2: Effect of HCL, H₂SO₄, Hot water on seed germinability in *Jatropha curcas*

	Jatropha curcas								
Time (min)	HCL (50%)			H ₂ SO ₄ (50%)			Hot water (50°c ± 2)		
	Percent Germination	Shoot length (cm)	Root length (cm)	Percent Germination	Shoot length (cm)	Root length (cm)	Percent Germination	Shoot length (cm)	Root length (cm)
0.5	50	19.21	16.33	80	20.51	16.37			
2	100	27.55	17.07	100	26.38	17.34			
4	100	28.69	19.96	100	29.62	17.71	40	16.10	12.98
6	40	17.55	12.07	80	18.32	12.67	100	18.77	12.67
10	20	12.42	10.11	20	16.53	10.18	100	24.48	16.22
15							100	28.88	18.16
20							80	22.91	14.52
S.D.	42.37	11.73	8.05	45.77	11.77	8.37	46.19	11.58	7.51
S.E. ±	16.01	4.43	3.04	17.30	4.45	2.92	17.46	4.38	2.84
C.D. (P=0.05)	41.16	11.40	7.82	44.46	11.43	7.51	44.87	11.25	7.30

 Table 3: Incidence of fungi on infected seeds of Jatropha curcas

Fungi	Incidence on seeds treated by HCL	Incidence on seeds treated by H ₂ SO ₄	Incidence on seeds treated by Hot water
Alternaria alternata	+	+	+
Aspergillus flavus	+	-	+
Aspergillus fumigatus	+	+	-
Aspergillus niger	+	+	+
Fusarium moniliforme	-		
Fusarium oxysporum	-	+	+
Mucur spp	-	+	+
Rhizopus stolonifer	+	+	+

-- = Absent + = Present.

The information regarding the identification of seed mycoflora which was found to be responsible for spoilage of seeds is available in the literature (Jain *et.al.*, 1982) isolated Fusarium oxysporum from the seeds of mustard. (Heller 1992) isolated Julus spp. from total loss of seedlings of Jaropha curcas. Chavan and Danai (1993) isolated F. moniliforme, F. dimerum, F. roseum, F. semitectum from seeds of groundnut. Gaikwad 2019) studied that the different chemical treatments as (IAA+ IBA and IAA+ NAA) on seed germination and seedling growth of Syzygium cumuni. It was observed that IAA + IBA at 600 ppm favoured maximum seed germination. On the other hand shoot length and root length was maximum at IAA+ NAA at 400 ppm. (Paramathma et.al 2004) suggested that healthy and bold seeds sown at 1.5 to 2.0 cm depth showed maximum germination. While Swamy and Singh (2006) reported maximum percent germination in J. curcas seeds soaked 24 hours in cold water for 2-3 minute in boiling water. Gaikwad and Borkar (2018) Pre-sowing treatment of seed plays vital role to enhance the seed germination under nursery conditions. Among the pre-sowing treatments, the best treatment for the sowing of seeds is 10% KOH and 10% NaOH for 10 minutes favored maximum seed germination of Syzygium cumuni.

CONCLUSIONS

Pre-sowing treatment of seed plays significant role to enhance the seed germination under nursery conditions. Among the pre-sowing chemical treatments, seeds of *J. curcas* treated with HCL, H_2SO_4 and Hot water showed significantly more germination and shoot length .Therefore, pre-sowing treatments of seeds with HCL, H_2SO_4 and hot water may be recommended for plantation scheme.

Acknowledgements.

Author is grateful to the Head, Department of Botany, Dr. Babasaheb Ambedkar Marathwada University, Aurangabad (M.S) for providing necessary facilities and UGC-SRF (Rajiv Gandhi National Fellowship) for financial supports.

REFERENCES

Achten WMJ, Mathijs E, Verchot L, Singh VP, Aerts R and Muys B (2007) *Jatropha* biodiesel fueling sustainability? *Biofuel Bioproduction and Bioresource*. 1(4),283–291.

- Adolf W, Opferkuch HJ, Hecker E (1984). Irritant phorbol derivatives from four Jatropha species. *Phytochemistry*, 23: 129-132.
- Chavan and Danai (1993). Fungi occurring on discolored seeds of pulses and oil seeds. *Indian bot. Reptr.* 12(1+3): 87-90.
- Gaikwad RS and Borkar SU (2018). Effect of pre-sowing treatment on seeds germination and plant growth of syzygium cumini(L.) Skeels *.International Journal of Research Analytical Reviews* .5 (4):186-189
- Gaikwad RS (2019). Effect of chemical pre-treatments on seed germination and seedling growth of syzygium cumuni (l.) skeels. *Think India Journal ISSN: 0971-1260.2019, 22: 31*
- Gurunathan, N (2006). Seed management studies in Jatropha curcas M.Sc., (Forestry). Thesis submitted to Tamil Nadu Agricultural University, Coimbatore.
- Heller J (1992). Studies on genotypic characteristic and propagation an cultivation method for physic nuts (*Jatropha curcas* L.). Dr. Kovac, Hamburg.
- Heller J (1996). Physic Nut. Jatropha curcas L. Promoting the Conservation and use of Underutilized and Neglected Crops. International Plant Genetic Resources Institute, Rome
- Jain SK, Singh PN and Singhal G (1982). Fungi associated with seed of Brassica. *Seed research*. 10(1): 87-90.
- Kumar S, Kaushik N and Roy S (2007). Genetic variability and divergence studies in seed traits and oil content of Jatropha (*Jatropha curcas* L.) accessions. Biomass Bioenergy; 31:497–502.
- Kaushik N., Roy S and Biswas, GC (2006).Screening of Indian germplasm of *Jatropha curcas* for selection of high oil yielding plants. *Indian Journal of Agroforestry*. 8: 54–7.
- Kaushik N and Roy S (2007) Genetic variability and divergence studies in seed traits and oil content of Jatropha *(Jatropha curcas L.)* accessions. Biomass Bioenergy. 31:497–502.
- Kaushik and Kumar.(2003). Propagation and uses of *J. curcas* L. paper presented in smposium on emerging trends in research and business management of medicinal and aromatic plants. 11-13. March, CCSHAU, Hisar
- Khafagy SM, Mohamed YA, Abdel SNA, Mahmoud ZF (1977). Phytochemical study of *Jatropha curcas*. *Planta Med*. 31: 274-277.
- Lalitha kumari D, Govindswami CV and Vidhyasekaran P (1970). Isolated seed-borne fungi from stored groundnut seed and their role in seed spoilage. *Madras Agric. J.* 57(9): 27-28.
- List PH, Horhammer L (1979). Hager's Handbuch der Pharmazeutischen Praxis, Springer, Berlin
- Makkar HPS, Becker K, Sporer F, Wink M (1997). Studies on nutritive potential and toxic constituents of different provenances of *Jatropha curcas. J. Agric. Food Chem.* 45: 3152-3157.
- Mujamdar AM, Upadyhe AS, Misar AV (2000). Studies on antidiarrhoeal activity of *Jatropha curcas* root extract in albino mice. *J. Ethnopharmacol.* 70: 183-187.
- Osoniyi O, Onajobi F (2003). Coagulant and anticoagulant activities in *Jatropha curcas. J. Ethnopharmacol.* 89: 101-105. (Osaniyi Clarify).
- Palanivelu V, Vijayavel K, Ezhilarasibalasubramanian S and Balasubramanian MP (2005). Impact of fertilizer (urea) on oxygen consumption and feeding the fresh water fish

Oreochromis mossambicus. Environmental Toxicology and Pharmacology.19: 351–355.

- Paramathma M, Parthban KT and Neelakantan KS (2004). Cultivation and management of *Jatropha* plantation. In: strategies for improvement and utilization of tree borne oilseeds. Forestry series-3. TNAU, mettupalayan. 218.
- Reddy and Ramesh A (2005). Parametric studies for improving the performance of a *Jatropha* oil-fuelled compression ignition engine. Renewable energy.31:1994-2016.
- Reddy and Ramesh (2005). Parametric studies for improving the performance of a *Jatropha* oil-fuelled compression ignition engine. Renewable energy.31: 1994-2016.
- Sinniah DG, Varghese G, Baskaran and Koo SH (1983). Fungal flora of *Strychnos nux vomica* seeds and neem oil toxicity. Malays. *Appl. Biol.* 12(1): 1-4.
- Swamy SL and Lalji Singh (2006). Strategies for Development of quality planting stock of *Jatropha curcas* for Biodiesel plantations. Biodiesel Conference Towards Energy Independence Focus on *Jatropha. Hyderabad.* 143-157.
- Staubmann R, Manfred S, Alois H, Theodor K (1999). A complex of 5- hydroxypyrrolidin-2-one and pyrimidine-2,4-dione isolated from *Jatropha curcas*. *Phytochemistry*. 50:337-338.

© 2018 | Published by IJLSCI

Submit your manuscript to a IJLSCI journal and benefit from:

- ✓ Convenient online submission
- ✓ Rigorous peer review
- Immediate publication on acceptance
- ✓ Open access: articles freely available online
- High visibility within the field

Email your next manuscript to IRJSE : editorirjse@gmail.com