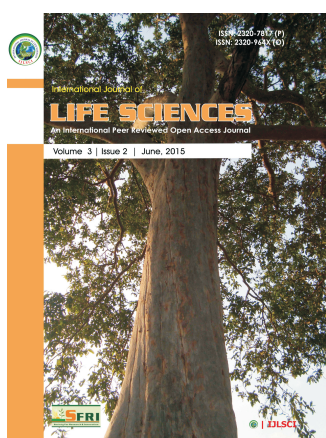


ISSN: 2320-7817 | e-ISSN: 2320-964X
Volume 3, Issue 2, June 2015

INTERNATIONAL JOURNAL OF
LIFE SCIENCES

AN INTERNATIONAL PEER REVIEWED OPEN ACCESS JOURNAL



Editor in Chief
Dr. Santosh S. Pawar

Managing Editor
Dr. Arvind Chavhan

STATUTORY WARNING

Articles, data, figures, scientific contents and its interpretation and authenticity reported by author(s) and published in IJLSCI are the exclusive views of author(s). The editorial board, IJLSCI is not responsible for any controversies arising out of them. In case of any plagiarism found, author(s) will have to face its consequences.

All enquiries and manuscript should be directed to

The Managing editor IJLSCI
46, Guruvandan, Jawahar Nagar,
VMV Road, Amravati -444604,
(MS) India

Contact: +91-942 077 5527
+91-997 055 9438
Email: editor@ijlsci.in
editorijlsci@gmail.com
web:www.ijlsci.in



All the materials in this journal is copyright and may not be reproduced except with the return permission of the publisher
© Copyright International Journal of Life Sciences



Cover image: *Terminalia arjuna* (Roxb.) Wt & Arn.

Description: The Arjuna is about 12–25 m tall; Branches glabrous; Smooth bark greenish white; Leaves sub-opposite, oblong or elliptic-oblong, rounded or sub cordate at base, Obtuse or sub-acute, shallowly crenate-serrate or subentire, glabrous, pale green above, brownish beneath; Petiole 6-10 mm long with 1-2 prominent glands at base of lamina; Flowers sessile, in short axillary and terminal panicles; Bractioles linear lanciolate, caducous. Calyx 4 mm long glabrous; teeth triangular, 1.5 mm long. Disc clothed with yellowish or redish hairs. Drupes 3-5 cm long, ovoid-oblong, dark brown, with 5 hard wings (Naik & Associates, 1998).

Locality: Near Sipna River bank, Semadoh Rest House

Distribution: Very Common through Maharashtra state along banks of rivers, Mahabaleshwar, Khandala, Pachgani, Satara, Western Ghats, South India, Gujarat, Lingmala.

Photo By: Mr. Anand Oak, Research Scholar, Dept of Botany, Shri Shivaji Science College, Amravati

Contents

RESEARCH ARTICLES

- 125 In vitro fungitoxic effect of some plant growth regulators on spore germination and germ tube emergence of *Alternaria solani*
Bhajibhuje MN
- 131 Effect of Domestic Sewage on Phytoplankton Community in River Rapti at Gorakhpur
Kushwaha VB | Agrahari M
- 141 Impact of Heavy metal, Arsenic trioxide on Biochemical profile of teleost, *Clarias batrachus* (Linn.)
Pundir Garima | Pundir Himanshu
- 147 Food and feeding of an economically important estuarine fish, *Sillago sihama* (forsskal)
Yeragi SS | Yeragi SG
- 152 Antioxidant and antimicrobial properties of *Adhatoda vasica* L. Nees
Wankhede TB
- 157 Seasonal Variation of Physicochemical and Microbial Parameters of water of Nal-Damayanti Sagar Dam, Morshi, Dist. Amravati, MS, India
Ghaware AU | Jadhao RG
- 162 Effect of moisture content on the production of protease by *Fusarium oxysporum* using agroindustrial waste
Vidhale NN | Deshmukh Rupali R
- 167 Rotifers diversity in Kudla Dam near Umri Nanded, MS, India
Bhojar VV
- 171 Diversity of Zooplankton in some Reservoirs in and around Karwar- Uttara Kannada District Karnataka
Vasanthkumar B and Kapsikar Gangadhar B
- 176 NF1 gene Analysis: New paradigm by computational approach
Jadhav VA | Laeequr Raheman

SHORT COMMUNICATION

- 181 Unusual sighting of Yellow-wattled Lapwing (*Vanellus malabaricus*) in Lucknow District, Uttar Pradesh, India
Kumar Adesh | Kanaujia Amita
- 185 Index

Editorial Board

Editor in Chief

Dr. Santosh Pawar
Department of Biology,
Institute of Forensic Sciences, Nagpur

Managing Editor

Dr. Arvind Chavhan
Department of Zoology,
D.B. College, Bhokar, Nanded

Associate Editor

Dr. Bela Volgyi
Department of Experimental Zoology
and Neurobiology, Pécs, Ifjúságu,
Hungary

Dr. Fazal Shirazi,
Anderson Cancer Center,
1515 Holcombe Blvd, Houston, Texas.

Dr. Gunjan Mukherjee, Ph.D.,
Department of Biotechnology and
Bioresources, (TERI)
Indian Habitat Centre (IHC), New Delhi

Dr. Suresh B. Zade
Professor in Zoology,
RTM University, Nagpur, MS, India

Dr. Satish A. Bhalerao
Professor in Botany,
Department of Botany,
Wilson College, Mumbai, India

Dr. M.N. Bhajbhuj
Department of Botany,
Jawaharlal Nehru College,
Wadi, Nagpur, India

Dr. B.N. Pandey
P.G. Department of Zoology,
Purnea College, Purnia, Bihar, India

Dr. Kamble SY,
Dy. Director, Botanical Survey of India,
Pune.

Dr. Mali Rajendra Prabhakar
Department of Zoology,
Yeshwant Mahavidyalaya,
Nanded (MS) India

Dr. Gyan Singh Shekhawat
Department of Botany,
Jai Narain Vyas University,
Jodhpur, India

Dr. Mumtaz Baig
P.G. Department of Zoology, G.V.I.Sc.,
Amravati. 444604(MS) India

Associate Editor

Dr. K. Madhava Chetty,
Plant Taxonomist, Department of
Botany, SVUCS, S.V. University,
Tirupati-517 502, AP, India.

Dr. Gurpreet Singh Dhillon
Institut national de la Recherche
Scientifique (INRS), Centre for Water,
Earth and Environment;
University of Québec, CANADA

Dr. Bankefa Emmanuel Olufemi
Department of Microbiology,
Federal University of Technology,
Akure, Ondo-State. Nigeria.

Dr. Zohair I.F. Rahemp
Department of Biology, University of
Salahaddin, Erbil, Kurdistan, Iraq

Dr. Kishor G. Patil,
P.G. Department of Zoology,
Institute of Science, Nagpur, India

Dr. Rajusing G. Jadhao
Department of Zoology, Shri Shivaji
Science College, Amravati, India

Dr. Mukund M. Dhore
Department of Botany,
B.B. Arts, N.B. Comm. & B.P. Science
College, Digras, MS India.

Dr. S.S. Nanware
Department of Zoology,
Yashawant College, Nanded, India

Dr. Vishal Marathe
Department of Botany, NES, Science
Collge, Nanded, India

Dr. Sangita Ingole
Department of Environmental
Science, Shri Shivaji Science
College, Amravati (MS) India

Dr. Sushant Punekar
Govt. Autonomous P.G. College,
Chhindwara (M.P.) India

Dr. Rajendra V. Tijare,
Department of Zoology,
Institute of Science, Nagpur, India

Dr. Uma Rani Agrawal
Department of Zoology,
CMP Degree College,
University of Allahabad, Allahabad,
India

Member of Editorial Board

Dr. Dhanraj V. Tayade
Department of Zoology
G.S. Gawande College,
Umardhed, MS India

Dr. Gayathri Narayan
Department of Zoology,
D.G. Ruparel College of Arts,
Science & Commerce,
Mahim (W), Mumbai 400 016.

Dr. Kumud Choudhary
Department of Zoology,
Shri R. R. Lahoti Science College,
Morshi, Amravati (MS) India.

Dr. Piyush Gupta
Department of Environmental
Chemistry, Science and Humanities
SRM university, Modinagar,
Ghaziabad, india

Dr. Tushar Wankhede
Depratmnet of Botany,
Shri Shivaji Science College,
Amravati, India

Dr. Sachin Tawade
Department of Botany,
D.B. College, Bhokar, Nanded

Prof. Aruna U. Kakade
Department of Environment,
Shri Shivaji Science College,
Amravati, India

Dr. Dinesh K. Dabhadkar
Department of Zoology,
G. S. Gawande College, Umardhed,
India.

About IJLSCI

International Journal of Life Sciences (IJLSCI) is a peer-reviewed, open access journal; it publishes original articles in all areas of Biology and Life Sciences. Authors are encouraged to submit complete unpublished and original works, which are not under review in any other journals. The scopes of the journal include, but limited to the following topic areas: Biology, Anatomy, Botany, Cytology, Genetics, Microbiology, Physiology, Zoology, Ecology, Environmental Science, Hydrobiology, Neurobiology, Developmental Biology, Immunology, Molecular Biology, Biochemistry, Biophysics, and Biotechnology, Genomics and Proteomics. It is an open access journal, which publishes original research articles, reviews and short communication in all areas of life sciences and at the interface of related disciplines.

TYPES OF RESEARCH PAPER

Original Articles:

These should describe new and carefully confirmed findings, and experimental methods should be given in sufficient detail for others to verify the work. The length of an original article is the minimum required to describe and interpret the work clearly.

Short Communications:

Short communication also contain abstract. The list of references should not exceed 15. The presentation of Short Communications should contain headings such as Introduction, Materials and Methods, Results and Discussion, etc.

Reviews Articles:

Submissions of review articles and perspectives covering topics of current interest are welcome and encouraged. Publication Frequency: Quarterly (published in March, June, September & December).

The journal is published in print as well as online version which is free access and downloads.

This Journal is circulated / Distributed Through Subscription only

Submission of Paper: Submission of paper is encouraged by e-mail to editorijlsci@gmail.com
Subscription Plan: Rs. 2500 /- or 150 US\$ (Individual Annual); Rs. 3500/- or 250 US\$ (Institutional)
Life Membership: Rs. 10000/- or 550 US\$ (Individual) Rs. 15000/- or 850 US\$ (Institutional)
Fellow Membership: Rs. 15000/- or 850 US\$ (Individual) Rs. 25000/- or 1700 US\$ (Institutional)

Publisher and Owner: Dr. Arvind Chavhan, Published from IJLSCI, 46, Guruvandan, Jawahar Nagar, VMV Road Amravati -444604, India

Printed at IJLSCI, 46, Guruvandan, Jawahar Nagar, VMV Road Amravati -444604, India

Managing Editor: Dr. Arvind Chavhan, Assistant Professor & Head, Department of Zoology, D.B. College, Bhokar, Nanded, India

**Publisher, Editorial Board and Editor in Chief take no responsibilities for the inaccurate, misleading data, opinion and statement appeared in the articles published in this journal.
All responsibilities of the contents rest upon the authors (especially upon the corresponding author).**

Indexing:

INNOSPACE (SJIF), ROAD, OAJI, Academic Keys, J-index.net, Scholar Steer, Research Bible, Google Scholar, DAIJ, DRIJ, DIIF, SIS, ISI, ASI.

**No Part of the this Journal can be reproduced without written permission of the publisher ©
Copyright 2013-15 By Dr. Arvind Chavhan, Managing Editor, International Journal of Life
Sciences, Amravati- 444604. India**

All Articles are freely accessible on the website of the International Journal of Life Sciences <http://www.ijlsci.in> As well as some of the Indexing agencies website via., oaji.net, research bible, Cite factors, ISI, etc.

RESEARCH ARTICLE

In vitro* fungitoxic effect of some plant growth regulators on spore germination and germ tube emergence of *Alternaria solani

Bhajbhuj MN

Seed Pathology Lab., Department of Botany, Jawaharlal Nehru Mahavidyalaya, Wadi, Nagpur- 440 023 (M.S.) India. | e-mail : dr_mnbhajbhuj@rediffmail.com

Manuscript details:	ABSTRACT
<p>Received: 29.04.2015 Accepted: 29.05.2015 Published : 30.06.2015</p> <p>Editor: Dr. Arvind Chavhan</p> <p>Cite this article as: Bhajbhuj MN (2015) <i>In vitro</i> fungitoxic effect of some plant growth regulators on spore germination and germ tube emergence of <i>Alternaria solani</i> . <i>Int. J. of Life Sciences</i>, 3(2): 125-130.</p> <p>Copyright: © 2015 Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.</p>	<p>The inhibitory effect of five plant growth regulators including indol-3-acetic acid; indol-3-butyric acid; naphthalene acetic acid; 2,4-dichlorophenoxy acetic acid; and phenyl acetic acid was evaluated <i>in vitro</i> against the leaf blight pathogen, <i>Alternaria solani</i>. All test chemical inducers at 10^{-2} to 10^{-4}M conc. significantly reduced spore germination and germ tube growth of the test pathogen. Pathogenic fungus, <i>Alternaria solani</i> had sensitive response against 10^{-2} conc. of naphthalene acetic acid, reducing spore germination by 84% and germ tube growth by 14% over untreated control. Also, complete inhibition spore germination of test fungal pathogen was confined when all chemical inducers were evaluated at 10^{-1}M conc. Fungal spore germination and germ tube growth increased significantly as the conc. of chemical inducers was decreased. Among the test growth regulators, indol-3-acetic acid had least inhibitory effect against test pathogen of these parameters undertaken. The results of the present study revealed the possibility of usage plant growth regulators in inducing phytoalexin compound in susceptible plant cultivars.</p> <p>Key words: Fungitoxicity, susceptible, abiotic elicitors, chemical plant resistance inducers.</p> <p>INTRODUCTION</p> <p>The Deuteromycetes ubiquitous fungal genus <i>Alternaria</i> comprises diverse saprophytic as well as Endophytic species and is known for its notoriously destructive plant pathogen members (Mamgain <i>et al.</i>, 2013). Out of the total 299 known species representing genus <i>Alternaria</i>, majority of them lack sexuality, although few species have been found to have sexual stage in</p>

their life cycle. The genus is characterized by the formation of polymorphous, multicellular and pigmented conidia either singly or in short or longer chains by asexual method. The conidia are broadest near the base; taper gradually to an elongated bead and provided with cross, longitudinal as well as oblique septa. It has been found to have a drastic effect on the members belonging in the plant families such as Cucurbitaceae, Brassicaceae, Solanaceae, Poaceae etc. which are having nutritional as well as economical food value. It is associated with a wide variety of substrates including seeds, plants, agricultural products, animals, soil and the atmosphere. It is also common allergen in humans, growing indoors and causing hay fever or hypersensitivity reactions that sometimes lead to asthma. Some species readily cause opportunistic infections in immuno-compromised people such as AIDS patients (Mamgain *et al.*, 2013). Several taxa are also important postharvest pathogens, causative agents of phaeohyphomycosis in immuno-compromised patients or airborne allergens.

Some saprophytic species representing genus *Alternaria*, are agents of decay and decomposition and growing profusely on dead and decaying debris of plant and animal origin producing a variety of primary and secondary metabolites. Several pathogenic species mostly causing leaf blight infection considered the major problems in agricultural production throughout the world, reducing yield and quality of crops and produce more than 70 phytotoxins of host selective (host specific) and nonspecific types (Trivedi *et al.*, 2013). Host-selective toxins (HSTs) are toxic only to host plants while nonspecific toxins can affect variety of non-host plants. The *Alternaria* HSTs involve a diverse group of low-molecular-weight substances such as alterotoxins, alternariol, tenuazonic acid; alternaric acid and were found in culture filtrates as families of closely related compound and were reported to play a crucial role in determining host specificity and contributing to disease development. The *Alternaria* HSTs cause necrosis on leaves of

susceptible cultivars at concentrations as low as 10^{-8} to 10^{-9} M and no necrosis on leaves of resistant cultivars even at higher concentrations (Otani *et al.*, 1995) The toxin from secondary metabolites penetrate host tissues, directly act on living host cell protoplasm and damage the metabolically active cells to influence the course of disease development (Mamgain *et al.*, 2013).

Majority species of *Alternaria* caused leaf spot and early blight resulting in defoliation, reduction in size and quality of fruits, ultimately adversely affects productivity (Mamgain *et al.*, 2013). The leaf spots may be control by foliar application of effective organic fungicides, but these are reported hazardous and their residual toxicity in plant parts, fruits poses carcinogenic disorders to consumers (Trivedi, *et al.*, 2013) and also helps to increase level of air pollutants. To overcome these, the concept of screening for disease resistance has been developed (Eckadt, 2011). Phytoalexin accumulation at infection site in leaves, stem, cotyledons and hypocotyl (Ingham, 1982), in response to wounding (Rahe and Arnold, 1975), to interaction with micro-organism (Iriti & Franco, 2009) or to treatment with certain chemicals (Ismile *et al.*, 1987; Bhajbhujje, 2013) makes a significant contribution to resistance, cultivars that are normally susceptible to a virulent race of pathogen thus providing protection in different plants. A little is known about induction of resistance by application of plant growth regulators in plants and control of leaf blight pathogen, it seemed to be worthwhile to report the fungitoxicity of plant growth regulators at variable concentrations against leaf blight pathogen, *Alternaria solani* (Ellis & Martin) Jones & Grout.

MATERIALS AND METHODS

The plant growth regulators of diverse chemical nature induce phytoalexin in plants, when applied in dilute concentration, were screened for fungitoxic effect against leaf blight causing pathogen *Alternaria solani*. The stock solution of

0.1M concentration for five plant growth regulators was prepared separately in volumetric flask and each was diluted to the concentration between 10^{-2} to 10^{-4} M. These solutions of different concentrations of test chemicals were screened for fungitoxic assay employing the slide germination technique (CMI, 2010). *Alternaria solani* was isolated from infested leaves and stored seeds of tomato as internal seed borne pathogen and maintained in laboratory on PDA nutrient medium at $25 \pm 1^{\circ}\text{C}$. One drop of different conc. of test chemicals was placed on cover slip and added one drop of spore suspension of test pathogen in the drop. The spores were allowed to grow in drop of water serve as control. The coverslip with spores in drop was inversely placed on cavity glass slide in triplicate. Slides of different treatments were randomly distributed into large Petri dishes made into moist chamber and kept these for 24 hrs. in darkness. One drop of lecto-phenol was put on each spot to fix the germinated spores. Germination of spores was counted in terms of percentage on the basis of 300 spores and germ tube growth was measured on the basis of 90 germlings from each spot observed randomly. These concentrations were selected on the basis of their effectiveness in inducing resistance in plants (Bhajbhujje, 2014).

RESULTS AND DISCUSSIONS

Altogether five plant growth regulators in aqueous dilute solution (10^{-2} to 10^{-4} M) are screened to study *in vitro* fungitoxic effect on spore germination & germ tube growth of pathogen following slide germination method (CMI, 2010). A drop of spore suspension in Czapek's Dox broth was placed 3 cm apart on each of three slides per treatment. The slides were randomly distributed into large Petri plates made into moist chamber and kept at room temperature in darkness. After 24 h of incubation, the percent spore germination was recorded from each spot on the basis of 50 spores and germ tube growth on the basis of 15 germlings.

The results presented in Table 1 revealed that an aqueous solution of all five plant growth regulators at different concentrations caused injury to spore of *Alternaria solani* inhibiting spore germination and germ tube growth. An absolute inhibitory effect was induced with naphthalene acetic acid (NAA) and phenyl acetic acid (PAA), when treated with 10^{-1} M aqueous solution of test chemical inducers. Naphthalene acetic acid (NAA) caused greater inhibitory effect at 10^{-2} M conc. reducing the spore germination by 84% over the untreated control, followed by phenyl acetic acid (PAA) causing reduction in this parameters by 71 %. Moderate inhibitory effect to the extent of 61% and 65% for spore germination was recorded with 2, 4-dichlorophenoxy acetic acid (2, 4-D) and Indol-3-butyric acid (IBA) at 10^{-2} M conc. respectively. Indol-3-acetic acid (IAA) had least inhibitory effect at this conc. on spore germination (Table 1).

The inhibitory effect for all the plant growth regulators was declined with dilution of aqueous stock solution. The greatest declining of inhibitory effect was recorded with naphthalene acetic acid (NAA), reducing the spore germination by 35% and 6%, when treated with conc. 10^{-3} to 10^{-4} M. Least inhibition of spore germination was confined at 10^{-4} M with Indol-3-acetic acid (IAA) while remaining test chemical inducers indol-3-butyric acid (IBA); 2,4-dichlorophenoxy acetic acid (2,4- D); phenyl acetic acid (PAA) had considerable to moderate inhibitory effect at conc. 10^{-3} M while it was declined to 2-9% when treated with 10^{-3} M conc. over untreated control (Table 1).

The similar trend was confined for germ tube emergence with all concentration of the plant growth regulators tested. The spores of leaf blight pathogen; *Alternaria solani* remained dormant and did not produce germ tube, when treated with 10^{-1} M aqueous solution of all five plant growth regulators excepting indol-3-acetic acid (IAA). All the test chemical inducers excluding indol-3-acetic acid (IAA) at 10^{-2} M concentration had 10 to 18% inhibitory effect on

germ tube growth compared to untreated control. Indol-3-acetic acid (IAA) at $10^{-2}M$ induced 3% inhibitory effect. Phenyl acetic acid (PAA) and 2,4-dichlorophenoxy acetic acid (2,4-D) caused greater inhibition at $10^{-2}M$ conc., reducing the mean germ tube growth to the extent of 18 and 17% respectively while remaining test chemical inducers had 10-14% inhibitory effect on the same parameter over untreated control (Table 1).

The inhibitory effect was declined with decrease in concentration of all the plant growth regulators tested. The greatest declining of inhibitory effect was recorded with Phenyl acetic acid (PAA) reducing the germ tube growth by 16% and 13% at conc. 10^{-3} to $10^{-4}M$ respectively. Indol-3-acetic acid (IAA) had little inhibitory effect; Naphthalene acetic acid (NAA) induced 9% inhibition while remaining test chemical inducers

Table 1: Effect of plant growth regulators at dilute concentration on spore germination and germ tube growth of *Alternata solani*

S. No.	Plant growth regulators	Percent spore germination			Mean germ tube growth		
		$10^{-2} M$	$10^{-3} M$	$10^{-4} M$	$10^{-2} M$	$10^{-3} M$	$10^{-4} M$
1.	Indol-3-acetic acid (IAA)	44 (-55.1)	71 (-27.6)	96 (-2.0)	92 (-3.1)	94 (-2.1)	95 (-1.0)
2.	Indol-3-butyric acid (IBA)	34 (-65.3)	64 (-34.7)	92 (-6.1)	86 (-10.4)	89 (-7.3)	91 (-5.2)
3.	Naphthalene acetic acid (NAA)	16 (-83.7)	48 (-51.0)	84 (-14.3)	83 (-13.5)	86 (-10.4)	87 (-9.4)
4.	2,4-dichlorophenoxy acetic acid (2,4-D)	38 (-61.2)	68 (-30.6)	94 (-4.1)	88 (-16.7)	92 (-4.2)	94 (-2.1)
5.	Phenyl acetic acid (PAA)	28 (-71.4)	56 (-42.9)	89 (-9.2)	79 (-17.7)	81 (-15.6)	84 (-12.5)
	Water (Control)	98	98	98	96	96	96

1. Results have been expressed as percentage in terms of control; 2.Average of 300 spores; 3.Average of 90 germlings; 4.Values in parentheses indicate percentage reduction or increase in terms of control

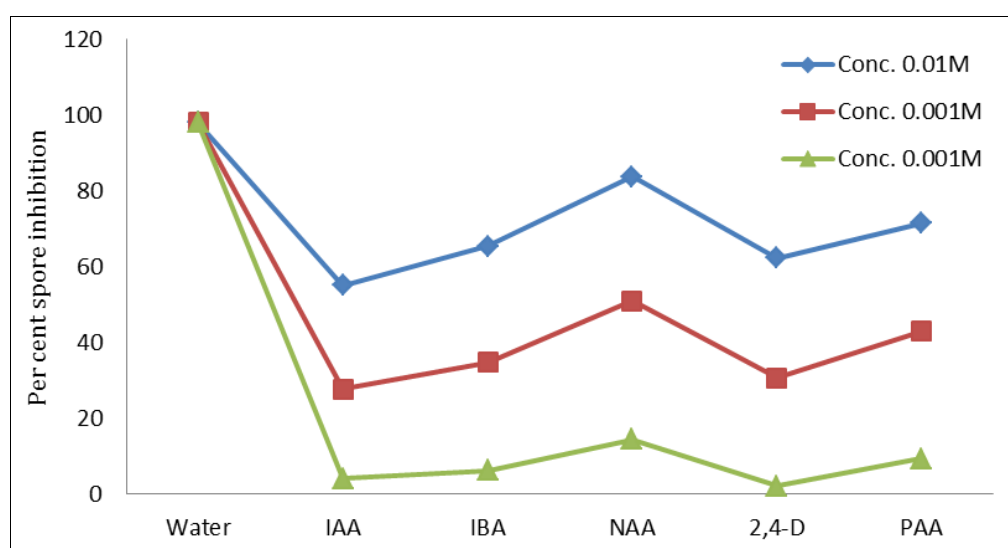


Fig. 1: Fungitoxicity of plant growth regulators on spore germination *Alternata solani* (IAA= indol -3-acetic acid; IBA=Indol-3-butarcic acid; NAA= Maphthalene acetic acid; 2,4-D+ 2,4-dichlorophenoxy acetic acid; PAA-phenyl acetic acid)

Indol-3-butyric acid (IBA); 2,4-dichlorophenoxy acetic acid (2,4-D); phenyl acetic acid had mild inhibitory effect at conc. 10^{-4} M compared to untreated control (Table 1). Of the five plant growth regulators screened against *Alternaria solani*, naphthalene acetic acid (NAA) caused greater inhibition of spore germination, but had moderate inhibitory effect on germ tube emergence, while Phenyl acetic acid (PAA) induced considerable inhibitory effect on spore germination but had greater reduction in germ tube growth. Moderate to considerable inhibitory effect was recorded with Indol-3-butyric acid (IBA) and 2,4-dichlorophenoxy acetic acid (2,4-D) while Indole-3-acetic acid had least inhibitory effect on both parameters studied (Fig. 1). It is in agreement with earlier finding of Ashraf and Ali (2007) who reported inhibitory response of chemical inducers on microbial community.

Ezzouhri *et al.*, (2009) reported the chemical tolerance level of some filamentous fungal organisms including *Alternaria alternata*, *Aspergillus niger*, *Geotrichum candidus*, *Penicillium sp.*, and *Fusarium sp.* Swami and Alane (2013) screened crude extract of various parts of some botanicals containing variable concentration of chemicals against some dominant seed borne fungal pathogens of green gram and reported the inhibitory level of the test crude extracts against these pathogens including *Alternaria alternata*, *Phytophthora sp.*, *Fusarium oxysporum*, *Aspergillus niger*, *Rhizoctonia solani*, *Curvularia lunata* and *Cladosporium* and reported the inhibitory level of the test crude extracts against these pathogens. The effectiveness of variable concentration of diverse group of chemicals was confirmed on spores of *Alternaria brassicicola* (Meena *et al.*, 2011), *Alternaria porae* (Feofilova *et al.*, 2012) and *Alternaria alternata* (Bhajibhuje, 2014).

Direct toxicity of heavy metal salts of varying origin to the fungal pathogen does not seem to explain the reduction of symptoms. Chlorides of copper and barium are non-toxic, provided stronger protection than mercuric and cadmium

chloride, a highly toxic one. These test chemicals may exert inhibitory influence upon fungal spores germination and impose upon them exogenous dormancy. This is clearly shown by sensitivity of fungal spores to chemicals by several researchers. The inhibition of spore germination may be attributed to variable toxic effect of test chemicals. Similar findings were reported with conidia of *Alternaria tenuis* (Bhajibhuje, 1989); *A. tenuissima* (Singh *et al.*, 2000); *A. alternata* (Meena *et al.*, 2011; Bhajibhuje, 2014), *A. porae* (Feofilova *et al.*, 2012), *A. solani* (Abdel-Kader *et al.*, 2012). The hydrolytic products of the chemicals possibly at low conc. induced dormancy or may cause injuries to fungal spores by dissolving the protective thick wall layers and plasma membrane or ruptured them making porous. Aqueous solution of test chemicals diffused through ruptured cell wall and porous plasma membrane to cytoplasm, react with functional cytoplasmic components of spore and seems to disturb a series of physiological processes of spore germination leading to any of the change (i) an inhibitors of trehalose degrading enzymes is destroyed; (ii) the trehalose degrading enzyme is synthesized from its precursor, the conversion being analogous to the trypinogen-trypsin transformation; (iii) the enzyme is thought to be spatially separated from its substrate inside a dormant spores and activation may bring the two together and (iv) a series of interlocking enzyme reactions are shifted from one steady state level (Feofilova *et al.*, 2012). In the present investigations, the variable inhibition of fungal spore germination and germ tube growth may be attributed to the differential toxic effect of the test chemicals.

CONCLUSION

The experimental findings reveal that an aqueous solution at 10^{-2} M concentration of plant growth regulators seemed to provide more vigorous defence response to virulent pathogen, *Alternaria solani*. These phytoalexin inducer test chemicals stimulated production of large amount of

fungitoxic substances in susceptible tissue on post-infection of virulent pathogen which make plant resistant to some extent and readily respond to infection. Of the test chemical inducers, naphthalene acetic acid at $10^{-2}M$ may serve as very promising compounds for use in plant disease control.

REFERENCES

- Abdel-Kader MM, El-Mougy NS, El-Gannal NG, Abd-El-Kareem F and Abd-Alla (2012) Laboratory evaluation of some chemicals affecting pathogenic fungal growth. *J. Appl. Sci. Res.*, 8(1) : 523-530.
- Ashraf A, Ali TA (2007) Effect of heavy metals on soil microbial community and mung beans seed germination. *Pak. J. Bot.*, 39(2): 629-636.
- Bhajibhuje MN (1989) Investigations on mycoflora associated with vegetable seeds from Vidarbha Region. Ph.D. Thesis, R.T.M. Nagpur University, Nagpur, M.S. India.
- Bhajibhuje MN (2013) Role of heavy metal salts on susceptibility of *Solanum melongena* L. seedlings to *Alternaria* early blight disease. *Int. J. of Life Sci.*, 1 (1) : 51-62.
- Bhajibhuje MN (2014) Response of heavy metal salts against *Alternaria* leaf spot infection on *Vigna mungo* (L.) Hepper seedlings by three techniques. *Int. J. of Life Sci.*, 2(2): 1-13.
- CMI (2010) Commonwealth Mycological Institute. Description of Pathogenic fungi and bacteria. Kew Surrey, England. Pp 451-460.
- Eckadt NA (2011) Induction of Phytoalexin Biosynthesis: WRKY₃₃ - Is a Target of MAPK Singling. *Plant Cell*, 23(4) : 1190.
- Ezzouhri L, Castro E, Moya M, Espinola F, Lairini K (2009) Heavy metal tolerance of filamentous fungi isolated from polluted sites in Tangier, Morocco. *Afri J. Microbiol. Res.*, 3(2) : 35-48.
- Feofilova E, Ivashechkin A, Alekhin A, Sergemma Y (2012) Fungal spores: Dormancy, germination, chemical composition & role in biotechnology (review), *Appl Biochem, & Microbiol.*, 48(1):1-21.
- Ingham JL (1982) Phytoalexins from the Leguminosae. In J. A. Bailey and J.W. Mansfield (Eds) Phytoalexins. Blackie and Son, Glasgow London, pp 289-318.
- Iriti M and Franco F (2009) Chemical Diversity and Defence Metabolism: How Plant Cope with pathogen and ozone pollution. *Int. J. Mol. Sci.*, 10 : 337-339
- Ismile IMK, Salama AAM, Ali MIA, Oaf SAE (1987) Effect of some phenolic compounds on spore germination and germ tube growth of *Asergillus fumigatus* and *Fusarium oxysporum* f. sp *lycopersici*. *Cryptogamie Mycology*, 8(1) : 51-60.
- Mangain A, Roychoudhary and Jagatpati Tah (2013) *Alternaria* pathogenicity and its strategic controls. *Res. j. of Biology*, 1 : 1-9
- Meena PD, Chattopadhyay C, Kumar A, Awasthi RP, Singh R, Kaur S, Thomas L, Goyal P, Chand P (2011) Comparative study on effect of chemicals on *Alternaria* blight in Indian mustard – A multilocation study in India. *J. Environ. Biol.*, 32(3) : 375.
- Otani H, Kohomoto K and Kodawa M (1995) *Alternaria* toxins and their effects on host plants. *Can. J. Bot.*, 73 (Suppl) : S543-S548.
- Rahe JE and Arnold RM (1975) Injury related phaseollin accumulation in *Phaseolus vulgaris* and its implications with regard to specificity of host-parasite interaction. *Can. J. Bot.*, 53: 921-927.
- Singh SK, Singh UP, Tuli L, Prithviraj B, Sarma BK (2000) . Effect of spore conc, of *Alternaria tenuissima* on germination & development of germ tubes on hosts & non-hosts. *Indian Phytopath.*, 53(4) : 419-422.
- Swami CS, Alne SK (2013) Efficacy of some botanicals against seed-borne fungi of green gram (*Phaseolus aureus* Roxb) . *Bioscience Discovery*, 4(1) : 107-110.
- Trivedi A, Sharma SK, Hussain T, Sharma SK, Gupta PK (2013) Application of biodynamic preparation, bio-control agent and botanicals for organic management of virus and leaf spots of black gram (*Vigna mungo* L. Hepper). *Acad. J. Agric. Res.*, 1(4): 60-64.

RESEARCH ARTICLE

Effect of Domestic Sewage on Phytoplankton Community in River Rapti at Gorakhpur

Kushwaha VB and Agrahari M

Department of Zoology, D.D.U. Gorakhpur University, Gorakhpur-273008, U.P., India
E mail: vbkddugkp@gmail.com

Manuscript details:

Received: 09.05.2015
Revised : 04.06.2015
Revised Received: 08.06.2015
Accepted: 12.06.2015
Published : 30.06.2015

Editor: Dr. Arvind Chavhan

Cite this article as:

Kushwaha VB and Agrahari M (2015) Effect of Domestic Sewage on Phytoplankton Community in River Rapti at Gorakhpur. *Int. J. of Life Sciences*, 3(2): 131-140.

Copyright: © 2015 | Author(s),

This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

ABSTRACT

This paper aims to study the effect of domestic sewage on phytoplankton population in river Rapti at Gorakhpur, U.P. and India. An increase in free CO₂, bicarbonate alkalinity, nitrate, phosphate and BOD while a decrease in pH, DO and carbonate alkalinity was observed at sewage mixing point. However, these parameters gradually changed at the station away from sewage mixing point (downstream) and were within the limits of Indian standards. During the study period total 29 species of phytoplanktons were observed belonging to 4 families: Bacillariophyceae (11), Chlorophyceae (10), Cyanophyceae (6) and Euglenophyceae (2). In present investigation it was observed that population of phytoplankton was very low at station R₂ where sewage mixed into river.

Keywords: Sewage, River, Phytoplanktons, Bacillariophyceae, Chlorophyceae, Cyanophyceae, Euglenophyceae

INTRODUCTION

Water is indispensable for all living organisms on the earth any of the water on this planet is stored in ocean and ice caps, which is difficult to recover for our diverse needs. Any of our demands for water are fulfilled by rain water, which gets deposited in surface and ground water resources. Now days, both surface and ground water resources are contaminated by various sources like industrial effluents, agricultural discharge and municipal waste water associated with large amount of inorganic and organic toxic pollutants along with harmful pathogens (Okoh et al., 2007). Sewage, used water of community generally contains organic as well as inorganic wastes from residences, business houses and industries that might lead to the. It affects physical,

chemical and biological characteristics of the water. Hence, continuous monitoring of river's water quality is very essential to determine the state of pollution in our rivers. Communication of this information to general public and government can help to develop the policies for the conservation of our natural fresh water resources (Ali et al., 2000). Phytoplanktons are a predominant type of a plant found in aquatic bodies. The quality and quantity of phytoplankton is good indicator of water quality. Phytoplanktons (algae) are those microscopic plants contain chlorophyll-a that float or swim too feebly to maintain a constant position against a water current (Lee 2008). Many workers have studied the plankton diversity and effect of domestic sewage on it, (Palmer 1980; Arya et al., 1987; Acharjee et al., 1995; Jha et al., 1997; Mathivanan et al., 2007; Hassan et al., 2008). The objective of this work is to show the effect of domestic sewage on phytoplankton population in river Rapti a tributary of river Ghaghara at Gorakhpur, U.P., India.

MATERIALS AND METHODS

Sampling stations:

Three water sampling stations were selected over 5 kms stretch of river Rapti. Station I (R₁): upstream of sewage discharge, station II (R₂): sewage mixing point, here a continuous discharge of city sewage through a large cemented drain occurs on the bank of the river. About 3 kms away from the station I, station III (R₃): downstream of sewage discharge point, about 2 kms away from station II (Agrahari and Kushwaha, 2012).

Sample collection and their physico-chemical analysis:

The study of the river Rapti at Gorakhpur was done for 12 months (Dec 2009 to Nov 2010). Samples of river water were collected in winter (Dec 2009) and summer (Jun 2010) season from all three stations. Samples were collected in plastic bottles for physico-chemical analysis. For

biochemical oxygen demand (BOD) and dissolved oxygen (DO) samples were collected in BOD bottles. Temperature, pH and dissolved oxygen were measured at the site (Agrahari and Kushwaha, 2012).

Methods for analysis of physico-chemical parameters:

The physico-chemical analysis was carried out using the methods given by APHA (1976). pH was measured using pH meter and temperature was measured using simple, mercury filled Celsius thermometer. Nitrate, phosphate and sulphate test kits were purchased from Hi-media Laboratories Pvt. Ltd., Mumbai, India (Agrahari and Kushwaha, 2012).

Collection and Preservation of phytoplanktons:

For the analysis of phytoplankton samples were collected by filtering 30 liters of water through standard plankton counting net. Phytoplanktons were counted with the help of Sedwicz Rafter slide. The phytoplankton samples were observed under Olympus microscope. The phytoplanktons were identified by using books and journals viz. (Agrahari and Kushwaha, 2014; APHA 1985; Jena et al., 2005; Pingale and Deshmukh, 2005; Hosmani, 2008; Perumal and Anand, 2008).

Calculation:

Abundance of phytoplankton was estimated as organisms/liter in the concentrated sample using the equation:

$$N \text{ (number of individuals)/Liter} = \frac{A \times 1000 \times C}{L}$$

Where,

A = Number of phytoplanktons/cc,

C = Volume of concentrated sample taken for counting

L = Volume of water in liter for collection of sample

1000 = Area of counting chambers

Data was subjected to Analysis of variance using software.

RESULTS AND DISCUSSIONS

The average physico-chemical quality of river Rapti at three sampling stations in Gorakhpur during summer, winter and rainy seasons are presented in Table 1.

The maximum water temperature 33.8 °C was observed at station R₂ in summer season and minimum 19.9 °C at station R₃ in winter season. The pH value of the river water at different stations was recorded to be within highest 8.5 at station R₁ in summer season and lowest 7.0 at station R₂ in winter season. Highest value of electrical conductivity 404µmhos/cm was recorded at station R₂ in summer season and lowest value 198µmhos/cm was recorded at station R₁ in rainy season and highest value of TDS 286 ppm was recorded at station R₂ in rainy season and lowest value 102 ppm was recorded at station R₁ in winter season. Dissolved O₂ ranged between 10.2 ppm at station R₁ during winter to 2.4 ppm at station R₂ summer season. Low level of DO is again indicative of polluted nature of water body. Free CO₂ ranged between 20 ppm at station R₂ in summer season and 2 ppm at station R₁ and R₃ in winter season. Chloride concentration ranged between 12 ppm at station R₁ in rainy season to 285 ppm at station R₂ in summer season. The carbonate alkalinity varied from and 24.4 ppm at station R₁ in summer season and 3.5 ppm at station R₂ in rainy season. Bicarbonate alkalinity varied from 81 ppm at station R₃ in rainy season to 762 ppm at station R₂ in summer season. Total hardness, Ca hardness and Mg Hardness varied from a maximum of 492 ppm at station R₂ in summer season and 122 ppm at station R₁ in rainy season, 216 ppm at station R₂ in summer season and 44.6 ppm at station R₃ in rainy season and 62.28 ppm at station R₂ to 11.9 ppm at station R₁ in rainy season respectively. Nitrate was ranged between 0.019 ppm at station R₁ to 1.8 ppm at station R₂ in summer season. Nitrate is one of the most important indicators of pollution of water. Phosphate was found to be maximum of 1.6 ppm at station R₂ in summer season and 0.016 ppm at

station R₃ in winter season. Sulphate concentration ranged between maximum of 18.5 ppm at station R₂ in summer season to a minimum of 10.4 ppm at station R₃ in rainy season. The BOD of river water varied from a maximum of 109.4ppm at station R₂ in summer season and 2.2 ppm at station R₃ in winter season. COD values ranged between 51 ppm at station R₂ in winter season to 19.2 ppm at station R₁ in rainy season.

Mean values of phytoplankton (units/ml) of river Rapti at three sampling stations (R₁, R₂ and R₃) in summer, winter and rainy seasons are presented in table 2. 29 species of phytoplanktons were observed belonging to 4 families: Bacillariophyceae (11), Chlorophyceae (10), Cyanophyceae (6) and Euglenophyceae (2).

Bacillariophyceae:

In present study the largest and diverse group is Bacillariophyceae. Diatoms were represented by 11 species *Amphora sp.*, *Navicula sp.*, *Fragilaria sp.*, *Nitzschia sp.*, *Gomphonema sp.*, *Pinnularia sp.*, *Syndra sp.*, *Gyrosigma sp.*, *Surirella sp.*, *Diatoma sp.* and *Melosira sp.*. In the present study maximum number of species of Bacillariophyceae 1983 units/ml in summer season was recorded at station R₁. Minimum number of species 533.1 units/ml in rainy season was recorded at station R₂ where sewage mixed into the river.

Chlorophyceae:

Chlorophyceae was the second group after Bacillariophyceae in the number of identified species observed. In this group 10 species were recorded which are *Chlorella sp.*, *Scenedesmus sp.*, *Zygnema sp.*, *Volvox sp.*, *Ankistrodesmus sp.*, *Ulothrix sp.*, *Cosmarium sp.*, *Mougeotia sp.*, *Pediastrum sp.* and *Spirogyra sp.* In the present study maximum number of species of Chlorophyceae 1616.5 units/ml in summer season was recorded at station R₁. Minimum number of species 241.6 units/ml in rainy season was recorded at station R₂.

Table 1: Mean values of physical and chemical parameters at different sampling stations of the river Rapti at Gorakhpur during summer, winter and rainy seasons.

Tests	Stations									Between Stations	Between Seasons
	Summer			Winter			Rainy				
	R _{S1}	R _{S2}	R _{S3}	R _{S1}	R _{S2}	R _{S3}	R _{S1}	R _{S2}	R _{S3}		
Temp (°C)	33.500	33.800	33.200	20.000	20.000	19.900	27.500	27.700	26.500	*(p<0.05)	** (p<0.01)
pH	8.500	7.400	8.200	8.000	7.000	8.200	8.200	7.200	7.500	*(p<0.01)	
Elec Cond, (µmhos/cm)	220.000	404.000	309.000	201.000	264.000	224.000	198.000	245.000	203.000	*(p<0.05)	*(p<0.05)
TDS (ppm)	122.000	260.000	182.000	102.000	207.000	164.000	140.000	286.000	185.000	*(p<0.01)	*(p<0.01)
DO ₂ (ppm)	7.600	2.400	6.900	10.200	3.600	8.200	8.800	3.600	8.000	** (p<0.01)	*(p<0.01)
Free CO ₂ (ppm)	8.000	20.000	6.000	2.000	6.000	2.000	4.000	18.000	5.400	*(p<0.01)	*(p<0.05)
Cl ₂ (ppm)	35.000	285.000	46.000	20.000	270.000	48.000	12.000	109.000	20.200	*(p<0.01)	
CO ₃ (ppm)	24.400	8.000	14.000	14.400	4.000	8.400	6.800	3.500	4.000	*(p<0.01)	*(p<0.01)
HCO ₃ (ppm)	372.000	762.000	608.000	208.000	660.000	301.000	92.000	115.000	81.000	*(p<0.05)	*(p<0.01)
Tot Hard (ppm)	200.000	492.000	268.000	188.000	424.000	220.000	122.000	450.000	198.000	** (p<0.01)	*(p<0.01)
Ca Hard (ppm)	65.400	216.800	68.600	49.700	95.600	69.600	54.500	65.800	44.600		
Mg Hard (ppm)	24.160	54.380	39.640	21.880	62.280	24.000	11.900	51.100	25.760	*(p<0.01)	
NO ₃ (ppm)	0.038	1.800	0.190	0.019	0.072	0.038	0.049	0.480	0.071	*(p<0.01)	
PO ₄ (ppm)	0.060	1.600	0.400	0.020	0.380	0.016	0.040	1.540	0.600	*(p<0.01)	
SO ₄ (ppm)	11.600	18.400	12.600	14.800	18.500	14.500	11.900	12.900	10.400	*(p<0.01)	*(p<0.01)
BOD (ppm)	6.800	109.400	5.900	3.200	69.400	2.200	4.000	70.400	5.000	*(p<0.01)	
COD (ppm)	23.200	46.500	32.500	22.000	51.000	39.800	19.200	49.600	30.800	*(p<0.01)	

** (p<0.01) indicates highly significant differences

* (p<0.01) and *(p<0.05) indicates significant differences

Table 2: Mean values of phytoplanktons (units/ml) at different sampling stations in the river Rapti at Gorakhpur during summer, winter and rainy seasons.

Tests	Stations									Between Stations	Between Seasons	
	Summer			Winter			Rainy					
	Rs1	Rs2	Rs3	Rs1	Rs2	Rs3	Rs1	Rs2	Rs3			
Bacillariophyceae	<i>Amphora sp.</i>	183.30	125.00	75.00	241.60	166.60	100.00	183.30	58.30	150.00	*(p<0.05)	
	<i>Navicula sp.</i>	266.60	166.60	166.60	233.30	200.00	66.60	225.00	100.00	83.30	*(p<0.01)	
	<i>Fragilaria sp.</i>	316.60	283.30	283.30	250.00	100.00	175.00	66.60	83.30	83.30		*(p<0.01)
	<i>Nitzschia sp.</i>	183.30	58.30	66.60	100.00	-	-	58.30	66.60	50.00	*(p<0.05)	
	<i>Gomphonema sp.</i>	208.30	175.00	141.60	200.00	166.60	100.00	100.00	66.60	-	*(p<0.01)	** (p<0.01)
	<i>Pinnularia sp.</i>	175.00	66.60	91.60	266.60	83.30	100.00	100.00	58.30	-	*(p<0.01)	*(p<0.05)
	<i>Synedra sp.</i>	225.00	200.00	166.60	-	-	-	100.00	-	83.30		*(p<0.01)
	<i>Gyrosigma sp.</i>	125.00	75.00	83.30	-	-	-	141.60	100.00	100.00	*(p<0.05)	*(p<0.01)
	<i>Surirella sp.</i>	-	-	-	-	-	-	100.00	-	66.60		*(p<0.01)
	<i>Diatoma sp.</i>	133.30	66.60	75.00	125.00	-	100.00	-	-	-		*(p<0.05)
	<i>Melosira sp.</i>	166.60	-	100.00	-	-	-	-	-	-		*(p<0.05)
Total	1983.00	1216.40	1249.60	1416.50	716.50	641.60	1074.80	533.10	616.50			
Chlorophyceae	<i>Chlorella sp.</i>	175.00	-	66.60	-	-	-	-	-	-		
	<i>Scenedesmus sp.</i>	133.30	50.00	175.00	150.00	-	200.00	150.00	66.60	91.60	*(p<0.01)	
	<i>Zygnema sp.</i>	275.00	83.30	141.60	116.60	-	58.30	-	-	-		*(p<0.01)
	<i>Volvox sp.</i>	133.30	66.60	91.60	-	-	-	150.00	50.00	83.30	*(p<0.05)	*(p<0.01)
	<i>Ankistrodesmus sp.</i>	300.00	166.60	225.00	250.00	75.00	66.60	-	-	-		*(p<0.01)
	<i>Ulothrix sp.</i>	100.00	58.30	75.00	200.00	75.00	91.60	150.00	50.00	100.00	*(p<0.01)	
	<i>Cosmarium sp.</i>	133.30	50.00	166.60	66.60	75.00	100.00	-	-	-		*(p<0.01)
	<i>Mougeotia sp.</i>	200.00	66.60	100.00	-	-	-	141.60	75.00	83.30		*(p<0.01)
	<i>Pediastrum sp.</i>	-	-	-	166.60	50.00	100.00	-	-	-		*(p<0.01)
	<i>Spirogyra sp.</i>	166.60	58.30	200.00	200.00	-	75.00	-	-	-		*(p<0.05)
Total	1616.50	599.70	1241.40	1149.80	275.00	691.50	591.60	241.60	358.20			

Table 2: Continued...

Tests		Stations									Between Stations	Between Seasons
		Summer			Winter			Rainy				
		R _{S1}	R _{S2}	R _{S3}	R _{S1}	R _{S2}	R _{S3}	R _{S1}	R _{S2}	R _{S3}		
Cyanophyceae	<i>Microcystis sp.</i>	291.60	300.00	100.00	133.30	166.60	150.00	-	-	-		*(p<0.01)
	<i>Oscillatoria sp.</i>	200.00	266.60	100.00	166.60	166.60	75.00	100.00	125.00	100.00	*(p<0.05)	
	<i>Anabaena sp.</i>	200.00	200.00	200.00	166.60	175.00	50.00	100.00	133.30	91.60		*(p<0.05)
	<i>Merismopedia sp.</i>	100.00	200.00	100.00	-	-	-	-	-	-		*(p<0.01)
	<i>Spirulina sp.</i>	100.00	100.00	66.60	-	-	-	-	-	-		*(p<0.01)
	<i>Nostoc sp.</i>	166.60	266.60	183.30	75.00	158.30	66.60	-	-	-	*(p<0.05)	*(p<0.01)
	Total	1058.20	1333.20	749.90	541.50	666.50	341.60	200.00	258.30	191.60		
Euglenophyceae												
	<i>Euglena sp.</i>	200.00	75.00	66.60	100.00	-	66.60	-	-	-		*(p<0.05)
	<i>Phacus sp.</i>	100.00	100.00	175.00	150.00	-	100.00	-	-	-		*(p<0.05)
	Total	300.00	175.00	241.60	250.00	-	166.60	-	-	-		

** (p<0.01) indicates highly significant differences

* (p<0.01) and * (p<0.05) indicates significant differences

Cyanophyceae:

Cyanophyceae was represented by 6 species of which *Microcystis sp.*, *Oscillatoria sp.*, *Anabaena sp.*, *Merismopedia sp.*, *Spirulina sp.* and *Nostoc sp.* In the present study maximum number of species of Cyanophyceae 1333.2 units/ml in summer season was recorded at station R₂. Minimum number of species 191.6 units/ml in rainy season was recorded at station R₃.

Euglenophyceae:

Only two species of Euglenophyceae were recorded that is *Euglena sp.* and *Phacus sp.* In the present study maximum number of species of Euglenophyceae 300 units/ml in summer season was recorded at station R₁. Minimum number of species 166.6 units/ml in summer season was recorded at station R₃. Group Euglenophyceae was absent in rainy season.

DISCUSSION

In present investigation an increase in electrical conductivity, TDS, free CO₂, bicarbonate alkalinity, total, Ca and Mg hardness, chloride, nitrate, phosphate, sulphate, BOD and COD while a decrease in pH, DO₂ and carbonate alkalinity was observed at sewage mixing point. However, these parameters gradually changed at the station away from sewage mixing point (downstream). A highest value of electrical conductivity was recorded at station R₂ in summer season. This might be due to the addition of sewage into it. An increase in electrical conductivity is regarded as pollution indicator in water bodies (Das et al., 2006; Agarhari and Kushwaha, 2012). An increase in TDS at station where sewage meets river water indicates an increase in pollution. Water with high dissolved solid is of inferior quality and may induce adverse response in the body of the consumer (Agarhari and Kushwaha, 2012; Mahor, 2011). Low level of DO is again indicative of polluted nature of water body. Such low level of oxygen was also noted by Iqbal et al.

(2006). At station R₂ saturation level of dissolved O₂ was very low in winter and rainy season. It may be due to high rate of oxygen consumption by oxidizable matter coming in along with sewage. Free CO₂ present in large amount at station R₂ can be attributed to high BOD load that comes with consumption of oxygen and release of CO₂ by the respiratory activity of the living organisms. Maximum values of free CO₂ recorded at station R₂ during summer might be due to acceleration in the rate of decomposition of organic matter by microbes, decrease of photosynthetic activity and high rate of respiration by benthic biota and microorganisms as observed by Hedge and Bharti (1985) and Sinha (1988). Maximum values of bicarbonates alkalinity recorded at station R₂ and R₃ were probably due to the input of domestic sewage. Shah (1988) noticed higher concentration of bicarbonate alkalinity in the domestic sewage during the study of river Jhelum. High fluctuation of Ca, Mg and total hardness were recorded at station R₂. In the present study higher values of Ca, Mg and total hardness observed at all the three sampling stations (Table 1 and 2) may be due to input of domestic sewage which contains organic matters. Cl₂ was found to be highly marked, higher values of Cl₂ recorded at station R₂ was due to the continuous influx of contaminated domestic sewage. Similar results are reported by Sinha (1988) in the case of river Yamuna and river Damodar. The maximum values of phosphate observed at station R₂, in comparison to other stations throughout the study period may be due to the discharge of contaminated domestic sewage containing decayed organic matter (Shah, 1988; Rana and Palria, 1988). Higher values were recorded at station R₂, owing to high amount of organic matter in domestic sewage (Paramshivam and Sreenivasan (1981) and Somashekar (1985) also reported that an increase in BOD and bacterial level as indicative of increasing pollution, which is supported by Sinha (1988). Station wise, maximum values of COD were recorded at station R₂ indicating presence of organic wastes in sewage.

In present investigation it was observed that population of phytoplankton was low at station R₂ where sewage mixed into river. This lowering of phytoplankton population at station R₂ is due to the presence of organic and inorganic matters of sewage that affects the physico-chemical quality of water as evidenced earlier by many workers (Kang et al., 2004; Shirodkar et al., 2010; Bhardwaj et al., 2010). And physico-chemical properties show effect on the phytoplankton diversity. Many previous studies showed the effect of physico-chemical parameters on plankton community. Hassan *et al.*, (2008) studied the effect of chemical and physical properties of River water in Shatt-AI-Hilla on phytoplankton communities, Sukumaran and Das, (2002) reported that the basic process of phytoplankton production was dependent upon temperature, turbidity and nutrients. The role of temperature in the development of algae has been emphasized by many workers from time to time (Palharya et al., 1993). Limnological studies of water quality are based on the principle that every type of aquatic ecosystem is associated with one specific community of organisms, the living communities that develop in aquatic ecosystems depend on specific physico-chemical characteristics of water and are noticeably modified when those conditions change (Kushwaha and Agrahari, 2014). Factors such as dissolved oxygen, transparency, depth, salinity, pH, temperature and nutrients influence the occurrence, abundance and distribution of Planktonic organisms (MBO, 2007) and effect their composition and distribution from place to place and year to year due to the dynamic nature of the aquatic system (FAO, 2006). In present observation temperature shows a moderate value in summer season which is favorable for growth of phytoplankton. Nutrients (nitrates and phosphates) showed low values in both seasons where as it increases at station-R₂ due to presence of domestic sewage. Dissolved Oxygen decreases at station R₂ that causes lowering of phytoplankton population. Presence of Dissolved oxygen is essential to maintain the biological life in the water Palharya, et al., 1993). In this

observation it was found that group Cyanophyceae showed higher population at station R₂ in comparison to other station because member of Cyanophyceae group are known to be highly adaptive and can colonize even the polluted area (Palharya, et al., 1993). Present observation also showed seasonal variations in phytoplankton communities, phytoplankton productivity was high during summer and low during winter season as evidenced earlier by Sadguru et al., (2002), Sharma et al., (2011) and Agrahari and Kushwaha (2012). The lowering of population of phytoplankton in winter can be attributed to low temperature (Sadguru et al., 2002; Hassan et al., 2008; Gross and Pfiester, 1988).

REFERENCES

- APHA (1976) Standard methods for the examination of water and waste water (14th Ed.) American Public Health Association, New York.
- Acharjee B, Dutta A, Choudhury M and Pathak B (1995) Phytoplankton species diversity indices in Dighali beel, Assam, India. *Environ. Ecol*, 13(3): 660-662.
- Agrahari M and Kushwaha VB (2012) Effect of domestic sewage on the physico-chemical quality of river Rapti at Gorakhpur. *Bioscan*, 7(1): 135-138.
- Ali M, Salam A, Azeem, A, Shafique M, Khan BA, (2000) Studies on the effect of seasonal variation on physical and chemical characteristics of mixed water from river Ravi and Chenab at union site in Pakistan. *J. Res. B. Z. Univ. Multan*, 2: 1-7.
- American Public Health Association (APHA) (1985) Standard methods for the examination of water and waste water, 15th Edition APHA, American Water Works Association, Water Pollution Control Federation, Washington D.C.

- Arya SC, Mudgal S and Shrivastava P (1987) Effect of sewage and industrial waste on river ecosystem. *Ind. J. Limnol.*, 15(1): 49-56.
- Bhardwaj V, Singh DS and Singh AK (2010) Water quality of the Chhoti Gandak river using principle component analysis, Ganga Plan, India. *Ind. J. Earth. Syst. Sci.*, 119(2): 117-127.
- Das R, Samal NR, Ray PK and Mitra D (2006) Role of electrical conductivity as an indicator of pollution in shallow lakes. *Asian J. Water. Env. Pollu.*, 3(1): 143-146.
- Food and Agriculture Organization (FAO) (2006) Interrelationship between fish and plankton in inland water. Retrieved Sept.29, 2007, from <http://www.fao.org/DOCREP/006/X7580E03.htm-106k-coached>.
- Gross JL and Pfiester LA (1988) Blue- Green Algae of Lake Thunderbird. *Proc. Oklahoma Academy Science*, 68: 39-44.
- Hassan FM, Kathim NF and Hussein FH (2008) Effect of chemical and physical properties of river water in Shatt Al-Hilla on phytoplankton communities. *E-Journal of Chemistry*. 5(2): 322-330.
- Hedge CR and Bharti, SG (1985) Comparative phytoplankton ecology of fresh water ponds and lakes of Dharwad, Karnataka State, India. *Proc. Nat. Symp. Pure Appl. Limnology* (Ed) Adoni A.D., Bull. Bot. Soc. Sagar., 32: 24-29.
- Hosmani P (2008) Ecology of Euglenaceae from Dharwar, Karnataka, *Indian Hydrobiology*, 11(2): 303-312.
- Iqbal PJ, Pandit AK and Javeed JA (2006) Impact of sewage waste from settlements on physico-chemical characteristics of Dal Lake, Kashmir. *J. Res. Dev.*, 6: 81-85.
- Jena M, Ratha SK and Adhikary SP (2005) Algal diversity changes in Kathajodi river after receiving sewage of Cuttak and its ecological implications. *Indian Hydrobiology*, 8(1): 67-74.
- Jha AK, Latif A and Singh JP (1997) River pollution in India: An overview. *J. Environ. Pollution*, 4(2): 143-151.
- Kang S, Su X, Tong L, Shi P, Yang X, Abe Y, Du T, Shen Q and Zhang J (2004) The impacts of human activities on the water-land environment of the Shiyang River Basin, an arid region in Northwest China. *Hydrological Sciences Journal des Sciences Hydrologiques*, 49(3): 413-427.
- Kushwaha VB and Agrahari M (2014) Effect of domestic sewage on zooplankton community in River Rapti at Gorakhpur, India, *World Journal Zoology*, 9(2): 86-92.
- Lee RE (2008) Phycology, Fourth edition, Cambridge University Press, Cambridge, ISBN 13 978-0-511-38669-5.
- Mahor RK (2011) Limnological study of fresh water reservoir Tighra, Gwalior (M.P.). *Int. Referred Research Journal*, 1(17): 47-48.
- Mathivanan V, Vijayan P, Sabhanayakam S and Jeyachitra O (2007) An assessment of plankton population of Cauvery River with reference to pollution. *Journal of Environmental Biology*, 28(2 Suppl): 523-526.
- MBO (2007) Zooplankton Retrieved Sept. 29, (2006) from <http://www.marine.bio.com/oceans/zooplankton.asp-62k>. ("Zooplankton - MarineBio.org". MarineBio Conservation Society. Web. Retrieved again on 2/7/2015. [http://marinebio.org/oceans/zooplankton/.](http://marinebio.org/oceans/zooplankton/))
- Okoh AI, Odjadjare, E.E. Igbinosa, E.O. and Osode, A.N. (2007) Wastewater treatment plants as a source of microbial pathogens in the receiving watershed, *Afr. J. Biotech.*, 6(25): 2932-2944.
- Palhrya JP, Siriah VK and Malvia S (1993) Environmental impact of sewage and effluent disposal on the river system. Ashish Publishing House, New Delhi, 1-66.
- Palmar CM (1980) Algae and water pollution. Castle House Publication Ltd. England, 123pp:123

- Paramshivam M and Sreenivasan A (1981) Changes in algae flora due to pollution in Cauvery Rivers. *Indian J. Environ Health* 23(3): 222-238.
- Perumal MG and Anand N (2008) Diversity of Desmids (Zygnamatales, Chlorophyceae) from Tiruchirappalli district of Tamil Nadu. *Indian Hydrobiology*, 11(2): 261-270.
- Pingale SD and Deshmukh BS (2005) Some freshwater algae from Amphitheatre of Wilson Dam. *Indian Hydrobiology*, 7(Supp): 97-100.
- Rana BC and Palria S (1988) Assessment, evaluation and abatement studies of a polluted river. The Bandi (Rajasthan). Ecology and Pollution of Indian Rivers, Ed. Trivedi R.K., Ashish Pub. House, New Delhi, 344-359.
- Sadguru P, Khalid K and Ansari K (2002) Seasonal dynamics of zooplankton in a fresh water pond developed from the waste land of brick kiln. *Pollut. Res.*, 21(1): 81-83.
- Shah AR (1988) Physico-chemical aspects of pollution in River Jhelum (Kashmir) during 1981-83. Ecology and Pollution of Indian River, Ed. Trivedy R.K., Ashish Pub. House, New Delhi, 163-207.
- Sharma I, Dhanze R, Kondal A and Dhiman R (2011) Seasonal abundances and diversity of Plankton of Banner stream, River Beas, Himachal Pradesh. *J. Env. Bio-Sci.*, 25(2): 279-284.
- Shirodkar PV, Pradhan UK, Fernandes D, Haldankar SR and Rao GS (2010) Influence of anthropogenic activities on the existing environmental conditions of Kandla Creek (Gulf of Kutch). *Curr.Sci.*, 98(6): 815-828.
- Sinha MP (1988) Effect of waste disposal on water quality of river Damodar in Bihar. Physico-chemical characteristics. Ecology and Pollution of Indian Rivers, Ed. Trivedy R.K., Ashish Publication House. New Delhi, 219-246.
- Somashekar RK (1985) Studies on water pollution of river Cauvery, India. *Proc. Nat. Sym. Pure and Appl. Limnology* (ed.) Adoni, A.D. Bull. Bot. SOC. Sagar. 32: 145-149.
- Sukumaran PK and Das AK (2002) Plankton abundance in relation to physico-chemical features in a peninsular manmade lake. *Environ. Ecol.*, 20(4): 873-879.

RESEARCH ARTICLE

Impact of Heavy metal, Arsenic trioxide on Biochemical profile of teleost, *Clarias batrachus* (Linn.)

Pundir Garima^{1*} and Pundir Himanshu²

¹Department of Zoology , R. G. (P.G) College, Meerut UP, India

²Department of Computer Science and Technology, Radha Govind Engineering College Meerut UP, India,

Address for correspondence Email: gaarimaa112@yahoo.co.in

Manuscript details:	ABSTRACT
<p>Received: 03.03.2015 Revised : 13.04.2015 Accepted: 03.06.2015 Published : 30.06.2015</p> <p>Editor: Dr. Arvind Chavhan</p> <p>Cite this article as: Pundir Garima and Pundir Himanshu (2015) Impact of Heavy metal, Arsenic trioxide on Biochemical profile of teleost, <i>Clarias batrachus</i> (Linn.). <i>Int. J. of Life Sciences</i>, 3(2): 141-146.</p> <p>Copyright: © 2015 Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.</p>	<p>The present investigation aims at evaluating the toxic effect of heavy metal, Arsenic trioxide on biochemical profile of <i>Clarias batrachus</i> after 30, 45 and 60 days of post treatment with experimental chemical , arsenic trioxide. Lc 50 value calculated for sublethal study of arsenic trioxide was 8.7 mg/l. Following biochemical parameters were analysed: Serum Protein, Serum Cholesterol. Serum Glucose, Acid phosphatase and Alkaline phosphatase. Decrease in serum protein observed in present study was due to liver cirrhosis or nephrosis. Increase in cholesterol indicated environmental stress in <i>Clarias batrachus</i> .The significant reduction in plasma glucose levels during acute treatment indicates hypoxic condition. Increased stimulation of alkaline phosphatase corresponds to pathological processes as liver impairment, kidney dysfunction and bone disease.</p> <p>Keywords: Arsenic trioxide, <i>Clarias batrachus</i>, Lethal concentration , Acid phosphatase, Alkaline phosphatase.</p> <p>INTRODUCTION</p> <p>The effect of heavy metals on aquatic organism is currently attracting wide spread attention particularly in studies related to industrial pollution. High toxicity of industrial pollutions have been known since long time, but their hazardous nature as pollution of aquatic environment has been matter of concern only after a large number of deaths of fishes occurring in different areas due to different metals. In aquatic environment, fishes are usually regarded as organisms of choice for assessing the effects of environmental pollution on aquatic ecosystems</p>

Gernhofer *et al.* (2001). Despite progress made in environmental waste management, heavy metals still pose immense health hazards to humans and biota unlike other classes of pollutants, which can be biodegraded and destroyed completely. The name "Arsenic" is derived from the Greek word "arsenikon", which means yellow orpiment. Arsenic compound have been mined and used since ancient times. The extraction of the element from arsenic compound was first reported by Albertus Magnus in 1250 A.D. Emsley (2001) Arsenic, a heavy metal ranks 20th in earth's crust, 14th in sea water and 12th in human body. Arsenic exhibit metallic as well as non-metallic properties. Arsenic is aknown chemical element that has the symbol 'As' and atomic number 33. Its atomic mass is 74.92 and is prevalent in the environment, occurring both naturally and as a result of environment pollution.

Sources of arsenic include treatment of wood using chromate copper arsenate, burning of coal in thermal power plants, operation of gold minning, as treatment of land with arsenical pesticides. Arsenic occurs naturally and its use is possibly aggravated by the use of over powering aquifers and by phosphorous from fertilizers, production of dyes from tanneries, application of some herbicides and insecticides. It is present in effluents from Laundring Tamaki and Frankenbeger (1992) Arsenic, an important environmental contaminant, is present in the aquatic environment as a result of geogenic and anthropogenic processes, Gonzalez *et al.* (2006); Singh and Banerjee (2008). Biochemical characteristics of blood are among the important indices of the status of internal environment of the fish organism (Luskova, 1997). The present study focuses on the impact of arsenic on biochemical profile of *Clarias batrachus*. Arsenic generally exists in the inorganic form in water samples. Under different redox conditions arsenic is stable in the +5, +3, -3, and 0 oxidation states. The pentavalent (+5) arsenic or arsenate species include AsO_4^{3-} , and $H_2AsO_4^-$. The trivalent (+3) arsenic or arsenite species include $As(OH)_3$, $AsO_2(OH)^-$, and AsO_3^- . The pentavalent arsenic

species are predominant and stable in the oxygen-rich aerobic environment, whereas the trivalent arsenic species are predominant in the moderately reducing anaerobic environment such as groundwater.

MATERIALS AND METHODS

Test fish:

Healthy living specimen of teleost, *Clarias batrachus* were collected from local fish market of Meerut. Fish measuring 15 ± 2 cm in length and 60 ± 8 gm in weight were selected for the present study. Selected fishes were acclimatised to the laboratory conditions for period of 15 days.

Preparation of stock solution and determination of 96 hr LC 50 value of Arsenic trioxide:

1gm of arsenic trioxide stock solution was prepared by dissolving arsenic trioxide in 1N HCl under constant heating. The pH was adjusted to 7.4 by adding 1N NaOH dropwise and the solution was filtered by passing through filter paper. For the determination of median tolerance limits or LC 50, different concentrations of arsenic trioxide (20, 30, 40, 50, 60, 70, 80 and 90 mg/l) were prepared from the stock and added in separate glass aquaria containing 50 L of water.

Chemical exposure and Experimental design:

Fishes were divided into 4 equal groups each comprising of 30 fishes. Each group was kept in separate glass aquaria of 250 litre capacity. First group was treated as control group. Fishes of other 3 groups were treated with sub-lethal concentration 8.7mg/l arsenic trioxide for period of 30, 45 and 60 days. Water in the aquariums were renewed after 24 hours and fresh solution of the toxicants were added to bring the concentration to the desired level.

Biochemical studies:

All biochemical studies were performed with the serum of control as well as treated groups of fishes.

Preparation and preservation of serum:

Fish blood was centrifuged at the speed of 3000 rpm. The serum was separated and preserved in the refrigerator at -20°C in the deep freezer. These vials were properly labelled according to the experimental design. Whenever the serum was required, it was first of all brought to the room temperature and then further estimations were done.

1. Determination of Serum Protein

Total serum protein was determined by Kjeldahl's digestion

2. Determination of Serum Cholesterol

Serum cholesterol was estimated with the help of one step method (Wybenga and Pilleggi).

3. Determination of Glucose

Glucose level was estimated by Kit method (End point o-toluidine).

4. Determination of Acid Phosphatase -

According to Kind and King's kit method.

5. Determination of Alkaline Phosphatase:

For the estimation of serum alkaline phosphatase Kind and king's

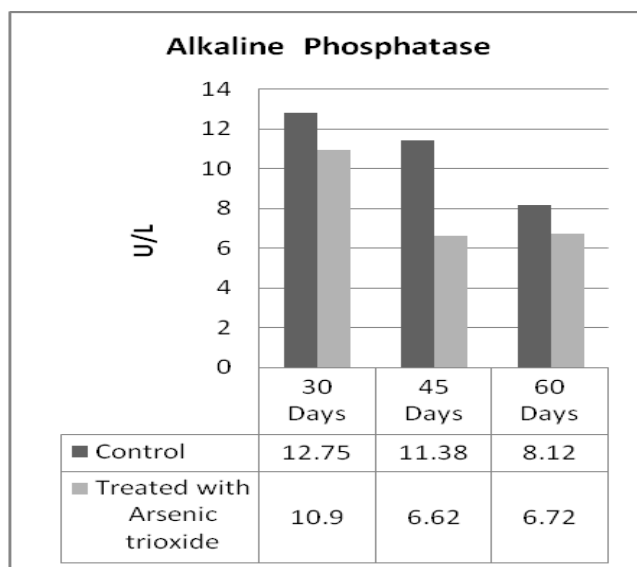
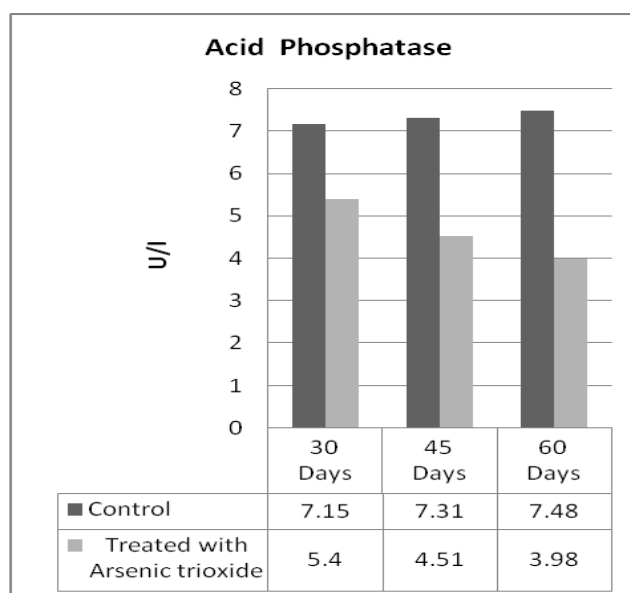
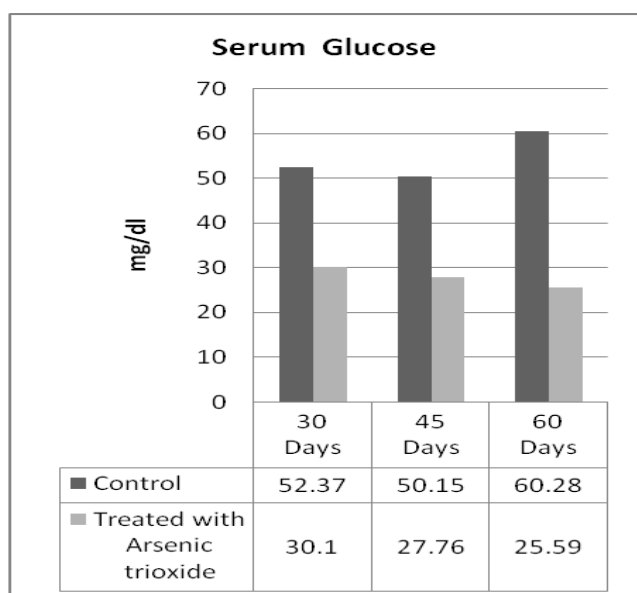
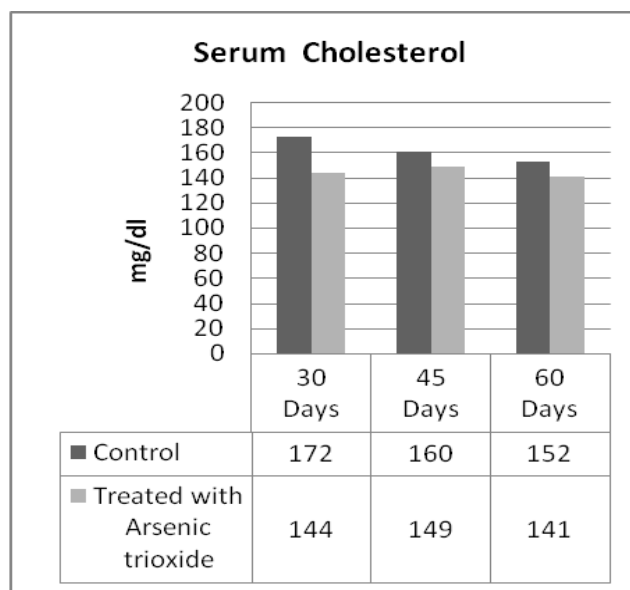
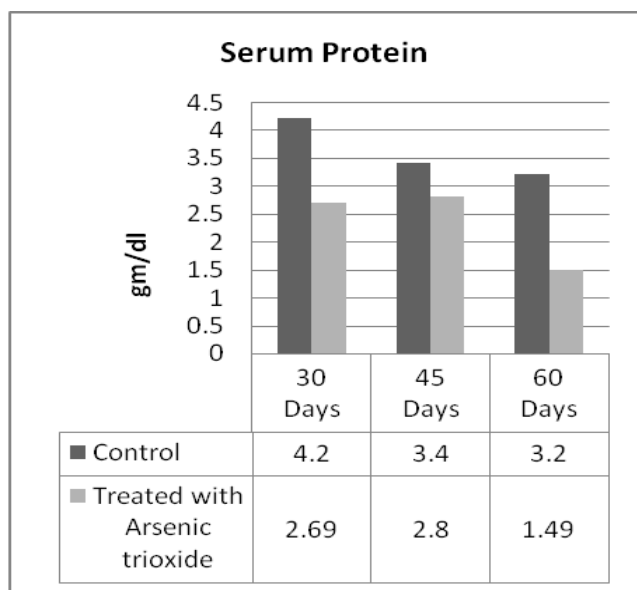
RESULTS AND DISCUSSIONS

The Serum protein was observed to be 2.69 gm/dl after 30 days of PT with arsenic trioxide. The observed value showed difference in parameter when ($p < 0.05$), compared with control values after 30 days. The Serum protein showed increase of 2.80 gm/dl after 45 days of PT with arsenic trioxide. While this parameter showed decline of 1.49 gm/dl after 60 days of PT with arsenic trioxide.

In the present investigation the decrease in Serum protein during acute and sublethal treatment is supported by the reports of Nandi *et al.*, 2005. Palaniappan and Vijayasundaram (2009) suggesting that the decrease in plasma

protein may be due to liver cirrhosis or nephrosis or might be due to alteration in enzymatic activity involved in protein biosynthesis. Pazhanisamy (2002) reported change in total protein content of various tissues in different fishes exposed to different heavy metals. Jana and Bandyopathyay (1981); Jatyajit (1996); Baskaran and Palanichamy (1995) have reported such a reduction in protein content when the fish *Channa punctatus* has been exposed to heavy metals such as mercury, arsenic and lead and *Channa striatus* exposed to mercury cadmium and lead. Gagnon *et al.* (2006) reported that due to metal complex formation, normal functioning of cell is disturbed and that in turn may result in variation on physiological and biochemical mechanisms of animals. Serum Cholesterol was observed to be 144mg/dl after 30 days of PT with arsenic trioxide. The observed value was found to decline when ($p < 0.05$), compared with control values after 30 days. This parameter showed increase 149 mg/dl after 45 days of PT with arsenic trioxide. 60 days of PT with arsenic trioxide showed decline of 141 mg/dl. The observed value showed decrease when ($p < 0.05$), compared with control values after 60 days. Serum Cholesterol showed initial increase after 45 days and decline was noted after 60 days. Heavy metals are known to have hazardous effects on cell structure, especially on the membranes. Therefore, it becomes evident that increase in cholesterol may be the indications of environmental stress. The present findings are in agreement with studies of Murray (1991); Gill and Epple (1993); Sastry and Shukla (1994) who pointed that hyper cholestrolemia observed in *Clarias* may be due to impairment of liver and inhibition of enzymes, which converts cholesterol into bile acid.

Serum Glucose was observed to be 30.10 mg/dl after 30 days of PT with arsenic trioxide. The observed value was found to decline when ($p < 0.05$), compared with control values after 30 days. This parameter showed decline of 27.76 mg/dl after 45 days of PT with arsenic trioxide. While after 60 days of PT with arsenic trioxide



this parameter showed further decline of 25.59mg/dl. The serum glucose showed decline after 45 and 60 days of exposure period Very little attention is paid on effect of arsenic on Serum glucose level in *Clarias batrachus*. Tseng, 2004 reported that chronic exposure of arsenic or its methylated metabolites induced diabetes mellitus in rats and this condition may be responsible for hyperglycemia. Thus an elevation of blood glucose level in the present study during sublethal treatment might be due to gluconeogenesis to provide energy for the increased metabolic demands imposed by arsenic stress. The significant reduction in plasma glucose levels during acute treatment might be

due to hypoxic conditions caused by arsenic leading to an excess utilization of stored carbohydrates.

Acid Phosphatase was observed to be 5.40U/L after 30 days of PT with Arsenic trioxide. The observed value was found to decline when ($p < 0.05$), when compared with control values after 30 days of exposure period. After 45 days of exposure period the observed value showed decline of 4.51 U/L, further decline in parameter was observed to be 3.98U/L after 60 days of PT with arsenic trioxide. Sastry and Gupta (1979) reported elevation in activity of acid phosphatase in *Channa punctatus* under lead exposure. The rise in the activities of acid phosphatase due to lead toxicity leads to hepatocellular damage in the organism Sharma (1999). This increase is associated with liver damage as this enzyme is known to be associated with lysosomal activity. It has been suggested that the acid phosphatase elevation causes proliferation of lysosomes in an attempt to sequester the toxic xenobiotic (Gill and Epple, 1992).

Alkaline phosphatase was noted to be 10.90U/L after 30 days of PT with arsenic trioxide. This value was found to be declined when ($p < 0.05$), compared with control values after 30 days. This parameter showed decline of 6.62 U/L after 45 days of PT with arsenic trioxide. The observed value showed decline in parameter when ($p < 0.05$), compared with control values after 45 days. Alkaline phosphatase showed increase of 6.72U/L after 60 days of PT with arsenic trioxide. Since very less work has been reported directly on this metal but the findings of present work coincides with findings of workers on other heavy metals. The result is in agreement with findings of Agarwal and Sastry (1979) who have observed significant increase in activity of ALP in *Channa punctatus* after 96 hr of post treatment with mercuric chloride. Gill *et al.*, 1991 and Ranjeeta (2008) recorded an increase in alkaline phosphatase activity in *Puntius conchoni* and *Clarias batrachus* under mercuric chloride intoxication and endosulfan exposure. Ilyas *et al.*,

(2007) also noticed the same result in *Labeo rohita*. Such result might be due to increase in osteoblastic activity or intra and extra hepatic obstructions of biliary passage Jyothi and Narayan (1999).

CONCLUSION

In the present study variations in all biochemical parameters were recorded with duration of exposure to experimental chemical Arsenic trioxide. Thus it is indicated that heavy metal arsenic trioxide is causing harmful alterations in biochemical profile of economically important food fish, *Clarias batrachus*.

REFERENCES

- Agrawal MK and Sastry KV (1979) HgCl₂ induced enzymological changes in kidney and ovary of a teleost fish, *Channa punctatus* Bull. Environ. Contamination Toxicol, 22(1-2): 38-43.
- Baskaran P, Palanichamy S, Visalakshi S, Balasubramanian MP (1989) Effects of Mineral fertilizers on survival of the fish *Oreochromis mossambicus*. Environ. Ecol, 7: 463-465.
- Emsley J (2001) Nature's Building Blocks: An A-Z Guide to the Elements. Oxford:Oxford University Press. pp. 43: 513,- 529.
- Gagnon A, Jumarie C, Hontela A (2006) Effects of Cu on plasma cortisol and cortisol secretion by adrenocortical cells of rainbow trout, *Oncorhynchus mykiss*. Aquat. Toxicol, 78: 59-65.
- Gernhofer M, Pawet M, Schramm M, Muller E and Triebkorn R (2001) Ultrastructural biomarkers as tools to characterize the health status of fish in contaminated streams. J. Aquat. Ecosystem, Stress and Recovery, 8:241-260.
- Gill TS and Epple A(1992) Impact of cadmium on the mummichog, *Fundulus heteroclitus* and the role of calcium in suppressing heavy metal toxicity, Comp. Biochem. Physiol, 101:519-523.

- Gill TS and Epple A (1993) Stress-related changes in hematological Profile of the American eel (*Anguillarostrata*). *Exotoxicology and Environmental Safety*, 25: 227-235.
- Gill TS, Tewari H and Pandey J (1991) In vivo and in vitro effects of cadmium on selected enzymes in different organs of the fish, *Barbus conchonus* Ham. (*Rosy barb*) *Comp. Biochem. Physiol*, 100: 501-505.
- Gonzalez HO, Roling JA, Baldwin WS and Bain LJ (2006) Physiological changes and differential gene expression in mummichogs (*Fundulus heteroclitus*) exposed to arsenic. *Aquat. Toxicol*, 77:43-52.
- Ilyas F, Quazi S, Feroz I, Sairy A and Hashmi S (2007) Effect of heavy metal mercuric chloride on the enzyme alkaline phosphatase from fish. *Labeo rohita*. *J. Aqua. Biol*, 22(2): 142-144.
- Jatyajit Hota (1996) Arsenic toxicity to the brain, liver and intestine on a freshwater fish. *Channa punctatus* (Bloch). *Geobios*, 23 :154 -156.
- Jana S and Bondyopadhyay (1981) Efficacy of heavy metal on some biochemical parameters in the fresh water fish, *Channa punctatus* *Environ Ecol*, 5: 488- 493.
- Jyothi B and Narayan G (1999) Certain pesticide induced carbohydrate metabolic disorders in the serum of freshwater fish *Clarias batrachus* (Linn). *Food and Chemical Toxicology*, 37: 417-421.
- Luskova V (1997) Annual cycles and normal values of hematological parameters in fishes. *Acta Sc. Nat. Brno*, 31(5): 70-78.
- Murray RK (1991) Harpers Biochemistry 22nd edition Prentice Hall International Inc pp : 678.
- Nandi D, Patra RC and Swarup D (2005) Effect of cysteine, methionine, ascorbic acid and thiamine on arsenic-induced oxidative stress and biochemical alterations in rats. *Toxicology* ,211:26-35.
- Palaniappan PLRM and Vijayasundurum V (2009) The effect of arsenic exposure and effect of DMSA on protein and lipids of the gill tissues of *Labeo rohita*. *Food.Chem.Toxicol*,47:1752-1759.
- Pazhanisamy K (2002) Studies on the impact of Arsenic on a fresh water fish, *Labeo rohita* (Hamilton). Annamalai university.
- Ranjeeta (2008) Endosulfan toxicity induced alterations in some biochemical parameters of the blood and surface ultrastructure of gill of *Clarias batrachus* (Linn). Ph. D Thesis, Ranchi university, India.
- Sastry KV and Gupta PK (1979) Histopathological and enzymological studies on the effects of chronic lead nitrate intoxication in the digestive system of a freshwater teleost, *Channa punctatus*. *Environ Res*,17: 472-479.
- Sastry KV and Shukla V (1994) Acute and chronic toxic effects of cadmium on some haematological, biochemical, and enzymological parameters in the fresh water teleost fish, *Channa punctatus*. *Acta Hydrochim. Hydrobiol*, 4: 71-176.
- Singh AK and Banerjee TK (2008) Toxic effect of sodium arsenate ($\text{Na}_2\text{HAsO}_4 \cdot 7\text{H}_2\text{O}$) on skin epidermis of air breathing catfish *Clarias batrachus* (L.). *Veterinarski Arhiv*, 78 (1):73-88.
- Sharma B (1999) Effect of carbaryl on some biochemical constituents of the blood and liver of *Clarias batrachus*, a fresh- water teleost. *J. Toxicol. Sci.*24(3):157-164
- Tamaki S and Frankenbeger WT (1992) Environmental biochemistry of arsenic. *Rev Environ Contam Toxicol* , 124: 79-110.
- Tseng C (2004) The potential biological mechanisms of arsenic- induced diabetes mellitus. *Toxicol. Appl. Pharmaco*, 197:67-83.

RESEARCH ARTICLE

Food and feeding of an economically important estuarine fish, *Sillago sihama* (forsskal)

Yeragi SS* and Yeragi SG

Department of Zoology, K.J. Somaiya College of Science, Vidhyavihar, Mumbai-400 077, MS, India

*Corresponding author E-mail : dryeragi@gmail.com.

Manuscript details:	ABSTRACT
<p>Received: 28.04.2015 Revised : 03.05.2015 Accepted: 25.05.2015 Published : 30.06.2015</p> <p>Editor: Dr. Arvind Chavhan</p> <p>Cite this article as: Yeragi SS and Yeragi SG (2015) Food and feeding of an economically important estuarine fish, <i>Sillago sihama</i> (forsskal). <i>Int. J. of Life Sciences</i>, 3(2): 147-151.</p> <p>Copyright: © 2015 Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.</p>	<p>The present investigation revealed that the stomach content of both juvenile (<130 mm SL) and adult (>130mm SL) <i>Sillago sihama</i> (Forsskal) in Mithbav (L.16° 20' N.L.17° 25') estuary of Sindhudurg District, Maharashtra from April 2011 to March 2012. The gut analysis was carried out using frequency of occurrence and point methods. Diatoms were found to be the most preferable food of plant origin in both the life stages like juvenile and adult in all the season, Diatoms, blue-green algae and dinoflagellates constituted main food of plant origin. Diatoms were found to be the most preferable food during starvation period of monsoon due to unexpected water currents. During monsoon, the habit of food and feeding was disturbed for short time. They do not remain in main stream of water but move towards the bank, bay, as well as in lagoons of the estuary. During monsoon it occurred in more than 11.3% of food item by point method. The feeding intensity of juvenile was noticed that on increasing along with increase in size group. The quality and quantity of food items were fluctuated seasonwise to season and juvenile to adult stages. Crustacean including shrimps, crabs, their larvae, copepods eggs, and larval forms comprised the maximum part of the food of animal origin. It was concluded that <i>Sillago sihama</i> in the coastal waters of Mithbav is planktonivorous and feeding on a wide range of food of planktonic and benthic organisms.</p> <p>Key words: <i>Sillago sihama</i>, feeding, Mithbav.</p> <p>INTRODUCTION</p> <p>Creeks, estuaries are an aquatic environment provide numerous commercially important resources that solve the problem of</p>

bread and butter of poor coastal natives. It is but natural, that these resources support the human life in different ways. During monsoon, the sea fishery is totally stopped for short time hence food remains the basic need of mankind for survival. The organisms like fish, shrimps, oysters, clams, mussels, crabs are harvested on large scales, of all trophic levels are available in estuarine ecosystem. Those of second, third and highest levels are known to be rich in protein in terms of quantity and quality particularly in eight amino acids necessary for human being. These amino acids are easily consumed through all through edible resource organisms.

In an aquatic ecosystem, every resource organism can search for favourable habitat to fulfil the demand of their food and safety. There they live, breed to form the nursery ground. Feeding ecology is the significant aspect of the life history strategy of every living species to understand the functional role of the fish within their ecosystem (Patole, 2009). It is an important aspect in an aquacultural practices. *Sillago sihama* (Indian, whiting fish) is the commercially important high market priced protein rich fish for coastal people. However without knowledge of the food requirements, feeding behavior pattern and predator-prey relationships is not possible to understand the predicted changes that might result from any natural or anthropogenic intervention (Yeragi, 1997). *Sillago sihama* (family-Sillaginidae), is commercially and recreationally important estuarine and near shore species of Mithbav coastal zone. It is eco-sensitive fish and in future could be used for exploitation of blue revolution to solve the problem of basic food. It is better to understand their hydrological status, selectivity of food and life history in coastal waters. In pre-monsoon time, their schools are migrating in Mithbav estuary for breeding, spawning as well as for nursing purposes. They always prefer sandy-muddy ground for quick burrow to avoid predation. It is golden yellow in colour with sharp mouth which enhance for burrow in soft sandy regions. It has two dorsal fin, first with 10-15

slender spines, second with one leading spine and 16-27 soft rays. Anal fin is long with two leading spines, with 14-20 soft rays. Family-sillaginidae includes 33 species but only one genus. This is the exceptional family amongst the finfish. Larvae and juveniles are pelagic and feeding on plankton. The body encloses two bladder with two anterior and two posterior extension. Sexual maturity attained about 13-19 cm. SL. Maximum length (SL) observed in this estuary was above 30-35 cm. but commercially marketable size was 20 cm and age is around seven years. The spawning was recorded in the month of July-August. In low salinity the growth is faster and within two month period development reached to fingerling stage. The first sexual maturity was observed at L130-140mm. at the age of one year. The ovulation is once in a year.

The objective of the present study were.

- 1) to explain the stomach contents of juvenile (130 mm<) and adult (130 > mm) of *S.sihama*
- 2) to justify dietary difference amongst variable size classes of juveniles
- 3) to determine seasonal changes in the diets of adults of said species
- 4) to compare feeding habits between the juvenile and adult.

MATERIALS AND METHODS

The study site, Mithbav estuary opening broadly to the west coast of India. Individuals of *Sillago sihama* of 130 mm. in standard length (SL) or more were defined as adults, following histological examination of the gonads. To examine seasonal dietary differences, adults were collected monthly from cast net, gill net and filter (Yendi) net. The fishery was conducted within the estuary from April 2011 to March 2012. The juveniles were collected 3-4 cm. length samples with help of Yendi along with *Penaeus indicus* (white shrimp) on mud-flat region of mangrove swamp. The adults were collected through the bottom fishery of gill-net (Tiyana), cast net and others throughout the year.

In the laboratory (Local), immediately of collection, SL and body weight were measured for each juvenile and adult specimens to the nearest 1mm and 0.1 gm, respectively. Juveniles were sorted into 5 size classes (≤ 10 mm SL, 11-40 mm SL, 41-70 mm SL, 71-100 mm SL and 101-129mm SL). Food items from the stomach contents of each specimen were identified to the lowest possible taxon and the percentage volume of each in the diet visually estimated under a binocular microscope.

RESULTS AND DISCUSSIONS

In the present investigation, many live specimens of *Sillago sihama* were used for examination. The species were dissected out to collect the guts

carefully under hygienic condition and after word they returned to the coastal people. The length range for adult was 14.21-21.80 cm (>130 mm SL) and for Juveniles (<130 mm SL). Bacillariophyceae (diatoms), Cyanophyceae (blue-green algae) and Dinophyceae (dinoflagellates) constitute the main food of plant origin in monsoon and post-monsoon seasons for both the size groups. The juveniles voraciously feed more than adult on cyanophyceae and Dinophyceae. The detritus was observed highest 11.15% in juvenile during monsoon period and then gradually decline upto pre-monsoon time. In monsoon adults also consumed more detritus to avoid starvation due to speedy water current and flood condition. The percentage composition of detritus was 19.15% in monsoon and lowest 1.17% in pre-monsoon.

Table 1 : Seasonal occurrence of food items of adult *Sillago sihama* in the coastal waters of Mithav estuary.

Food Categories	Monsoon %	Post monsoon %	Pre-monsoon %
Diatoms	11.13	29.17	07.18
Blue-green algae	05.11 ⁰	04.28	01.17
Dinoflagellates	04.21	05.19	03.15
Decapods	23.15	24.17	21.15
Mollusca	0.5	01.51	04.17
Foraminifera	01.21	0.15	01.18
Copepods	12.15	26.51	28.91
Polychaetes	01.25	02.12	03.17
Detritus	19.15	02.15	01.17
Animal derivatives	11.19	13.85	15.36
Eggs	08.71	03.15	05.18
Fish	02.24	07.78	08.21

Table 2 : Showing seasonal variation in food items of Juvenile *S. Sihama*.

Food Categories	Monsoon %	Post monsoon %	Pre-monsoon %
Diatoms	17.13	18.24	06.17
Blue-green algae	07.15	09.19	04.45
Dinoflagellates	08.32	09.26	02.18
Decapods	20.18	18.27	17.62
Mollusca	01.71	02.51	08.91
Foraminifera	0.8	02.15	03.31
Copepods	13.18	21.63	24.19
Polychaetes	0.19	01.92	02.61
Detritus	11.15	3.25	03.21
Animal derivatives	07.93	8.26	13.65
Eggs	08.84	03.12	09.18
Fish	03.42	02.22	04.52

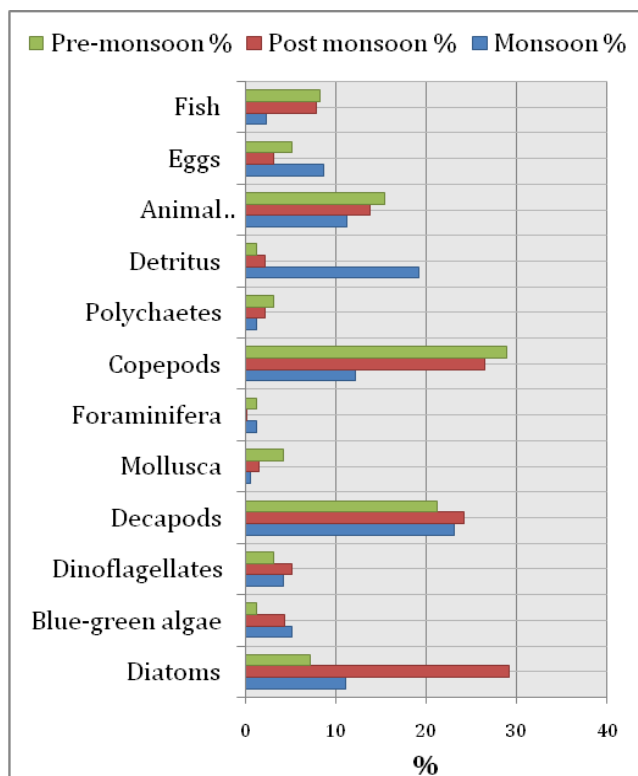


Fig. 1: Histogram Showing seasonal variation in food items of Adult *S. Sihama*

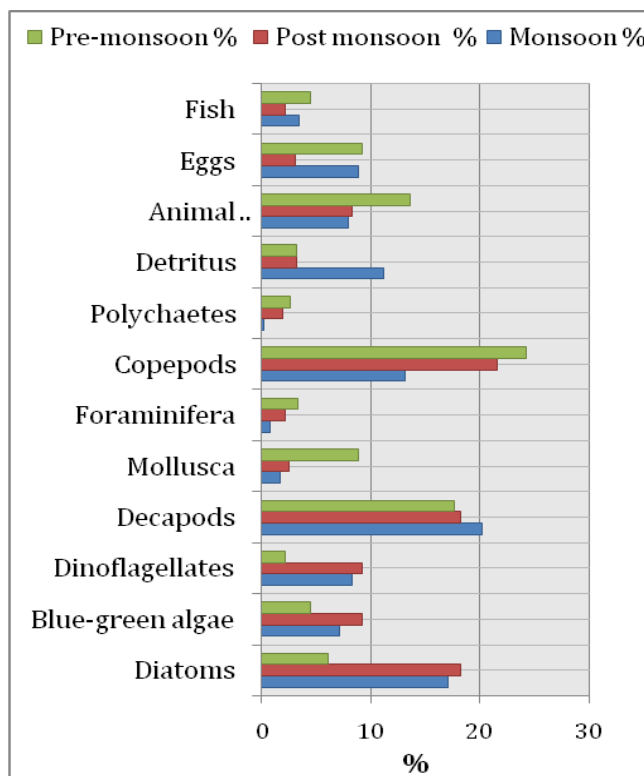


Fig. 2: Histogram showing seasonal Variation in food items of Juvenile *S. sihama*.

During flood time these fish move towards the bank of estuary to avoid unfavorable conditions. The coastal living people catch the fish easily in lagoon, bays and near by the bank with help of cast net. The major food items found in the stomach of *S. sihama* are shown in Table. 1 & 2 In point method, diatoms were found to be the most selective food of plant origin. It was occurred in more than 17.13% in monsoon, 18.24 in post monsoon while decline in pre-monsoon to 6.17%. It was noticed that during pre-monsoon the juveniles voraciously feed on copepods whose percentage composition was 24.19% highest in all the seasons. It is also seen that the Diatoms and copepods were inversely proportion as food items. In adult the parentage of Diatom as food item was highest in monsoon (11.13%) and lowest (7.18%) in pre-monsoon due to availability of copepods. Decapods, like juvenile shrimps, crabs, hermit crab were found to be highest in adult during post monsoon (24.17%) while in juverules maximum in monsoon

(20.18%). The percentage compositions in both the groups were found to be slightly changed from season to season. It also observed that the highest peak was in monsoon followed by post monsoon. Copepods and animal derivatives as a groups contributed about 11.19% monsoon and highest 15.30% in pre-monsoon in adult while in juvenile lowest 7.91% in monsoon and maximum 9.18% in pre-monsoon. The percentage compositions of food items like mollusks, foraminifera, were least in all the seasons. The molluscan food items was highest in both during pre-monsoon because of clams, oysters, mussels, solri, *Perna*, larvae found to be plenty in pre-monsoon season. The juveniles shrimps were detected heights in monsoon and pre-monsoon period. Animal derivatives (eggs, scales, crustaceans appendages, etc) were peak during post monsoon and pre-monsoon. The choises of food items were found to be maximum in post and pre-monsoon their monsoon than. The intensity, of feeding is directly corelated to season

as well as size groups. The juveniles spend more time in feeding than adults. The adults were restricted to the feeding ground and prefer deep water areas, while juveniles spend more time towards the bank of estuary.

From the above mentioned observation, it is clearly understood that this species could be easily cultivated in fish ponds like Shrimp. The *S.Sihama* are tasty fish having high priced therefore the coasted people should use this fish for fish farming to get highest yield production to solve the problem of food. (Yeragi 2004).

The dietary compositions of juveniles of *S.sihama* changed progressively with increasing body size. The change included a shift from the ingestion of small zooplankton, such as calanoid copepods by small juveniles to the consumption of larger benthic prey, such as polychaetes, shrimps, similar to those of adult of the species. The overall feeding habits of juvenile *S. sihama* ≤ 10 and 11-45mm. SL size classes preferred major food items of calanoid copepods. In large size classes, however, this prey item was replaced by polychaetes and mollusks. The results also indicated that the percentage of copepods differed significantly between season wise and size wise.

CONCLUSION

S. sihama is ecologically and economically important estuarine fish. Cultivation of this fish is likely to be profitable because of the consumer demand both in local and export markets. It has high rate of tolerance and planktivores. The growth is very fast. The rate of fecundity is also high. It is advisable to local native that they can easily make the fish farm in the adjacent vacant mud-flat region to get high yield production to solve the problem of required amino acids to mankind. considering high rate of tolerance, fast growth, high population dynamic, it is better used for fish farming as model example.

REFERENCES

- Bam Deo Pandey and Yeragi SG (2000) The importance of live feeds in aquatic seed production. *Aquaculture, Info fish* 4:31-35.
- Patole VM (2009) Biodiversity and ecology of Mangroves in Mochamad creek, Vengurla, Maharashtra, Ph.D. Thesis, University of Mumbai.
- Yeragi SS and Yeragi SG (1997) Species Composition and distribution of Prawn juveniles in Mangroves of Mithbav estuary, Maharashtra state, India *J.Aqua.Biol.Vol.12* (1 &2):16-17.
- Yeragi SG and Yeragi SS (2004) Need of an aquaculture Prospects of marine edible resources in Kharland lagoons of Sindhudurg District. Maharashtra, India. *J.Aqua. Biol.Vol 19*(2):169-174.

RESEARCH ARTICLE

Antioxidant and antimicrobial properties of *Adhatoda vasica* L. Nees

Wankhede TB

Department of Botany, Shri Shivaji Science College, Amravati – 444604, MS, India

Manuscript details:	ABSTRACT
<p>Received: 10.03.2015 Revised : 21.03.2015 Revised received: 13.05.2015 Accepted: 29.05.2015 Published : 30.06.2015</p> <p>Editor: Dr. Arvind Chavhan</p> <p>Cite this article as: Wankhede TB (2015) Antioxidant and antimicrobial properties of <i>Adhatoda vasica</i> L. Nees. <i>Int. J. of Life Sciences</i>, 3(2): 152-156.</p> <p>Acknowledgement: The author is thankful to Dr. N.A. Ghanwate, Department of Microbiology, Sant Gadge Baba Amravati University, Amravati for providing the laboratory facility.</p> <p>Copyright: © 2015 Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.</p>	<p>Plant <i>Adhatoda vasica</i> L. Nees (Acanthaceae) commonly known as Malabar nut is an evergreen two – three m tall shrub, sometime used as hedge, branches opposite and stem yellowish. Leaves simple, 10-20 cm long and 3 to 7.5 cm or sometime much more broad, elliptical, ovate-lanceolate, and tapering towards apex. Inflorescence terminal or sub terminal spikes, flowers white bilabiate and fruits two-valve capsule, which dehisces when mature, or dry. The plant leaves, bark and root known for traditional medicinal use in Ayurveda. The plant parts generally bitter and useful in cough, bronchitis, asthma, skin disease, eczema and scabies. The leaves extensively employed in preparations indicated in respiratory ointments and particularly in cough syrups. An antioxidant is a molecule capable of slowing or preventing the oxidation of other molecules. Oxidation is a chemical reaction that transfer electrons from a substance to oxidizing agent oxidation reaction can produce free radicals, which start chain reaction that damage cells. Antioxidants terminate these chain reactions by removing free radical intermediates and other oxidation reaction by being oxidize themselves. In present investigation preliminary antioxidants evaluated from the experimental plant and antimicrobial sensitivity test carried out against some human pathogenic microbial strains to support out the potential compounds of pharmacognostic interest.</p> <p>Keywords:Antioxidants, antimicrobial sensitivity, pharmacology</p> <p>INTRODUCTION</p> <p>Antioxidants remarkably occur in plants having potential to protect the plant from severe damage. More danger of free</p>

radicals, plants produce more antioxidant. Antioxidants are nutraceuticals whose deficiency states are associated with a variety of dreaded conditions, viz. cardiovascular diseases, diabetic, cataracts, rheumatoid arthritis, Alzheimer's disease and others. The medicinal properties of plants have been investigated in the recent scientific development, throughout the world, due to their antioxidant activities, no side effect and economic viability (Anndy *et al.*, 2003). Flavonoids and phenolic compounds widely distributed in plants which have been reported to exert multiple biological effects, including antioxidant, free radical scavenging abilities, anti-inflammatory, anti-carcinogenic etc. (Miller, 1996). They are also suggested to be iron chelators i.e. the novel natural antioxidant and radical scavenging properties (Boyer *et al.*, 1988; Havsteen 1983). Basic clinical and epidemiological research has suggested a potential protective effect of antioxidant nutrients such as (Vitamin C) Ascorbic acid, Anthocyanin, β carotene, Lycopene, chlorophyll etc. on the risk of cancer cardiovascular diseases and aging (Aviram, 2000). Ascorbic acid is an important antioxidant essential for the normal regulation of the colloidal condition of connective tissue, osteoid tissue, dentine and the intercellular cement substance of the capillaries. It is concerned in the hydroxylation of proline and hydroxyproline as an important constituent of collagens. Severe ascorbic acid deficiency produces scurvy. Lycopene is a powerful antioxidant that categorically retards damage caused to DNA and protein. Lycopene offers distinctly and appreciable much better skin protection against the UV-light than β -carotene. It especially accumulated in the various segments of human body viz. skin, adrenal gland, prostate glands, testes etc. It also renders adequate protection against cancer. Lycopene noticeably arrests the insulin-like growth factor-1 stimulation of cancerous growth (Xianquan *et al.*, 2005). In recent years, numerous studies have shown that anthocyanin displays a wide range of biological activities including antioxidant, anti-inflammatory, anti-microbial and anticarcino-

genic activities, improvement of vision, induction of apoptosis and neuroprotective effects. In addition, anthocyanin displays a variety of effects on blood vessels and platelets that may reduce the risk of coronary heart diseases (Mazza, 2007). Chlorophyll is nowadays a solution of many problems like vitamin deficiency, pollution after effect, countering heart and diabetic diseases. It is prescribed in the form of leaf juices, liquid chlorophyll, chlorophyll tablets and leafy pastes. This is all due to the biochemical nature of chlorophyll which on dissociation and substitution reaction gives rise to various essential organic compounds like hemoglobin and vitamin B12 (Ursula *et al.*, 2005). The phytol tail of chlorophyll dissociates into two β -carotene molecules which is a precursor of vitamin A. Hence, plant pigments especially of medicinal plants act as sources of antioxidants (Hsu *et al.*, 2013). *Adhatoda vasica* is reported to prevent oxidative damage of carbon tetrachloride induced hepatotoxic effect in rats (Pandit *et al.*, 2004). Maurya and Singh, (2010) accounted the highest amount of phenolic compounds which scavenge free radicals and exhibit the greatest antioxidant activity. The positive effect of gamma irradiation on the natural antioxidants of *Justicia Adhatoda* showed the release of phenolic compounds (Rajurkar *et al.*, 2012). The ethanolic extract of *A. vasica* showed high antioxidant activity with cytoprotective potential in cell culture (Mamta *et al.*, 2013). Interestingly, the methanolic and aqueous extract of *A. vasica* has potential phytochemical composition of flavonoids, phenols with antioxidant and cytotoxic effects (Rao *et al.*, 2013). Recently, (Kumar *et al.*, 2014) evaluated pharmacological screening of leaf extract of *A. vasica* against dysentery and diarrhea due to the presence of chemical compounds tannins, alkaloids, saponins and flavonoids.

MATERIALS AND METHODS

Fresh leaves, stem and root are metabolically active parts of the plant and site of synthesis for many chemical compounds hence chosen for analysis. Plant materials collected from Melghat

forest areas brought laboratory cleaned and preserved. Estimation of antioxidants like (Vit-C) Ascorbic acid, Anthocyanin, Lycopene, Chlorophyll from plant materials carried out as per the protocols of Thimmaiah (1999).

The plant parts cut in small pieces, cleaned carefully and washed under tap water to remove impurities followed by shade drying. Dried plant parts crushed in blender, powdered and preserved in airtight bottles. Soxhlet extraction process followed in petroleum ether ethanol, methanol, and acetone and different solvent fractions obtained. Dried extracts were stored in labeled sterile wide mouthed screw capped bottles at 4°C and used for further study (Parekh and Chanda, 2008). The standard pathogenic bacterial and fungal strain obtained from Microbial Type Culture Collection and Gene Bank (IMTECH), Chandigarh, India. The bacteria rejuvenated in Nutrient broth (Hi-media laboratories, Mumbai, India) at 37°C for 18 hrs and then stored at 4°C on Nutrient agar. The fungal organisms were sub cultured on Sabaroud's dextrose agar. Four bacterial strains like gram-negative *Proteus vulgaris* (MTCC-744), *Shigella flexneri* (MTCC-1457), *Salmonella typhimurium* (MTCC-98), gram-positive *Staphylococcus aureus* (MTCC-96) and one fungal pathogen *Aspergillus niger* (MTCC-28) were selected. Disc diffusion method was used for the antibacterial sensitivity test by following the standard methods (NCCLS, 1990). The results were compared with the standard bacterial antibiotics like (10 µg/ml) Tetracycline and Nystatin for fungi.

RESULTS AND DISCUSSION

Determination of antioxidants

Antioxidants mostly known to protect our body from the formation of free radicals. Ascorbic acid is not synthesized in human being and dietary or oral consumption only provide this vitamin. The high quality of ascorbic acid was found in fresh leaves of *Adhatoda vasica* showed 1200 µgm of ascorbic acid content (Table- 1). The normal human body when fully saturated contains about 5000 mg of vitamin C, at which 30mg found in adrenal glands, 200mg in extra cellular fluids & really distributed in varying concentrations throughout the cells at the body. (Danne, 1990). Lycopene is one of the over 600 or more carotenoids pigments. Some studies reported that lycopene could inhibit the growth of cancer and endometric cancers (Rao and Agarwal 2000). The moderate lycopene value of 0.84 µgm found in *Adhatoda vasica* (Table -1). The moderate quantity of anthocyanin was found in fresh leaves of *Adathoda vasica* i.e. 62.25 µgm. The antioxidant activity (Scavenging free radicals metal chelation; protein binding) of anthocyanin including the protection of LDL against oxidation has been demonstrated in a number of *In vitro* systems. (Aviram, 2000) The total chlorophyll content in *Adhatoda vasica* found 0.60 µmg from the fresh leaves of the plant (Table-2). Chlorophyll has anti inflammatory, antioxidant and wound healing properties. It is efficient delivery of magnesium helps the blood to carry oxygen to cell and tissues. Chlorophyll also removes carbon dioxide and carbon monoxide, and has been found to

Table 1: Observations for *Adhatoda vasica*

Sr. No	Name of the compound	Plant part taken for analysis	Weight of plant part	Vol. of extract	Vol. of extract taken for analysis	Absorbance (nm)	Value found in µgm
1.	Ascorbic acid	Leaves	1 g	10 ml	1ml	0.193	1250 µgm
2.	Lycopene	Leaves	1 g	10 ml	1ml	0.028	0.87 µgm
3	Anthocyanin	Leaves	2g	10 ml	1ml	0.249	62.25 µgm
4.	Chlorophyll	Leaves	1g	Total chlorophyll		0.545	0.555 µmg

Table 2: Preliminary antimicrobial sensitivity test of *Adhatoda vasica*

Sr. No.	Solvent Extract	Zone of Inhibition [mm]				
		<i>Proteus vulgaris</i> [MTCC-744]	<i>Shigella flexneri</i> [MTCC-1457]	<i>Staphylococcus aureus</i> [MTCC-96]	<i>Salmonella typhimurium</i> [MTCC-98]	<i>Aspergillus niger</i> [MTCC-281]
1.	Petroleum Ether	08	06	09	09	15
2.	Ethanol	12	11	10	15	17
3.	Methanol	09	08	10	11	13
4.	Acetone	09	13	07	11	12
5.	Tetracyclin [control]	27	29	34	30	-
6.	Nystatin [control]	-	-	-	-	31

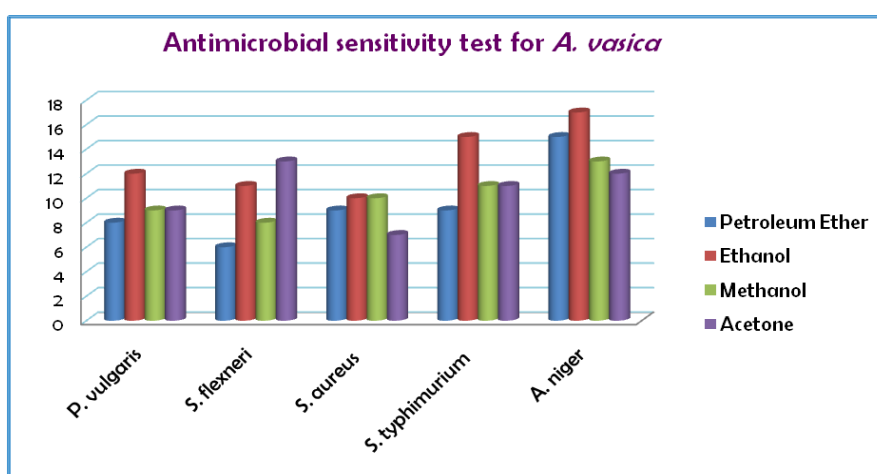


Fig. 1: Analysis of antimicrobial sensitivity test

reduce fecal, urinary, and body odor. Chlorophyll may reduce the binding of carcinogens to DNA in the liver and other organs (Hsu *et al.*, 2013).

Antimicrobial sensitivity of *Adhatoda vasica*

The various types of extract showed consistently positive result against maximum microbial pathogens (Table 2, Fig. 1). The petroleum ether extract of the plant exhibited significant interaction with fungi *Aspergillus niger* with 15mm zone and less with *S. flexneri* pathogens. Subsequently, the ethanol extract found much sensitive with positive results against microorganisms like *P. vulgaris*, *S. aureus*, *S. typhimurium* and *A. niger* with maximum zone of 17 mm. The greenish black coloured extract of methanol was found sensitive to various pathogens with positive interaction like *P. vulgaris*, *S. aureus*, *S. typhimurium* and *A. niger*

with 13mm zone but less reactive to *S. flexneri* with 8mm zone (Rao *et al.*, 2013) The acetonic extract of the plant showed least response against microorganisms *P. vulgaris* and *S. aureus*, and moderate active against *S. flexneri* *S. typhimurium* while greatest against fungi *A. niger* with zone 12mm (Table-2, Fig -1). The ethanol and methanol extract of the plant found more sensitive to all the pathogens as compared to the, petroleum ether and acetone extract. As compare to bacterial strains the fungal strain *A. niger* showed highest and remarkable antifungal sensitivity (Mamta *et al.*, 2013).

CONCLUSION

From the analysis and results it revealed that *Adhatoda vasica* is important medicinal plant with rich antioxidant potential and antimicrobial

sensitivity against pathogenic microorganisms. Its noteworthy that the conventional drugs more sensitive to gram positive bacteria (*S. aureus*) but in present investigation the extracts were more sensitive to gram negative bacteria (*P. vulgaris*, *S. typhimurium*, and *S. flexneri*). Hence, besides conventional drug practice more advance exploration needed for pharmacognostic uses.

REFERENCES

- Anndy B, Ferreira F, Blasina C, Laftop F, Arredondo F, Dajas R and Tripathi PC (2003) Screening of antioxidant activity of three Indian medicinal plants, traditionally used for the management of neurodegenerative diseases. *Ethano pharmacol* 84: 131-138.
- Aviram M (2000) "Review of human studies on oxidative damage and antioxidant protection related to cardiovascular diseases." *Free Radic Res* 33 suppl: S85-97. PMID-11 191279.
- Boyer RF, Clark HM and Laroche AP (1988) Reduction and release of ferritin iron by plant phenolics. *J Inorg Biochem*; 32:171-81.
- Danne JL (1990) *Nutrition Alwanac* Mc. GRAO HLL -4445.
- Havsteen B (1983) Flavonoids, a class of natural product of high pharmacological potency *Biochem Pharmacol* 30:1141-1148.
- Hsu CY, Chao P, Hu S and Yang C (2013) "The Antioxidant and Free Radical Scavenging Activities of Chlorophylls and Pheophytins," *Food and Nutrition Sciences*, Vol. 4 No. 8A, 2013, pp. 1-8.
- Kumar M, Dandapat S, Kumar A and Sinha MP (2014) Pharmacological screening of leaf extract of *Adhatoda vasica* for therapeutic efficacy. *Global Journal of Pharmacology* Vol. 8. No. 4; 494-500.
- Mamta P, Sujata B and Rachna (2013) Antioxidant activity and cytoprotective potential of ethanolic extract of *Adhatoda vasica*. *International Journal of Pharmtech Research*, Vol.5, No. 2; 501-510.
- Maurya S and Singh D (2010) Quantitative analysis of total phenolic content in *Adhatoda vasica* Nees. extract. *International Journal of Pharmtech Research*, Vol.2, No. 4; 2403-2406.
- Mazza G (2007) Anthocyanin and heart health *ANN IST SUPERSANITA* Vol. 43.No.4.369-374
- Miller AL (1996) Antioxidant Flavonoids; structure function and clinical usage. *Alt Med Rev* 1:103-111.
- NCCLS (1990) Manual on "Performance Standards for Antimicrobial Disk Susceptibility Tests". Approved Standard NCCLS Publication, M2-A4, Villanova, PA, USA., (1990 a-b).
- Pandit S, Sur K, Jana U, Debnath PK, Sen S, Bhattacharya D (2004) Prevention of carbon tetrachloride hepatotoxicity in rats by *Adhatoda vasica* leaves. Vol.36, No. 5: 312-313
- Parekh J and Chanda S (2008) Antibacterial activity of aqueous and alcoholic extracts of 34 Indian Medicinal plants against some bacterial species. *Turk. J. Biol*, 32: 63-71.
- Rajurkar NS, Gaikwad KN and Razavi MS (2012) Evaluation of free radicals scavenging activity of *Justica adhatoda*: A gamma radiation study. *International Journal of Pharmacy and Pharmaceutical Sciences*. Vol. 4, Suppl. 4; 93-96.
- Rao KVB, Munjal M, Patnayak A, Karthik L and Kumar G (2013) Phytochemical composition, Antioxidant, Antimicrobial and Cytotoxic potential of Methanolic extract of *Adhatoda vasica* (Acanthaceae). *Res.J. Pharm and Tech*, Vol.6, No.9; 997-1002.
- Rao AV and Agarwal S (2000) Role of Antioxidant Lycopene in Cancer and Heart Disease *Journal of the American College of Nutrition*, Vol. 19, No. 5; 563-569
- Thimmaiah SR (1999) Standard Methods of Biochemical Analysis, Kalyani Publishers, Ludhiyana. ISBN 81-7663-067-5.
- Ursula M, Lanfer-Marquez, Rosa MC, Barros and Patricia Sinnecker (2005) Antioxidant activity of chlorophylls and their derivatives. *Food Research International* Volume 38, Issues 8-9, October-November 2005, Pages 885-891
- Xianquan S, Sti J, Kakuda Y (2005) "Stability of lycopene during Food processing and storage" *J Med Food* 8(4) ; 413-22.

RESEARCH ARTICLE

Seasonal Variation of Physicochemical and Microbial Parameters of water of Nal-Damayanti Sagar Dam, Morshi, Dist. Amravati, MS, India

Ghaware AU and Jadhao RG*

Department of Zoology, Shri Shivaji Science College, Amravati, 444603 Maharashtra. India.

*Correspondent author- Dr. R.G. Jadhao, Associate Professor, Department of Zoology, Shri. Shivaji Science College, Amravati

Manuscript details:	ABSTRACT
<p>Received: 12.04.2015 Revised : 29.04.2015 Accepted: 25.05.2015 Published : 30.06.2015</p> <p>Editor: Dr. Arvind Chavhan</p> <p>Cite this article as: Ghaware AU and Jadhao RG (2015) Seasonal Variation of Physicochemical and Microbial Parameters of water of Nal- Damayanti Sagar Dam, Morshi, Dist. Amravati, MS, India. <i>Int. J. of Life Sciences</i>, 3(2): 157-161.</p> <p>Copyright: © 2015 Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non- commercial and no modifications or adaptations are made.</p>	<p>This study was aimed to estimate current status of Physico-chemical characteristics and level of pollution indicator bacteria and their variation at whole stretch of dam. Some environmental parameters such as Temperature, pH, Turbidity, Dissolved oxygen, Sulphates and Nitrates were monitored. In addition, the microbial analyses involved total viable bacterial and fungal counts. The results of physicochemical parameters showed varied value; from conclusions revealed that large number of sewage drains in Morshi and agricultural discharge is mainly responsible for pollution in Nal-Damyanti Sagar Dam.</p> <p>Keywords: Environment, DO, Sulphate, Agricultural Discharge.</p> <p>INTRODUCTION</p> <p>Water is one of the abundantly available substances in nature, which man has exploited more than any other resources for the sustenance of life. Water of good quality is required for living organisms. Dams are the most important water resource. Unfortunately, the dams are being polluted by indiscriminate disposal of sewage, industrial wastes and human activities. The dams are always the victims of the negative impacts of urbanization. Most water bodies become contaminated due to incorporation of untreated solid and liquid waste. Now a days due to increased human population and man-made conditions, the water quality is deteriorating everywhere Jayabhaye (2008).</p>

Water quality provides current information about the concentration of various solutes at a given place and time. Water quality parameters provide the basis for judging the suitability of water for its designated uses and to improve existing conditions. For optimum development and management for the beneficial uses, current information is needed which is provided by water quality programmers. Prevention of river pollution requires effective monitoring of physicochemical and microbiological parameters Chandra et al. (2006). In most countries, the principal risks to human health associated with the consumption of polluted water are microbiological in nature WHO (1997). The bacteriological examination of water has a special significance in pollution studies, as it is a direct measurement of deleterious effect of pollution on human health APHA (1981). Coliforms are the major microbial indicator of monitoring water quality (Brenner et al., 1993; Grant, 1997). The detection of *Escherichia coli* provides definite evidence of fecal pollution. This work aimed to assessment of the water quality of Nal-damyanti Sagar dam and relates the physicochemical characteristics and microbial quality of water with standard guidelines for safe consumption or usage.

MATERIALS AND METHODS

Water samples for physic-chemical analysis were collected from Nal-damyanti Sagar Dam, geographical coordination Longitude 21° 16' 35" N and Latitude 78° 3' 26" E. Morshi, (M.S) India, during Feb 2010-Jan 2011 in the early morning between 8 am to 11 am in the first week of every month from Feb10-Jan11. The samples were collected in acid washed plastic container from a depth of 5-10 cm. below the surface of water. Samples were aseptically collected in sterile brown bottles (500 ml capacity), transported to laboratory and stored at 4°C until bacteriological analysis completed within 6 h of sampling.

Physicochemical Analyses: The physico-chemical characteristics of the dam water like water temperature, turbidity, pH, Sulphates and nitrate were determined in summer, monsoon and winter according to standard methods APHA (2005); Trivedy and Goel (1984).

Bacteriological analysis: Spread plate method was used for enumeration of Total Viable Bacterial Count at 37°C and Fungal count at 25°C.

RESULTS AND DISCUSSIONS

Physicochemical Analyses

Temperature: The temperature of water affects some important physical properties and characteristics of water such as density, viscosity, conductance, salinity, solubility of dissolved gases etc. and also, chemical and biological reaction rates increase with temperature.

In present investigation, the maximum value was recorded 25.45 ± 1.06 (°C) recorded during summer; minimum value was recorded 19.72 ± 0.58 (°C) recorded during winter. Low temperature recorded in winter may due to lesser solar radiation, low atmospheric temperature and high temperature in summer because of low water level, high solar radiation and clear atmosphere. Similarly, results have been reported by (Anita *et al.*, 2005; Jawale and Patil, 2009).

pH: In present investigation, the maximum value was recorded 7.50 ± 0.35 recorded during summer; minimum value was recorded 6.45 ± 0.02 recorded during monsoon. pH range shows that the water of all sampling sites of dam was slightly alkaline and acidic in nature. High value of pH during summer might be due to low water levels and concentration of nutrient in water. pH shows high significant positive relationship with water temperature. Similar trend was also reported (Narayana *et al.*, 2008; Reddy Vasumathi *et al.*, 2009; Kadam *et al.*, 2007; Anita, 2002).

Table 1 : Seasonal variation in physic-chemical and microbial parameters of Nal-Damyanti Sagar Dam, Morshi (During Jan10-Dec 11)

Parameter	Summer	Monsoon	Winter
Temperature	22.45 ± 1.06	22.77 ± 0.28	19.72 ± 0.58
pH	7.50 ± 0.35	6.85 ± 0.21	6.45 ± 0.02
Dissolve oxygen	6.4 ± 0.06	6.8 ± 0.18	7.3 ± 0.075
Turbidity	42.26 ± 0.47	47.62 ± 1.16	43.56 ± 0.86
Sulphate	17.69 ± 0.18	19.48 ± 0.39	16.03 ± 0.79
Nitrate	0.65 ± 0.024	0.73 ± 0.06	0.67 ± 0.028
Total Bacterial Count	25 ± 4.4	26.5 ± 2.87	15.5 ± 1.65
Total Fungal Count	2 ± 0.70	6.25 ± 1.6	10 ± 1.58

Dissolved oxygen: It is a very important water quality parameter and is also an index of physical and biological processes going on in water. In present investigation, the maximum DO value was recorded 7.3 ± 0.075 mg/l during winter and minimum value 6.4 ± 0.06 mg/l during summer. Kataria et al. (2006) reported that depletion of dissolve oxygen in water is due to high temperature and increased microbial activity, on their study on water quality of Dahod dam, India. The level of DO was found minimum in summer. This is because of the low solubility of gases at high temperature (Hynes, 1978).

Turbidity: Suspension of particle in water interfering with the passage of light is called Turbidity. Turbidity has been long known to hinder disinfection by shielding microbes, some of them perhaps pathogens. In the present investigation, the maximum turbidity value was recorded 47.62 ± 1.16 during monsoon and minimum turbidity value 42.26 ± 0.47 was during summer. These observations were also supported by (Prasanna and Ranjan, 2010, Shraddha et al., 2008; Trivedi et al., 2009). High values of turbidity in monsoon may be due to influx of rain water from catchments area, cloudiness, less penetration of light, washes silts, sand, high organic matter and low transparency due to suspended inert particulate matter. However, low values of turbidity in summer may be due to clear

atmosphere, evaporation of water and high light penetration.

Sulphate: Sulphate is present in fertilizers they contribute to water pollution and increase sulphate concentration in water body. In the present investigation, the maximum sulphate values obtained 19.48 ± 0.39 mg/l during monsoon and minimum value 16.03 ± 0.79 mg/l during winter. Maximum sulphate concentration during monsoon may be due to the dilution and utilization of sulphate by aquatic plants. However, the low sulphate concentration was noted during winter may be due to biodegradation and low water level. Similarly, results have been reported (Reddy et al., 2009; Telkhade et al., 2008; Shanthi et al., 2006).

Nitrate: Nitrate is the most highly oxidized form of nitrogen compounds commonly present in natural waters, because it is a product of aerobic decomposition of organic nitrogenous matter. In the present investigation, maximum values of nitrate obtained 0.73 ± 0.06 mg/l during monsoon and minimum value obtained 0.65 ± 0.024 mg/l during winter season. Nitrate levels in surface water often show a marked seasonal fluctuation with higher concentration being found during monsoon month compared to winter months. Similarly results have been reported (Gohram, 1961, Rajashekhar et al., 2007).

Microbial analysis:

Total bacterial count: High bacterial density in water indicates sewage contamination. As long as *E. coli* is present in water, there is every possibility of the presence of some pathological bacteria in water and this will affect or alter the diversity of organisms and sometime obstruct the aquatic organisms especially fishes and crabs. Bahadoor *et al.*, (2004); Obiri *et al.*, (2003) reported the interaction between coliform bacteria and its aquatic environment. In the present investigation, maximum bacterial count in CFU/ml obtained 26.5×10^4 during monsoon and minimum value obtained 15.5×10^4 during winter season. Continuously increase in population of Morshi around the dam area is mainly responsible for increased level of pollution. Most of sewage water often added from residential area. Higher bacterial population during monsoon is due to increased land run off and higher faecal inputs in to dam from connecting rivers and various sources. An increase in the bacterial level after rainfall was reported by (Shehane *et al.*, 2005). THB load in the present study is significantly correlated with dissolve oxygen.

Total fungal count: In the present research shows that maximum fungal count in CFU/ml obtained 10×10^3 during winter and minimum count value obtained 2×10^3 during summer season. Total fungal count found to be higher in winter season than the respective level found in summer and monsoon. Shridhar and kaveriappa (1989) also observed that the total number of water fungi was lowest during summer season. Occurrence of maximum number of fungal species during winter and spring season in the present study might be due to moderate temperature and slightly higher percentage of organic and inorganic matter.

CONCLUSION

The obtained results of the present study concluded that the water quality along the studied area in Nal-Damyanti Sagar Dam was remarkably influenced by wastewater discharge from drains located on its sides regarding both physicochemical and microbial characteristics. Agricultural and sewage wastes are the key factors in this environmental problem. The water of Nal-Damyanti Sagar Dam is subjected to fecal pollution and continuous monitoring of microbial quality of water is recommended to control the spreading of pathogens transmitted by contaminated water.

REFERENCES

- Anita G, Chandrasekhar SVA and Kodarkarm MS (2005) Limnological studies on Mir Alam Lake, Hyderabad. *Poll. Res.*, (3): 681-687.
- Anitha G (2002) Hydrography in relation to benthic macro-invertebrates in Mir- Alam Lake Hyderabad Andhra Pradesh, India. Ph.D. Thesis submitted to Osmania University. Hyderabad.
- APHA (1981) Standard methods for the examination of water and wastewater. 15th ed. APHA; Washington, DC, USA.
- APHA (2005) Standard methods for the examination of water and waste waters, 21st Edn., Washington, DC. USA.
- Bahadoor N, Patil DN, Baseri SM and Kapadnis BP (2004) Distribution and seasonal variation of microbial faecal pollution indicators and pathogenic bacteria in ground water of city. *Nature Environ. Poll. Tech.*, 3:331-335.
- Brenner KP, Rankin CC, Roybal YR, Stelma GN, Scarpino PV and Dufour AP, (1993) New medium for the simultaneous detection of total coliforms and *Escherichia coli* in water. *Appl. Environ. Microbial.* 59: 3534-3544.
- Chandra R, Singh S and Raj A(2006) Seasonal bacteriological analysis of Gola River water contaminated with pulp paper mill waste in Uttaranchal, India. *Environ. Monit. Assess.* 118: 393-406.

- Gohram (1961) The chemical composition of some waters from Dune slacks at Sand scale, North Lancashire, *J. Ecol.*, 49(1): 79-82.
- Grant MA (1997) A new membrane filtration medium for simultaneous detection and enumeration of *Escherichia coli* and total coliform. *Appl. Environ. Microbiol.* 63: 3526-3530.
- Hynes HBN (1978) The biology of polluted water, Liverpool Uni. Press, Liverpool, 200-204.
- Jawale AK and Patil SA (2009) Physico-chemical characteristics and Phytoplankton abundance of Mangrul dam, Dist-Jalgaon, Maharashtra, *J. Aqua. Biol.*, 24(1): 7-12.
- Jayabhaye UM, Pentewar MS and Hiware CJ, (2008) A study on physico-chemical parameters of a minor reservoir, Sawana, Hingoli District, Maharashtra. *J. Aqua. Biol.*, 23(2): 56-60.
- Kadam MS, Pampatwar DV and Mali RP (2007) Seasonal variations in different physico-chemical characteristics in Masoli Reservoir of Parbhani district, Maharashtra. *J. Aqua. Biol.*, 22(1): 110-112.
- Kataria HC, Singh Arun and Pandey SC (2006) Studies on water quality of Dahod Dam, India. *Poll Res.* 25 (2):553-556.
- Narayana J, Puttaiah ET and Basavaraja D (2008) Water quality characteristics of anjanapura reservoir near Shikaripur, District Shimoga, Karnataka *J. Aqua. Biol.*, 23(1): 59-63.
- Obiri-Danso K, Okore-Hanson A and Jones K (2003) The microbiological quality of drinking water sold on the streets in Kumasi, Ghana. *Lett. Appl. Microbiol.* 37 (4), 334-339.
- Prasanna MP and Ranjan PC (2010) Physicochemical properties of water collected from Dhamra estuary. *Int. J. of env. Sciences* 1(3) pp- 334-342.
- Rajashankar AV, Lingaiah A, Rao Satyanarayana and Piska Ravi Shankar (2007) The studies on water quality parameters of a minor reservoir, Nadergul, Rangareddy district andhra Pradesh. *J. Aqua. Biol.*, 22(1): 118-122.
- Reddy Vasumathi K, Laxmi Prasad K, Swamy M and Reddy Ravinder (2009) Physico-chemical parameters of Pakhal Lake of Warangal district Andhra Pradesh, India. *J. Aqua. Biol.*, 24(1): 77-80.
- Shanthi V, Muthumeena S, Jeyaseeli A and Florence Borgia VJ (2006) Physico-chemical status of Varaga River at Theni district, Tamil Nadu. *J. Aqua. Biol.*, 21(2): 123-127.
- Shehane SD, Harwood VJ, Whitelock JE and Rose JB (2005) "The influence of rainfall on the incidence of Microbial faecal Indicators and the dominant sources of faecal pollution in a Florida river." *Journal of Applied Microbiology.* 98, 1127-1136.
- Shraddha S, Savita D, Praveen J, Shah KW and Vishwakarma R (2008) Statistical evaluation of hydrological parameters of Narmada river water at Hoshangabad city, India. *Environ Monit. Assess.* 143: 195-202.
- Shridhar KR and Kaveriappa KM (1989) Colonization of leaves by water borne hyphomycetes in a tropical stream. *Mycological Research.* (92): 392-396.
- Telkhade PM, Dahegaonkar NR, Zade SB and Lonkar AN (2008) Quantitative analysis of Phytoplanktons and zooplanktons of Masala Lake, Masala, Dist. Chandrapur, Maharashtra. *Environ. Cosr. J.* 9(1 and 2): 37- 40.
- Trivedi P, Bajpai A and Thareja S (2009) Evaluation of water quality: Physicochemical characteristics of Ganga River at Kanpur by using correlation study. *Nature and Science*, 1(6): 91-94.
- Trivedy RK and Goel PK (1984) Chemical and biological methods for water pollution studies, Environmental Publications, Karad, India. 122.
- WHO (1997) Guidelines for Drinking Water Quality: Surveillance and Control of Community Supplies. World Health Organization, Geneva, pp: 3.

RESEARCH ARTICLE

Effect of moisture content on the production of protease by *Fusarium oxysporum* using agroindustrial waste

Vidhale NN and Deshmukh Rupali R

Department of Microbiology, Shri. Shivaji Science College, Amravati 444604, M.S., India

Email: vidhalenana@gmail.com

Manuscript details:	ABSTRACT
<p>Received: 23.04.2015 Revised : 12.05.2015 Revised received: 23.06.2015 Accepted: 25.06.2015 Published : 30.06.2015</p> <p>Editor: Dr. Arvind Chavhan</p> <p>Cite this article as: Vidhale NN and Deshmukh Rupali R (2015) Effect of moisture content on the production of protease by <i>Fusarium oxysporum</i> using agroindustrial waste. <i>Int. J. of Life Sciences</i>, 3(2): 162-166</p> <p>Copyright: © 2015 Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.</p>	<p>The effect of moisture content on the production of protease from <i>Fusarium oxysporum</i> was studied using agro industrial waste as substrates such as dal mill waste, oil mill waste, molasses, fruit waste and vegetable garbage under solid state fermentation. Dal mill waste, oil mill waste and vegetable garbage produced maximum protease activity in presence of all the types of moistures (25%, 35%, 45%, 55%, 65%, & 75%) after 96 hrs of incubation. However molasses and fruit waste gave highest protease production in presence of 55 % and 65% moisture content after 7th day of incubation. Among all the substrate dal mill waste and oil mill waste were promising in being utilized faster for the production of protease enzyme.</p> <p>Keywords: Agoindustrial waste, <i>Fusarium oxysporum</i>, protease</p> <p>INTRODUCTION</p> <p>Protease is one of the most commercial enzyme used in food processing, detergent industry, dairy industry, silver recovery, medical purpose, leather making, meat processing, and chemical industry as well as in waste water treatment (Negi and Benerjee, 2006). This enzyme occurs widely in plants and animals, but commercially proteases are produced exclusively from microorganisms. Molds of the genera <i>Aspergillus</i>, <i>Penicillium</i> and <i>Rhizopus</i> are especially used for producing proteases (Sandhya <i>et al.</i>, 2005). Solid state fermentation has gained tremendous attention for the low cost production of industrially important enzymes by utilization of various types of waste as agro industrial waste. In search for cheaper fermentation processes with a high enzyme yield, SSF was found to be more attractive (Kota <i>et al.</i>, 1999). Different mticroorganisms were utilized for</p>

the production of protease fungi such as *Aspergillus*, *Penicillium*, *Rhizopus*, *Chrysosporium*, *Mucor*, *Scedosporium* and bacteria like *Bacillus licheniformis*, *Bacillus firmus*, *Bacillus alcalophilus*, *Bacillus subtilis*, *Bacillus thuringiensis*, *Streptomyces spp.* etc. Hence study was carried out to study protease production potentiality of Fungus *Fusarium oxysporum*.

MATERIALS AND METHODS

Substrates preparation

Agro industrial waste such as dal mill waste, oil mill waste, Molasses, fruit waste and vegetable garbage were collected and powdered to size about 2 mm in homogenizer and then sieved through 20-40 mesh screens to obtain a particle having diameter between 0.42 to 0.85 mm. Each of such substrate was supplemented with 0.83 gm K_2HPO_4 and 0.16gm $MgSO_4$, 1.5% agar-agar and 10 ml distilled water and autoclaved for 15 min. at 15 lb/inch². Initial utilization of these substrates for production of protease by *Fusarium oxysporum* strain was studied under Solid state fermentation, incubated for 7 days at room temperature (25 to 30°C). Initially 20 g of 50% moistened substrate was sterilized and thoroughly mixed with 1ml spore suspension of 7 day old culture of *Fusarium oxysporum*. This substrate along with spore suspension was poured in sterilized Petri dish and allowed to incubate at room temperature for 7 days.

Enzyme estimation

After every 24 hrs of interval 1gm fermented substrate was harvested from petri plate and transferred to test tube containing 5ml phosphate buffer. The contents were homogenized and centrifuged at 2000 rpm for 30 min to remove all particulate matter. Protease activity was assayed as suggested by Keay and Wrildi (1970). To 1 ml of culture filtrate, 1ml of 2% casein solution was added and the mixture was incubated at 37°C for 10 min. The reaction was terminated by adding

2ml 0.4 M TCA (Trichloro acetic acid), again incubated at 37°C for 20 min. and filtered through Whatman filter paper no1. One ml of the filtrate was added to 5 ml of sodium carbonate (0.4M) and 1ml Folin- Ciocalteus's reagent and incubated at 37°C for 30 min. The Absorbance was measured at 660 nm. in Spectrophotometer .

Effect of Moisture content

All the five substrates were provided with different moisture percents such as 25%, 35%, 45%, 55%, 65% and 75%. Protease production in SSF in all the five substrate under above moisture content was studied. Protease production was estimated from second day of incubation up to 7 days.

RESULTS AND DISCUSSIONS

All the agro industrial waste substrates were provided with moisture 25%, 35%, 45%, 55%, 65% & 75% and incubated for 7 days at room temperature. Dal mill waste, oil mill waste and vegetable garbage produced maximum protease activity in presence of all the types of moistures after 96 hrs of incubation. However molasses and fruit waste gave highest protease production in presence of 55 % and 65% moisture content after 7th day of incubation. Among all the substrate dal mill waste and oil mill waste were promising in being utilized faster for the production of protease enzyme. Results of enzyme estimation are summarized in Table-1 and fig- 1, 2, 3, 4, and 5. The highest enzyme production (121.50 ug⁻¹) was obtained at 60% initial moisture content by *Streptomyces sp.* (N902) (Lazim *et al.* 2009). A similar observation has been reported in case of *Streptomyces sp.* 594 protease production by De Azerodo *et al.* (2005). Study of Germano *et al.* (2003) indicated the requirement of 55 and 63% initial moisture content for maximum proteases production by *Penicillium* LPB- 9 and *A. falvus* (Malathi and Chakroborty (1991) respectively, in SSF. In the study of Chutmanop *et al.* (2008), the optimum initial moisture level was about 50%

Table 1: Effect of moisture percent on production of protease by *Fusarium oxysporum* in a solid state fermentation using agro industrial waste as substrate.

Type of waste	% of moisture	Production of protease ($\mu\text{g/ml}$) at different Incubation period						Mean \pm SD Value
		24 hrs.	48 hrs.	72 hrs.	96 hrs.	120 hrs.	144 hrs.	
Dal mill waste	25	7.2	7.8	7.9	10.9	10.5	10.3	9.1 \pm 1.49
	35	6.3	7.4	8	10.9	10.7	10.3	8.93 \pm 1.78
	45	6.4	7.6	8.1	10.9	10.8	10.6	9.07 \pm 1.78
	55	6.5	7.7	8.4	10.8	10.7	10.4	9.08 \pm 1.65
	65	6.4	7.4	8.2	10.8	10.3	9.2	8.72 \pm 1.55
	75	6.4	7.3	8.3	10.9	10.4	9.1	8.73 \pm 1.60
Oil mill waste	25	6.6	11.1	9	10.9	9.6	9.7	9.48 \pm 1.48
	35	5.8	11.1	10.1	10.8	8.9	9.8	9.42 \pm 1.77
	45	6.2	10.9	10.3	10.7	8.1	10.3	9.42 \pm 1.71
	55	6.7	8.7	10.4	10.8	7.8	10.3	9.12 \pm 1.51
	65	6.4	8.7	10.3	11	8	10.1	9.08 \pm 1.57
	75	7	8.4	10.3	10.9	6.6	5	8.03 \pm 2.07
Molasses	25	2.3	2.8	2.4	4.2	7.4	7.4	4.42 \pm 2.20
	35	1.9	1.6	2.1	5.1	6.2	6.3	3.87 \pm 2.04
	45	1.4	1.3	2	4.5	7.4	7.5	4.02 \pm 2.65
	55	1.3	1.2	2	5.2	7.8	7.8	4.22 \pm 2.86
	65	1	1.1	1.6	4.6	7.7	7.8	3.97 \pm 2.93
	75	2.5	1.3	2.2	4	7.6	7.5	4.18 \pm 2.51
Fruit waste	25	2.2	3.3	3.6	4.9	7.6	7.7	4.88 \pm 2.11
	35	2.2	3.2	4.2	5.6	7.9	8	5.18 \pm 2.21
	45	2.4	2.7	3.5	5	7.1	7.4	4.68 \pm 1.99
	55	2.2	3.4	4.3	5.4	8.5	8.6	5.4 \pm 2.43
	65	2.1	3	3.7	4.9	8.6	8.6	5.15 \pm 2.58
	75	2	3.5	3.8	6.1	8.1	8.2	5.28 \pm 2.35
Vegetable garbage	25	3.7	3.2	6.1	8.1	8.3	9.5	6.48 \pm 2.37
	35	2.8	4.2	6.9	9.8	9.3	8.9	6.98 \pm 2.65
	45	3.3	4.1	6.1	9.1	8.6	6.6	6.3 \pm 2.13
	55	3.5	3.6	7.2	6.9	7.3	7.5	6 \pm 1.74
	65	3	3.9	5.9	9.7	9.7	8.2	6.73 \pm 2.66
	75	3.8	3.5	7.1	8.7	8.7	7.6	6.57 \pm 2.14

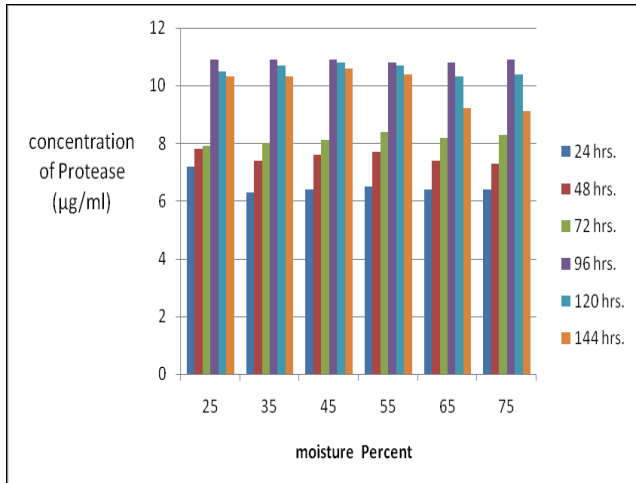


Fig 1: Effect of moisture percent on production of protease by *Fusarium oxysporum* in a SSF using dal mill waste as substrate.

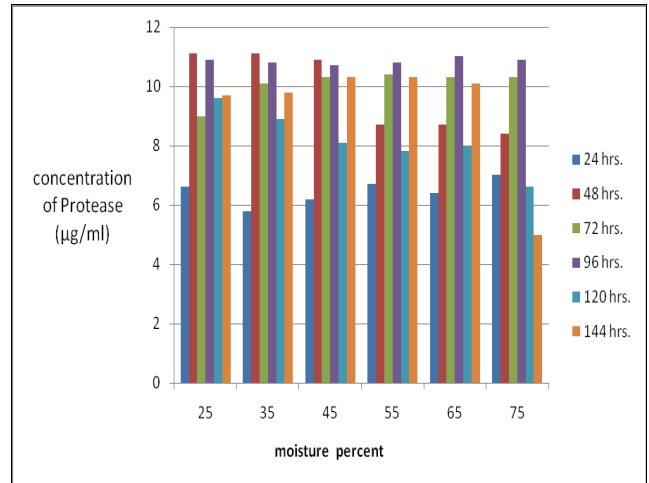


Fig 2: Effect of moisture percent on production of protease by *Fusarium oxysporum* in a SSF using oil mill waste as substrate.

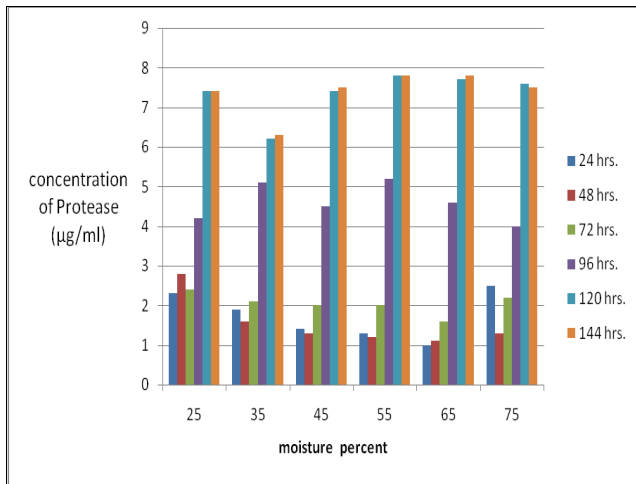


Fig 3: Effect of moisture percent on production of protease by *Fusarium oxysporum* in a SSF using molasses as a substrate.

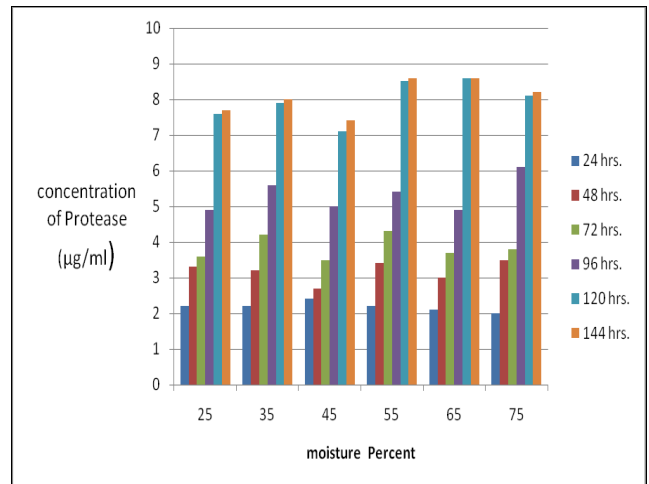


Fig 4: Effect of moisture percent on production of protease by *Fusarium oxysporum* in a SSF using fruit waste as a substrate.

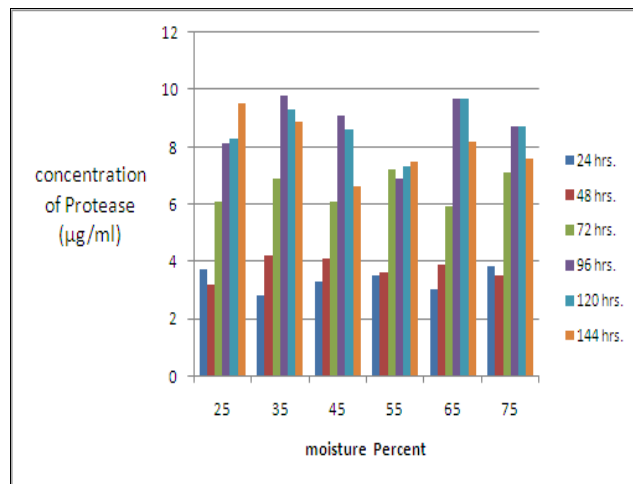


Fig5: Effect of moisture percent on production of protease by *Fusarium oxysporum* in a SSF using vegetable garbage as substrate.

which afforded a high protease activity value.. In the study carried out by Sajeed and Vidhale (2013) the effect of initial moisture on protease production was maximum at initial moisture level of 50% (w/w).

REFERENCES

- Abdus Saboor and Altaff K (1995) Qualitative and Quantitative analysis of Zooplankton population of tropical pond during summer and rainy season. *Ecobiol.*, 7(4):269 – 275.
- APHA (1998) Standard methods for the examination of water and waste water (20th edition). *American Public Health Association*, pp.10-161.
- Chutmanop Jarun, Sinsupha Chuichulcherm, Yusuf Chisti and Penjit Srinophakun (2008) Protease production by *Aspergillus oryzae* in solid state fermentation using agroindustrial substrates. *Chemical Technology*, 1018: 1012- 1018.
- De Azeredo, LAI, Lima MB, de-Coleho RRR and Freire DMG (2006) Thermophilic protease production by *Streptomyces* sp. 594 in submerged and solid state fermentation using feather meal. *J. Appl. Microbiol.* 100: 641–647
- Germano S, Pandey A, Osaka CA, Rocha SN, Soccol CR (2003) Characterisation and stability of proteases from *Penicillium* sp. produced by solid-state fermentation. *Enzyme Microb Technol.*, 32:246–251
- Hadeer Lazim Hadeer, Houda Mankai, Nedra Slama, Insaf Barkallah and Ferid Limam (2009) Production and optimization of thermophilic alkaline protease in Solid state state fermentation by *Streptomyces* sp. CN902. *J Ind. microbial Biotechnol.* vol.36: pp 531-537.
- Keay L and Wildi BS (1970) Proteases of the genus *Bacillus*. I. Neutral proteases. *Biotechnol. Bioeng.*, 12: 179–212. doi: 10.1002/bit.260120205
- Kota KP and Sridhar P (1999) Solid state cultivation of *Streptomyces clavuligerus* for cephamycin C production. *Process Biochem.*, 34:325-328.
- Malathi S and Chakraborty R (1991) Production of Alkaline Protease by a New *Aspergillus flavus* Isolate under Solid-Substrate Fermentation Conditions for Use as a Depilation Agent. *Applied and Environmental Microbiology*, 57(3):712-716..
- Negi S and Benergee R (2006) Optimization of amylase and protease production from *Aspergillus awamori* in single bioreactor through EVOP factorial design technique. *Food Technol Biotechnol.*, 44:257-261).
- Sajeed Ali Syed and Vidhale NN (2013) Protease Production by *Fusarium oxysporum* in Solid-State Fermentation Using Rice Bran. *American journal of microbiological research*,1,(3): 45-47.
- Sandhya C, Sumantha A, Szakacs G and Pandey, A. (2005) Comparative evaluation of neutral protease production by *Aspergillus oryzae* in submerged and solid-state Fermentation. *Biochemical process.*, 40: 2689-2694.

RESEARCH ARTICLE**Rotifers diversity in Kudla Dam near Umri Nanded, MS, India****Bhoyar VV**

Department of Zoology, L. B. D.G. College, Umri, Dist Nanded (MS) India.

Manuscript details:	ABSTRACT
<p>Received: 12.03.2015 Revised : 26.03.2015 Revised received: 13.05.2015 Accepted: 18.05.2015 Published : 30.06.2015</p> <p>Editor: Dr. Arvind Chavhan</p> <p>Cite this article as: Bhoyar VV (2015) Rotifers diversity in Kudla Dam near Umri Nanded, MS, India. <i>Int. J. of Life Sciences</i>, 3(2): 167-170.</p> <p>Copyright: © 2015 Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.</p>	<p>The water quality and nutrients influence population of zooplanktons. The population observed in the distribution of zooplankton is due to abiotic factors (e.g. temperature, salinity DO and other dissolved ions), to biotic factors (e.g. nutrients). Many species of zooplanktons are limited by temperature, dissolved oxygen, salinity and other physico-chemical factors. Zooplanktons constitute an important link in food chain as (primary or secondary consumers) and serve as food for fishes directly or indirectly. Therefore any harm to these will be harmful to the fish populations. The occurrence of zooplankton depends upon its productivity, which is influenced by physico-chemical parameters and the level of nutrients in water. Hence the present study was carried out on kudla dam for a period of one year from Jun 2013 to May 2014. In the present study 19 species of Rotifers were found.</p> <p>Key words: zooplanktons, dam, water quality</p>
	<p>INTRODUCTION</p> <p>Most important energy source to all living organisms is the sun. The solar energy is converted into organic compounds by the process of photosynthesis. This process of converting carbon dioxide and water into carbohydrates is performed by primary producers. These primary producers in water are called as phytoplanktons. These are consumed by zooplanktons, which are again consumed by fishes and so on. These all organisms depend upon water for their growth and reproduction. Therefore any change in the quality of water affects the nature and productivity of these organisms. Change in physico-chemical parameters and nutrient content of water body plays an important role in the production of plankton which act as the natural food of many species of fishes, mainly zooplanktons form important food</p>

source of many fishes and support the necessary amount of diet for the growth of Larval forms (Rahman and Hussain, 2008)). Phytoplankton being the primary producers forms the lowest trophic level in the food chain of freshwater ecosystem , moreover, number and species of phytoplankton's serves to determine the quality of water body (Bahura, 2001). Because of interference of man with nature, the water clarity becomes less and with the addition of nutrients and organic matter, the primary production in water increased. Zooplanktons form the important link between phytoplankton and fishes. The productivity of zooplanktons is influenced by physic-chemical para-me ters and organic contents in water. Zooplanktons feed upon the phytoplankton and make them available to fishes in the food chain (Michael, 1973).

MATERIALS AND METHODS

The samples were collected from four sampling stations of the dams. The present study was conducted in Kudla dam for period of one year i.e.- from Jun 2013 to May 2014. Plank tonic

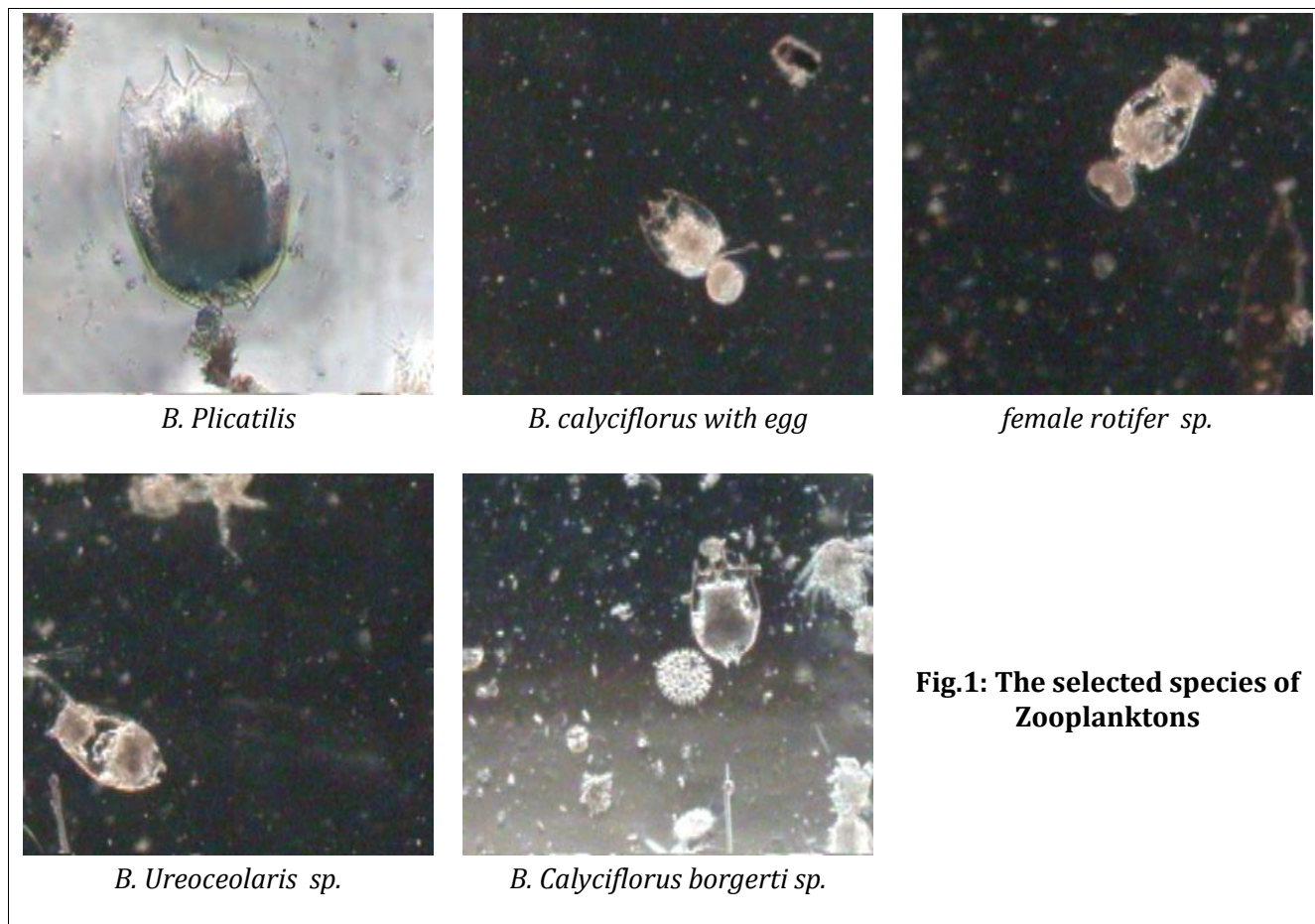
samples were collected on monthly basis from four stations A, B, C and D. Planktons were collected using plankton net made up of bolting silk cloth (Trivedi and Goel, 1986). Filtered samples were fixed and preserved by adding 4% formalin. For counting planktons a Sedgwick Raftor Plankton Counting Cell was used. Identification of planktons was done with the help of methods described by Sehgal (1983); Battish (1992); Dhanapathi (2000) (APHA, 1998).

RESULTS AND DISCUSSIONS

During the study period the high incidence of rotifers in summer season indicating the influence of temperature on positive co-relation between temperature and rotifers population. Similar observations were made by Sinha and Sinha (1983); Singh (2000); Kaushik and Sharma (1994), while working with other reservoir. The maximum density of rotifers was observed during spring peak in April. This was followed by a marked decrease in abundance during the clear water phase in May, 2014.

Table 1 : Zooplanton species abundance

Zooplankton	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Jan	Feb	Mar	Apr	May
Rotifera												
<i>Rotararia</i>	-	-	-	+	+	-	+	-	+	+	-	+
<i>B.calyciflorus</i>	+	+	+	+	+	-	+	+	+	+	+	+
<i>Euchlanis brahmae</i>	+	-	+	+	+	+	-	+	-	+	-	+
<i>Asplanchna</i>	-	-	+	+	-	+	-	+	+	+	+	+
<i>Keratella quadrata</i>	+	+	+	+	+	+	+	+	+	+	+	+
<i>Keralella cochleris</i>	+	+	+	-	-	+	+	+	+	+	+	+
<i>Notholca squamala</i>	-	-	+	-	+	+	+	+	-	+	-	+
<i>Plationus Platulus</i>	-	+	+	+	+	+	+	+	+	+	-	-
<i>Lecane (monostyla) bulla</i>	-	-	-	+	+	+	+	+	+	+	+	+
<i>B. bidentata</i>	+	+	-	-	+	+	+	+	+	+	+	+
<i>L.Papuana</i>	+	+	-	-	+	+	+	+	+	+	+	+
<i>L.doryssa</i>	-	-	-	-	-	+	+	+	+	+	+	+
<i>Pseudoeuchlanis longipedis</i>	-	+	+	+	+	-	+	-	-	+	-	+
<i>B.caudatus</i>	+	+	-	+	+	-	+	+	+	+	+	-
<i>B. c.v. hymani</i>	-	-	+	+	+	+	-	-	-	+	+	+
<i>B.plicatilis</i>	+	-	+	-	-	+	+	+	+	+	+	-
<i>B. quadridentatus</i>	+	+	+	+	-	+	+	+	+	+	+	-
<i>B. durgae</i>	+	+	+	+	+	-	+	+	+	+	+	+
<i>Aaplancha brightwelli</i>	-	-	-	-	+	+	-	+	-	+	-	+



Rotifers play an important role as grazers, suspension feeders and predators within the zooplankton community. The difference in population density of different rotifers can be analyzed by the biotic interactions. The monthly variations of rotifers were ranged between 04 to 07/mL at Station-A, 05 to 15/mL at Station B, 06 to 14/mL at Station C and 06 to 17/mL at Station D, in the year 2013-14. The minimum population rotifers were recorded in the month of August and the maximum population of rotifer was recorded in the month of April, 2014.

High rotifer population indicates population from organic matter due to direct entry of untreated domestic sewage from catchments area (Arora, 1967). Rao (1982) has reported less effect of abiotic factors on the abundance and fertility of pelagic rotifers. Chanadrsekhar (1962) observed

that in summer and monsoon, the factors like water temperature, turbidity, transparency and dissolved oxygen (play an important role in controlling the diversity and density of rotifers).

Abdus and Altaff (1995) studied on qualitative and quantitative analysis of zooplankton population of a tropical pond during summer and rainy season and observed that the zooplankton during the month of July and November shows difference in the density of planktonic species. Dominance of rotifers over other groups has also been reported in other water bodies of the world (Michael 1968; Singh and Sahai, 19768).

The sequence of dominance of various groups was Rotifera > Cladocera > Copepoda. According to George (1966) the abundance of Rotifers is followed by cladocera is an indication of the

eutrophic nature of water bodies. The abundance of Rotifers may be attributed to their dependence on phytoplankton detritus matter and bacteria as food. Higher plankton number was recorded during summer due to an increase in Rotifer number. Present findings also support this.

REFERENCES

- Abdus Saboor and Altaff K (1995) Qualitative and Quantitative analysis of Zooplankton population of tropical pond during summer and rainy season. *Ecobiol.*, 7(4):269 – 275.
- APHA (1998) Standard methods for the examination of water and waste water (20th edition). *American Public Health Association*, pp.10-161.
- Arora HC (1966) Rotifers as indicators of trophic nature of environments. *Hydrobiological*, 27(1 & 2):146 – 149.
- Bahura CK (2001) Phytoplanktonic community of highly eutrophicated temple tank, Bikaner Rajasthan. *J. Aqua Biol.*, 16 (1 &2): 1 -4.
- Battish SK (1992) Freshwater zooplankton of Idierd, Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi.
- Chandrashekhar SVA (1966) Ecological studies on Saroonagar Lake, Hyderabad with special reference to zooplankton Communities, Ph.D. Thesis, Osmania University, and Hyderabad.
- Dhanapati MVSSS (2000) Taxonomic notes on the rotifers from India (1889 – 2000) IAAB publication, Hyderabad, Pg. 175, Publ. No. 101.
- Kaushik S and Sharma N (1994) *J. Environment and Ecology*, 12(2): 429 – 434.
- Michael RG (1968) Studies on Zooplankton of tropical fish ponds. *Hydrobio*, 32:47 – 68.
- Michael RG (1973) A guide to the study of freshwater organism, 2 Rotifera. *J. Madurai, University Suppl.* 1:23-36.
- Rao IS (1982) Ecology of the Manjira Reservoir, Sangareday, Andhra Pradesh Ph.D., /thesis, Osmania University, Hyderabad, Pg. 294.
- Singh DN (2000): *Geobios*, 27 (2-3): 97 – 100.
- Singh SB and Sahai R (1976) Study on some limnological features of Jalvanic pond of Ghorakhpur, *Proc. Nat. Sci India*, 49 (B) II.
- Sinha KK and Sinha DK (1983) *Journal of Ecobiology*, 5(4):299 – 302
- Trivedi RK and Goel PK (1986) Chemical and Biological methods for water pollution studies Pg. 209, *Enviromedia publications Karad*.
- Sehgal KL (1983) Planktonic copepoda of freshwater *Ecosystem Environ. Sci. Series Interprint*, New Delhi, Pg. 1 – 69
- George MG (1966) Comparative planktonic ecology of five fish tanks in Delhi, India, *Hydrobiologia*, 27: 81 – 108.

RESEARCH ARTICLE

Diversity of Zooplankton in some Reservoirs in and around Karwar- Uttara Kannada District Karnataka

Vasanthkumar B¹ and Kapsikar Gangadhar B²,

Dept of Zoology, Govt. Arts and Science College, Karwar- Karnataka

Address for correspondence e mail: ugc.bvk@gmail.com

Manuscript details:	ABSTRACT
<p>Received: 12.03.2015 Revised : 26.03.2015 Revised received: 13.05.2015 Accepted: 18.05.2015 Published : 30.06.2015</p> <p>Editor: Dr. Arvind Chavhan</p> <p>Cite this article as: Vasanthkumar B and Kapsikar Gangadhar B (2015) Diversity of Zooplankton in some Reservoirs in and around Karwar-Uttara Kannada District Karnataka. <i>Int. J. of Life Sciences</i>, 3(2): 171-175.</p> <p>Copyright: © 2015 Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.</p>	<p>The present work was carried out from October 2012 to December 2013. The main aim of the study was to analyse the diversity, evenness and richness of zooplankton species in some Reservoirs of Karwar. The samples were collected during morning hours and were analyzed monthly for different parameter. Results showed that the larval forms were dominant throughout the study period followed by copepods and protozoa group.</p> <p>Key words: Zooplankton, Hydrobiology, Karwar, Diversity, Correlation</p> <p>INTRODUCTION</p> <p>Zooplankton are the heterotrophic component of the plankton community, which range from microscopic to few feet in size. Even though there are many permanent members, much of its composition is made by the juveniles of some animal groups. They form an important link in the aquatic food chain as 'secondary producers' accumulating the carbon for transferring to the other level of food chain, the consumers. Kali River exhibits different type of biotopes such as estuary, backwater, fresh water and mangrove etc. Five reservoirs were selected for study. The environmental parameters of aquatic biotope fluctuate periodically dependable on the three conspicuous seasons, pre-monsoon (February-May), southwest monsoon (June-September) and post-monsoon (October-January). The pre monsoon season is identified by high temperature and salinity, the south- west monsoon season is characterized by heavy</p>

rainfall and the post-monsoon season is known for stable environmental conditions and a high biological productivity rate.

MATERIALS AND METHODS

Water samples were collected on monthly basis. Water samples were collected using a clean plastic container for the study of various physico-chemical and biological parameters.

The plankton samples were collected on monthly basis. Plankton samples were collected by filtering 100 litres of water through plankton net made up of bolting silk. The zooplankton samples were preserved in 5 percent formalin. The preserved samples were brought to the laboratory for qualitative and quantitative analysis and the identification was done with the help of methods described by Pennak (1953), Arora (1963), Sehgal (1983), Battish (1992) and Murugan *et al.* (1998).

Community structure analysis: Three indices were used to obtain the estimation of species diversity, species richness and species evenness.

1. Shannon and Weaver (1949) and Simpson (1949) diversity index value was obtained using the following equation:

$$D = \sum_{i=1}^I P_i^2 (\log P_i) \text{ (Shannon's index)}$$

$$i = I$$

$$D = \sum_{i=1}^I P_i^2 \text{ (Simpson index)}$$

$$i = I$$

Where

Pi = is the proportion of the first species. The proportions are given $P_i = n_i/N$

2. Species richness (D or R1 and R2) was obtained using the equation.

$$R1 = (S - 1) / \log N \text{ (Margalef, 1951)}$$

$$R2 = S \sqrt{n} \text{ (Menhinick, 1964)}$$

Where:

R = is the index of species richness

S = total number of species

N = total number of individuals

3. Species equitability or evenness was determined by using the expression of Pielou (1966) and Sheldon (1969).

$$E1 = \frac{N1}{N0} \text{ (Pileou's Evenness)}$$

$$E1 = \frac{N1}{N0} \text{ (Sheldon evenness)}$$

Where:

N0 = number of species in the sample

N1 = number of abundant species in the sample.

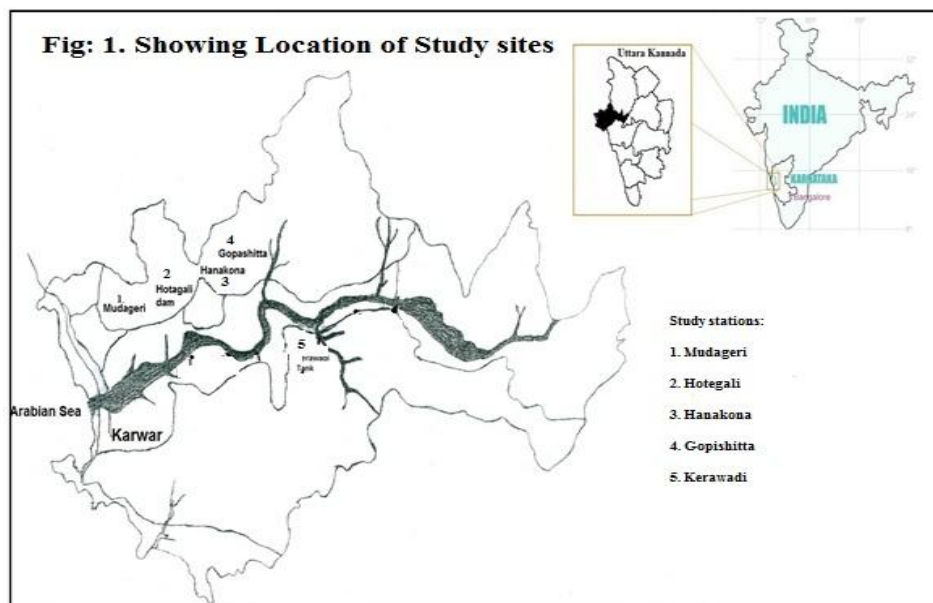


Table: I Study stations with Lat/Long.

Sl.No	Name of the Reservoir	Distance from Karwar	Geographical position
1	Mudageri	13.3km	14°-53'-52.27" N latitude/ 74°-07'-55.06" E longitude
2	Hotegali	15km	14°-54'-13.45" N latitude/ 74°-10'-12.08" E longitude
3	Hanakon	17km	14°-54'-05.29" N latitude/ 74°-11'-28.25" E longitude
4	Gopashitta	19.3km	14°-54'-39.24" N latitude/ 74°-13'-08.24" E longitude
5	Kerawadi	33km	14°-52'-46.32" N latitude/ 74°-15'-46.63" E longitude

RESULTS

Table 2: Monthly Variation in Hydrographical Parameters at Station I

Parameters	O	N	D	J	F	M	A	M	J	J	A	S	O	N	D
Water temp (°c)	28	28	27	26	28.	29	30	32	28	26	28	28	29	27	27
pH	6.9	7.5	7.8	7.6	7.2	7.5	7.5	7.2	7.3	6.6	7.1	6.6	7.4	7.5	7.8
D.O (ml/l)	5.3	6.0	5.5	5.3	5.2	5.1	5.3	5.2	6.1	6.5	5.5	5.8	5.9	5.1	5.4

Table 3: Monthly Variation in Hydrographical Parameters at Station II

Parameters	O	N	D	J	F	M	A	M	J	J	A	S	O	N	D
Water temp (°c)	29	28	29	29	31	28	30	29	26	28	28	28	28	28	29
pH	6.9	7.1	6.5	6.6	7.6	7.8	7.5	7.7	6.8	6.3	6.7	6.7	7.0	6.8	6.7
D.O (ml/l)	5.7	6.1	5.8	5.5	5.4	5.1	4.9	5.1	6.5	6.4	6.6	5.8	5.5	5.5	5.1

Table 4: Monthly Variation in Hydrographical Parameters at Station III

Parameters	O	N	D	J	F	M	A	M	J	J	A	S	O	N	D
Water temp (°c)	29	28	28	28	29	30	29	30	29	28	27	29	29	28	29
pH	6.9	6.9	7.0	6.7	7.3	7.5	7.9	8.2	7.2	6.3	6.4	6.9	7.3	7.5	7.2
D.O (ml/l)	5.6	5.9	5.4	5.4	5.5	5.2	5.5	21.2	6	6.36	6.2	5.7	5.8	5.3	5.3

Table 5: Monthly Variation in Hydrographical Parameters at Station IV

Parameters	O	N	D	J	F	M	A	M	J	J	A	S	O	N	D
Water temp (°c)	27	27	26	26	27	30	30	30	27	26	28	28	29	28	28
pH	7.0	7.3	7.8	7.5	7.3	7.3	7.6	7.7	7.6	6.6	6.9	7.2	7.3	7.5	7.1
D.O (ml/l)	5.3	6.0	5.3	5.6	5.4	5.1	5.0	5.5	6.2	6.3	5.9	5.6	6.2	5.4	5.7

Table 6: Monthly Variation in Hydrographical Parameters at Station V

Parameters	O	N	D	J	F	M	A	M	J	J	A	S	O	N	D
Water temp (°c)	28	27	28	27	29	30	30	30	27	27	26	28	28	28	28
pH	7.6	7.9	7.9	6.3	7.4	7.6	7.9	8	6	6.4	6.5	7.1	6.8	6.7	6.8
D.O (ml/l)	5.7	5.9	5.5	5.2	5.6	5.4	4.9	4.8	6.1	6.4	6.7	6.3	6.0	5.1	5.2

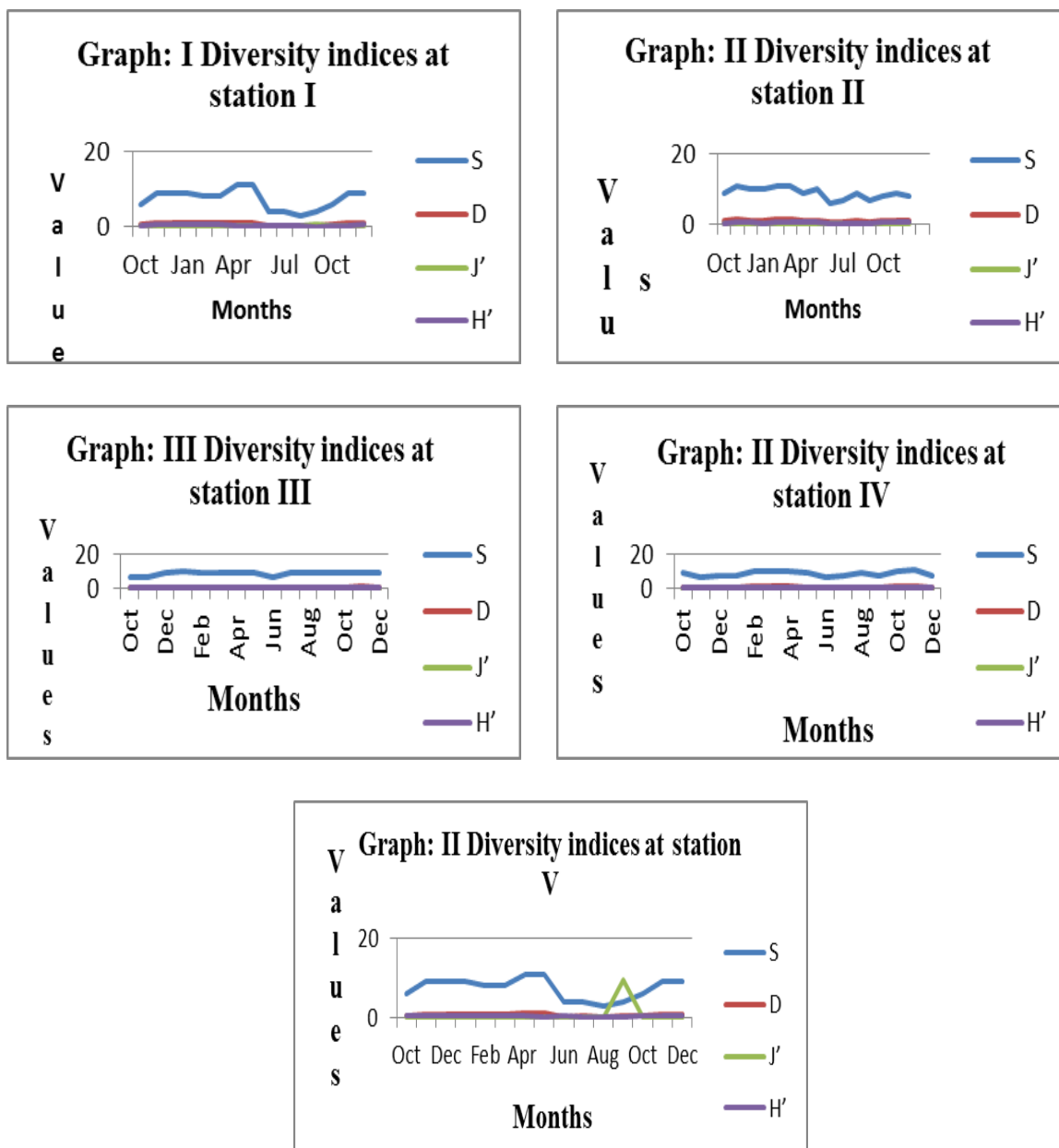


Fig. 1 : Results of Diversity Indices, Species Richness and evenness of zooplankton species

DISCUSSION

Totally 12 groups of zooplankton were identified during the present study. The larval forms ranked 1st (1264-3067/m³) followed by copepods (97-1420/m³) and *protozoa* (41.54/m³). The larval forms constituted about 83-85% of the total species present in all the stations. Copepods constituted 11-13% while *protozoa* constituted only 2-3%. Other groups constituted about 15-

17% of the zooplankton diversity. From the study it is clear that the zooplankton population of the study region was found to be dominated by larval forms followed by copepods and protozoans.

Although zooplankton exists under a wide range of environmental conditions, yet many species are limited by DO, temperature and other physico-chemical factors. According to (Magurran 1988) the diversity indices based on 1. Stable

communities have diversity value and unstable ones have low diversity and 2. Stability in diversity is an index of environmental integrity and wellbeing. Species diversity indices of the Zooplankton groups showed Margalef index (d) ranged between 0.4 and 1.68 and Shannon-Weiner (H) ranged between 0.40 and 0.78 and Evenness (J') ranged between 0.20 and 0.32. In all the stations highest numbers of zooplankton species were observed in the month of May and lowest in the month of October. According to McDonald (2003) the value of diversity indices ranging between 1.5 and 3.4 represents low diversity and species richness value above 3 indicates high diversity. In the present study, the value of Shannon-Weiner (H) ranged 0.40 and 0.78.

REFERENCES

- Arora HC (1963) Studies on Indian Rotifera. Part. II. Some species of the Genus Brachionus from Nagpur. *Journal of Zoological Society India* 15: 112-121.
- Battish (1992) Fresh water Zooplankton of India. Oxford and IBH publishing Co. New Delhi, pp; 233.
- Magurran A (1988) Ecological diversity and its measurements. Princeton University press.
- Margalef R (1951) Diversidad de especies en las comunidades naturales. *Publ. Inst. Biol. Apl.*, 9, 5-27
- McDonald K (2003) The abundance of herbivorous and predatory fishes in relation to *Diadema antillarum* along the west coast of Dominica. *ITME Research reports*. Pp. 11-21.
- Menhinick EP (1964) A Comparison of some species - Individuals diversity indices applied to samples of field insects. *Ecol.*, 45, 859-881.
- Murugan N, Murugaval P and Koderkar MS (1998) *Indian Association of Aquatic Biologists Organization*. (IAAB), 1- 47pp
- Pennak RW (1953) Freshwater invertebrates of the United States. New York: The Ronald Press Company. 769 pp.
- Pielou EC (1966) The measurement of diversity in different types of biological collections. *J. Theoret. Biol.*, 13, 131-144
- Sehgal KL (1983) Planktonic copepods of Freshwater ecosystem. Interprint, new Delhi. 169pp.
- Shannon CE and Weaver W (1949) The mathematical theory of communication Urban. *Univ. Illinois Press. Illinois*. p. 125.
- Sheldon A (1969) Equitability indices: dependence on the species count. *Ecol.*, 50, 466-467
- Simpson EH (1949) Measurement of diversity. *Nature*, 163, 688.

RESEARCH ARTICLE**NF1 gene Analysis: New paradigm by computational approach****Jadhav VA^{1,2} and Laeequr Raheman³**¹Department of Biophysics, D.B. College, Bhokar, Nanded, MS, India -431801²School of Life Science, SRTM University, Nanded, 431606, MS, India.³MGM'S college of CS & IT, Nanded, 431601, MS, India

Manuscript details:	ABSTRACT
<p>Received: 22.04.2015 Revised : 21.05.2015 Revised received: 13.06.2015 Accepted: 16.06.2015 Published : 30.06.2015</p> <p>Editor: Dr. Arvind Chavhan</p> <p>Cite this article as: Jadhav VA and Laeequr Raheman (2015) NF1 gene Analysis: New paradigm by computational approach. <i>Int. J. of Life Sciences</i>, 3(2): 176-180.</p>	<p>Now a day, we are having good stimulation and regulation due to small piece of evolutionarily developed nucleotides working dynamically called gene (eg.NF1). The malfunctioning of NF1 is autosomal dominant condition, contributes a set distinct genetic disorder that that cause tumors to grow along various types of nerve. In addition, it can affect the development of non-nervous tissue such as bone and skin. The NF1 gene, encodes for protein called neurofibromine, belongs to family of protein that serve as negative regulators ras oncogene. The GRD region encoded by exons 20-27a, is the function ascribed region. We are aiming to identify and analyze with structure prediction.</p> <p>Keywords: structure prediction, autosomal dominant, NF1, GRD.</p>
<p>Abbreviation: NF1 : Neurofibromatosis GRD: GAP related domain</p> <p>Copyright: © 2015 Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.</p>	<p>INTRODUCTION</p> <p>Neurofibromatosis type 1(NF1) is one of the most common genetic disorders in human and is characterized by neurofibromas (Riccardi, 1992) It encompasses a set of distinct genetic disorder within neurons, brains, bones, skins etc that cause tumors to grow various nerves and non-nervous tissue. Neurofibromatosis cause to tumor to grow anywhere on or in the body. The NF1 codes for protein neurofibromine, it posses a region that shares a high homology with the family of GTPase-activating proteins, which are negative regulators of RAS function and thereby control cell growth and differentiation (Serra <i>et al.</i>, 1997). NF1 patients show 'two hit' hypothesis with one allele inactivated and another somatically mutated. While considering importance of impaired regulation (Sebastian, 2011)</p>

We are analyzing NF1 locus in benign neurofibromas in NF1 gene. The further research will help in active site prediction and possible outcomes for pharmacokinetics.

MATERIALS AND METHODS

In this analysis, we have retrieved nucleotide as well as protein sequence of NF1 gene from NCBI Gene database (<http://www.ncbi.nlm.nih.gov/gene>) and Protein database (<http://www.ncbi.nlm.nih.gov/protein>). After retrieval of protein sequence of NF1 gene we analyzed primary structure protein using ProtParam tool, which computes various physico-chemical properties of given protein sequence. It is available online in proteomics category of ExPASy server <http://web.expasy.org/protparam>. The secondary structure analysis was carried out by ANTHEPROT integrated protein sequence software. It provides analysis by different

method, out of which GOR and DPM method were used in secondary structure analysis. In consequence we predicted motif, domain, coiled region of NF1 protein sequence using Pfam (<http://pfam.xfam.org/search/sequence>) and Inter Pro Scan (<http://www.ebi.ac.uk/Tools/pfa/iprscan5>). The PDB File format was used to analyze active region i.e. Motif and domain for the basis of protein ligand interaction.

RESULTS AND DISCUSSIONS

Neurofibromin is cytosolic protein with molecular weight of 280kDa. Atomic composition of neurofibromin protein shows 2818 total amino acid. The physico-chemical parameters are specific volume 0.74cm³/g, Extinction Coefficient 282685/m²cm, Estimated half-life >10 hours (E.coli, in vivo), Instability index computed to be 43.35 and GRAVY value is estimated to be -0.129 (Table 1).

Table 1: Physico-chemical parameter of Neurofibromin

Number of amino acids: 2818	Total number of negatively charged residues (Asp + Glu): 299
Molecular weight: 317032.5	Extinction coefficients:
Theoretical pI: 6.90	Extinction coefficients are in units of M ⁻¹ cm ⁻¹ , at 280 nm measured in water.
Total number of positively charged residues (Arg + Lys): 290	Ext. coefficient 282685
Atomic composition:	Abs 0.1% (=1 g/l) 0.892, assuming all pairs of Cys residues form cystines
Carbon C 14141	Ext. coefficient 278810
Hydrogen H 22457	Abs 0.1% (=1 g/l) 0.879, assuming all Cys residues are reduced
Nitrogen N 3813	Estimated half-life:
Oxygen O 4158	The N-terminal of the sequence considered is M (Met).
Sulfur S 144	The estimated half-life is: 30 hours (mammalian reticulocytes, in vitro).
Formula:	>20 hours (yeast, in vivo).
C14141H22457N3813O4158S144	>10 hours (Escherichia coli, in vivo).
Total number of atoms: 44713	Instability index:
	The instability index (II) is computed to be 43.35
	This classifies the protein as unstable.
	Aliphatic index: 94.36
	Grand average of hydropathicity (GRAVY): -0.129

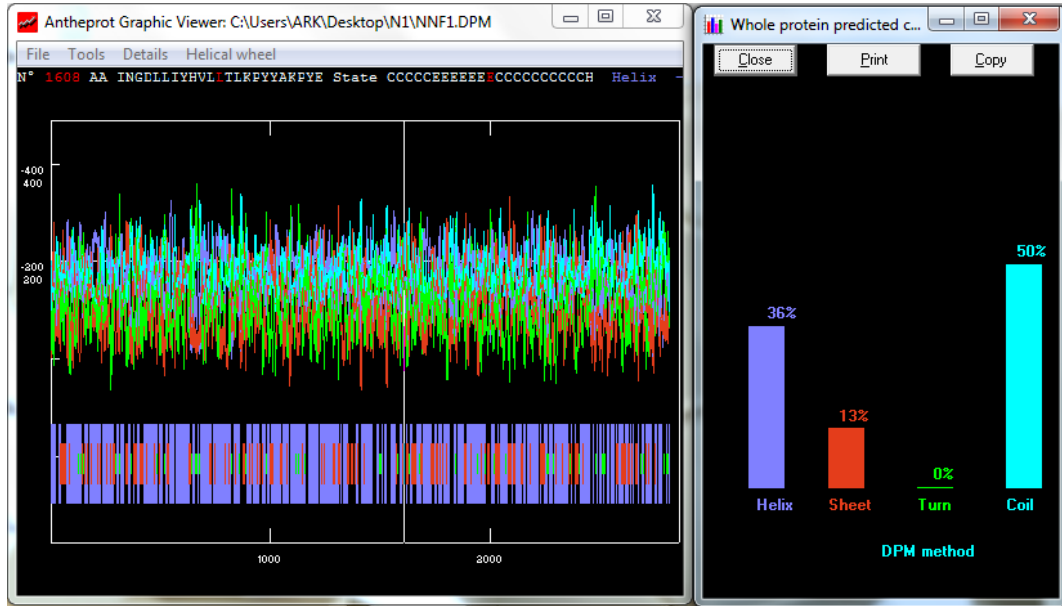


Fig.1: Secondary Structure prediction using Antheptot (a) By DPM method

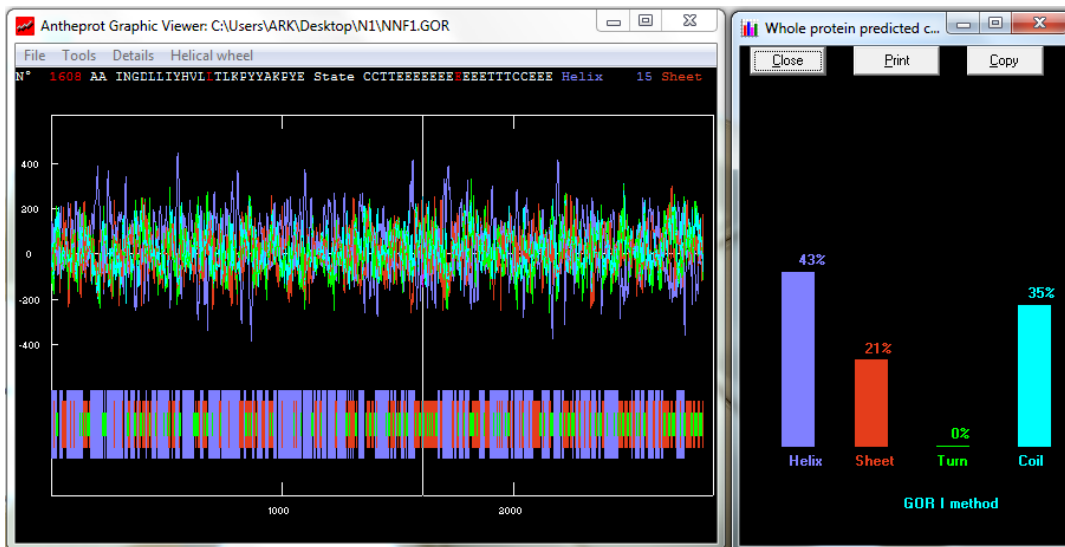


Fig. 1: (b) By GOR methods

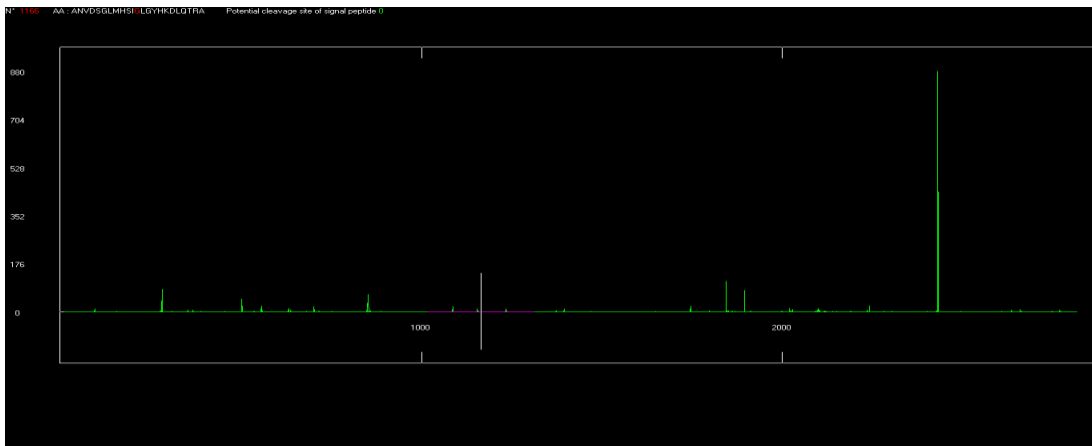


Fig.2: Potential Cleavage Site Using Antheptot (Eukaryotes)

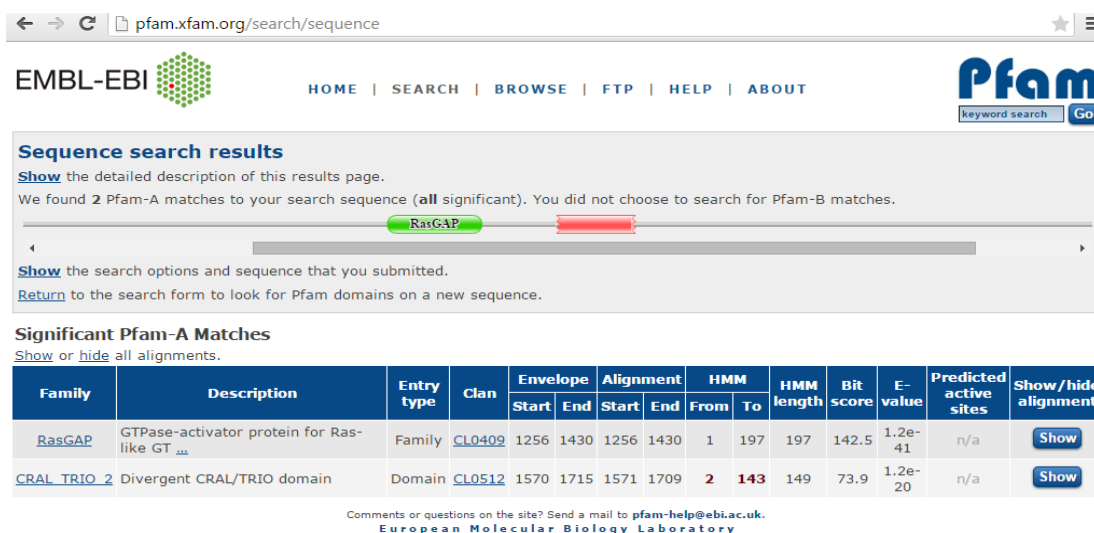


Fig 3: Pfam result showing RasGap related Protein

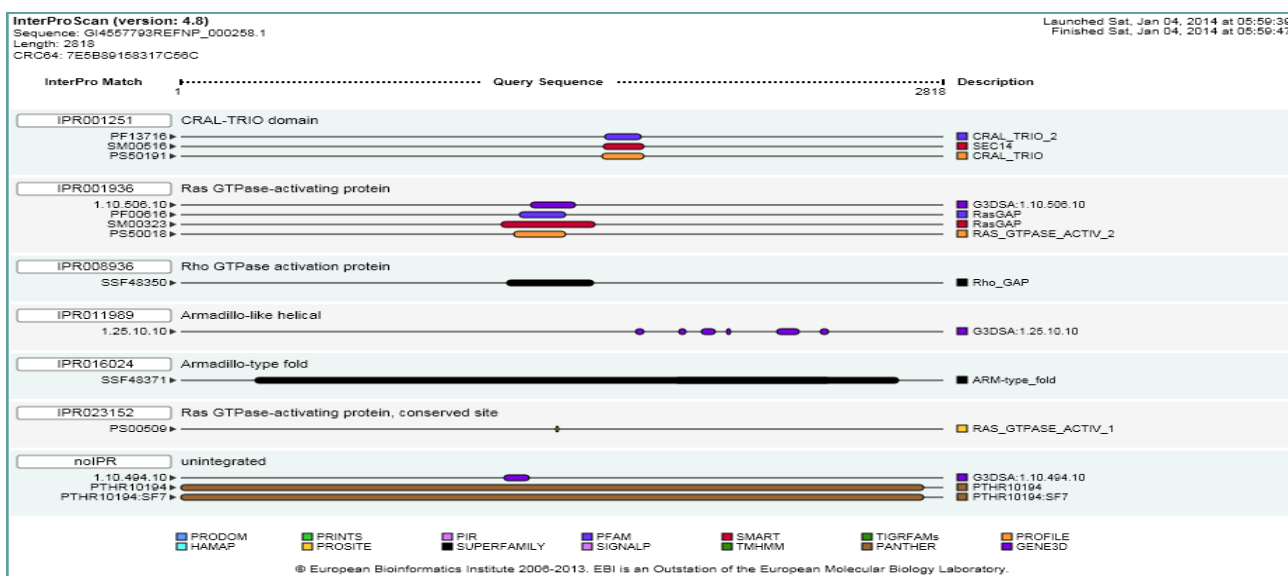


Fig.4: InterPro Scan showing different domain in prote

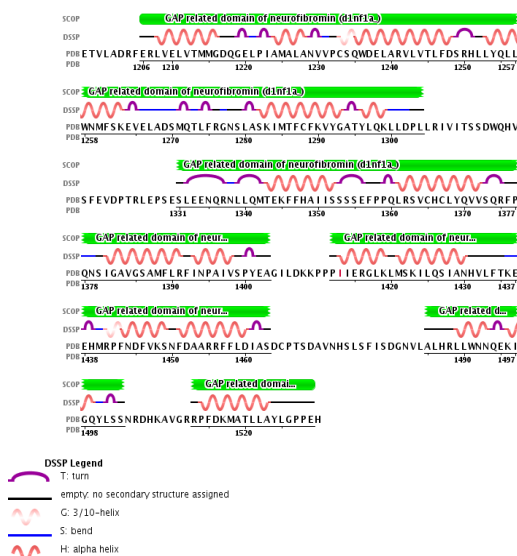


Fig.5: GAP related domain of neurofibromin and 3D view of GAP related domain of neurofibromine using JSmol

The secondary structure comprises alpha helix, β -sheets, turns and coiled region. The analysis shows 36% Helix, 13% Sheet, 0% Turns, 50% coiled region whereas 43% Helix, 21% Sheet, 0% Turns, 35% coiled region according to DPM and GOR method respectively (fig. 1.a & b). The potential cleavage site of signal peptide (Eukaryote) shown in (fig 2). We have found 11 Pfam-A matches to our search sequence (2 significant and 1 insignificant). The graphics below shows (fig.3) the arrangement of matches on our sequence. InterproScan showing different domain in protein (fig.4) GAP related domain of neurofibromin consist of 260 residues (fig.5). 3D Structure of GAP related domain of neurofibromin showing in fig.5.

CONCLUSION

The various protein parameters give significant information about atomic composition, bonding, interactions etc. so it can use to regulate the functioning of neurofibromin protein. This approach is important in new paradigm of computational drug design.

REFERENCES

- Bernards A, Haase VH, Murthy AE, Menon A, Hannigan GE, Gusella JF (1992) Complete human NF1 cDNA sequence: two alternatively spliced mRNAs and absence of expression in a neuroblastoma line. *DNA Cell Biol.*, 11:727-734.
- Deléage G, Combet C, Blanchet C and Geourjon C (2001) ANTHEPROT: integrated protein sequence analysis software with client/server capabilities. *Comput Biol Med.*, 31(4):259-67.
- Li Y, O'Connell P, Breidenbach HH, Cawthon RM, Stevens J, Xu G, Neil S, Robertson M, White R and Viskochil D (1995) Genomic organization of the neurofibromatosis 1 gene (NF1). *Genomics*, 25:9-18.
- Murzin AG, Brenner SE, Hubbard T, Chothia C (1995) SCOP: a structural classification of proteins database for the investigation of sequences and structures. *J.Mol.Biol.*247: 536-540.
- Riccardi VM (1992) *Neurofibromatosis: Phenotype Natural History and Pathogenesis*, 2nd ed. Johns Hopkins Uni. Press, Baltimore, MD.
- Scheffzek K, Ahmadian MR, Wiesmuller L, Kabsch (1998) Structural analysis of the GAP-related domain from neurofibromin and its implications. *EMBO J.* 17(15): 4313-4327.
- Sebastian Laycock-van Spyk, Nick Thomas, David N Cooper, Meena Upadhyaya (2011) Neurofibromatosis type 1-associated tumours : their somatic mutational spectrum and pathogenesis. *Hum Genomics*, 6(6):623-690
- Serra E, Puig S, Otero D, Gaona A, Kruyer H, Ars E, Estivill X, Lazaro C (1997) Confirmation of a double-hit model for the NF1 gene in benign neurofibromas. *AJHG*, 61 (3): 512-519
- Wallace MR, Marchuk DA, Andersen LB, Letcher R, Odeh HM, Saulino AM, Fountain JW, Brereton A, Nicholson J, Mitchell AL, Brownstein BH and Collins FS (1990) Type 1 neurofibromatosis gene: identification of a large transcript disrupted in three NF1 patients. *Science*, 249:181-186.
- Xu G, O'Connell P, Viskochil D, Cawthon RM, Robertson M, Culver M, Dunn D, Stevens J, Gesteland R, White R and Weiss R (1990) The neurofibromatosis type 1 gene encodes a protein related to GAP. *Cell*, 62:599-608.
- Zody MC, Garber M, Adams DJ, Sharpe T, Harrow J, Lupski JR, Nicholson C, Searle SM, Wilming L, Young SK, Abouelleil A, Allen NR, Bi W, Bloom T, Borowsky ML, Bugalter BE, Butler J, Chang JL and Nusbaum C (2006) DNA sequence of human chromosome 17 and analysis of rearrangement in the human lineage. *Nature*, 440:1045-1049.

Web references

- <http://www.ncbi.nlm.nih.gov/gene>
- <http://www.ncbi.nlm.nih.gov/protein>
- <http://web.expasy.org/protparam>
- <http://pfam.xfam.org/search/sequence>
- <http://www.ebi.ac.uk/Tools/pfa/iprscan5>

SHORT COMMUNICATION**Unusual sighting of Yellow- wattled Lapwing (*Vanellus malabaricus*) in Lucknow District, Uttar Pradesh, India****Kumar Adesh * and Kanaujia Amita**

Biodiversity & Wildlife Conservation Lab, Department of Zoology, University of Lucknow, Lucknow, UP, India

*Corresponding author Email: adesh.science@gmail.com

Manuscript details:	ABSTRACT
<p>Received: 26.04.2014 Accepted: 10.05.2015 Published : 30.06.2015</p>	<p>The present study concerns the survey of Yellow-wattled Lapwing in Lucknow district. It is a wader and its habitats preferences includes any sort of open ground, dry fields and the largest concentrations are found in and near wetlands. It is medium-sized pale brown waders with a black crown which is separated from the brown on the neck by a narrow white band and large yellow facial wattles. It is obligate visual forager, meaning catch its prey at the substrate boundary layers, by picking small invertebrates from the surface or from low vegetation cover. Yellow-wattled Lapwing plays a prominent role in ecosystems. The Yellow-wattled Lapwing contributes to maintaining ecosystem food chain because they regulate and maintain the populations of many invertebrates (Pests) which are harmful for agricultural crops. Yellow-wattled Lapwing is a good example of territorial and social behaviour. The present survey was carried out at winter and summer season during February 2014- March 2015.</p> <p>Keywords: Ecosystem, Yellow-wattled Lapwing, Social behavior.</p>
<p>Editor: Dr. Arvind Chavhan</p> <p>Cite this article as: Kumar Adesh and Kanaujia Amita (2015) Unusual sighting of Yellow-wattled Lapwing (<i>Vanellus malabaricus</i>) in Lucknow District, Uttar Pradesh, India. <i>Int. J. of Life Sciences</i>, 3(2): 181-184.</p>	
<p>Acknowledgement: Thanks are due to the to Prof. Madhu Tripathi, Head, Department of Zoology, University of Lucknow, Lucknow for her valuable assistance and providing necessary facilities to carry out the research work. We express our sincere gratitude to Dr. Ashish Kumar, Asst. Prof, Dept. of Zoology, University of Lucknow, for their valuable support.</p>	
<p>Copyright: © 2015 Author(s), This is an open access article under the terms of the Creative Commons Attribution-Non-Commercial - No Derivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.</p>	<p>INTRODUCTION</p> <p>Yellow- wattled Lapwing is prominent and unique wader with a black crown, white supercilium, large yellow facial wattles and tail is white, tipped black belong to family Charadriidae and order Charadriiformes. The long legs are yellow in colour. The breeding season is March-May to the lead of the monsoon. Its habitats preferences include any sort of open ground, dry fields and the largest concentrations are found in and near wetlands fringes (Kumar, 2015). They make short distance movement in</p>

response to rain. It is obligate visual forager, meaning catch its prey at the substrate boundary layers, by picking small invertebrates from the surface or from low vegetation cover. Yellow-wattled Lapwing plays a prominent role in ecosystems. The Yellow-wattled Lapwing contributes to maintaining ecosystem food chain because they regulate and maintain the populations of many invertebrates (Pests) which are harmful for agricultural crops. Yellow-wattled Lapwing is a good example of territorial and social behaviour. There appears to be slightly decreases in its status over the past few decades and is considered as least concerned bird. Only few works have done by several workers on Yellow-wattled lapwing (Jayakar and Spurway, 1965; 1968; Dhindsa, 1983; Gupta and Kaushik, 2010; Santharam, 1980; Sethi et al., 2010). Though, no one has studied Yellow-wattled Lapwing so far in Lucknow in context of status, ecology and threats, hence the present study was primarily effort.

MATERIALS AND METHODS

The Geographic coordinates of Lucknow is 26.8470° N and 80.9470° E. The study area involves Lucknow and its associated areas (up to 100 km). The capital of Uttar Pradesh is situated 123 meter above sea level. In summer

temperature ranges from 25-45°C while in winter from 2-20 °C, the average annual rainfall is about 896.2 mm (35.28 inch). Lucknow covers an area of 2528 sq.km.

Ecological survey of Yellow-wattled Lapwing was carried out at a fixed time- interval from February 2014- March 2015. Species was observed and monitored twice in a day in the morning and evening hours. Observations and monitoring were done with the aid of an Olympus 10x50 binocular and photography was done with 60 D SLR Cannon camera.

RESULTS AND DISCUSSIONS

The yellow-wattled lapwing (*Vanellus malabaricus*) is a lapwing and a group of largish waders in the family Charadriidae that is endemic to the Indian Subcontinent. Dr. Amita Kanaujia along with research scholar Mr. Adesh Kumar had gone to wetlands survey and bird watching on 2nd February 2014 at Magaiyapurva Jheel, Gosaiganj (N 26°38.707' and E 081°03.237') in district Lucknow of Uttar Pradesh. They saw a flock of medium size birds looking rather common Myna or others waders but with long yellow legs and brown colour (Fig. 1 & 2). For few moments they show like little ringed plover but size was larger than plover. On comparison



Fig 1: Yellow- wattled Lapwing in resting and in flight



Fig 2: A flock of Yellow- wattled Lapwing

with colour plates in the Pictorial Guide of Ali & Ripley (1995), Grimmett *et al.*, (2011), Ali (2002) identified the waders as Yellow- wattled Lapwing (*Vanellus malabaricus*). Total 31 lapwings were sighted here. However Yellow- wattled Lapwing was not sighted before at this site. Thereafter a follow up was kept every year at the Magaiyapurva Jheel, Gosaiganj in winter and sighted 46 lapwings. Kumar (2015) observed nesting biology of Yellow- wattled Lapwing in agricultural environment of Punjab.

On 17th January 2015, the authors visited a wetland near Sanjay Gandhi Post Graduate Institute of Medical Science (SGPGI, N 26°44'24.6" and E 080°57'16.2") and Behda pond in Nagar Chaungwa, Mahona (N 27°05.700' and E 080°53.980') in Lucknow district of Uttar Pradesh. Again authors visited Sanjay Gandhi Post Graduate Institute of Medical Sciences (SGPGI) and Behda pond on 2nd February 2015 and 17th March 2105. Then we visited the backside of the Deen Dayal Upadhyaya Park (SGPGI) and we were delighted to see a flock of around 25-35 Yellow- wattled Lapwing and 22-40 from Behda pond. Although Yellow- wattled Lapwing seen in past but very little in number but these study sites number were quite much. A study on threats to nests of Yellow- wattled Lapwing in Kurukshetra has performed by Gupta and Kaushik (2012). A.S.F.L Lok and Subaraj

(2009) studied the status of Lapwings in Singapore.

CONCLUSION

The gathering of large number of Yellow-wattled Lapwings at above study sites in Lucknow for feeding, resting and roosting is due to the abundance of food such as macrophytes, macrobenthic organisms and insects. Accessibility to food resources and availability of exposed mudflats and shorelines of wetlands provides an ideal location feeding and roosting. Still, no one has studied Yellow-wattled Lapwing so far in Lucknow in context of status, ecology and threats; hence the present study was primarily effort. Preliminary study performed in the discussed area would reveal a baseline data which is important for further research and conservation.

REFERENCES

- Ali S and Ripley SD (1995) A Pictorial Guide to the Birds of the Indian Subcontinent. Bombay Natural history society, Mumbai.
- Ali S (2002) The Book of Indian Birds. Bombay Natural History Society and Oxford University Press, Mumbai.

- Dhindsa MS (1983) Yellow-wattled Lapwing: a rare species in Haryana and Punjab. *Pavo*, 21(1-2):103-104.
- Grimmett R, Inskipp C and Inskipp T (2011) Birds of the Indian Subcontinent. London: Oxford University Press.
- Gupta R C and Kaushik T K (2010) On the causative factors responsible for the pathetic plight of Yellow wattled Lapwing in Kurukshetra suburbs. *Journal of Nature Conservation*, 22 (2):181-187.
- Gupta R C and Kaushik T K (2012) Spectrum of threats to nests of Yellow-wattled Lapwing *Vanellus malabaricus* in Kurukshetra outskirts-a case study. *Journal of Applied and Natural Science* 4 (1): 75-78.
- Jayakar SD and Spurway H (1965) The Yellow wattled Lapwing *Vanellus malabaricus* (Boddaert), a tropical dryseason nester. II. Additional data on breeding biology. *Journal of Bombay Natural History Society*, 62:1-14.
- Jayakar SD and Spurway H (1968) The Yellow wattled Lapwing *Vanellus malabaricus* (Boddaert), a tropical dryseason nester. III. Two further Seasons' breeding. *Journal of Bombay Natural History Society*, 65:369-383.
- Kumar C (2015) First record of a regularly occupied nesting ground of Yellow-wattled Lapwing, *Vanellus malabaricus* (Boddaert) in agricultural environs of Punjab with notes on its biology. *Journal of Entomology and Zoology Studies*, 3 (1): 129-134
- Lok A S F L and Subaraj R (2009) Lapwings (Charadriidae: Vanellinae) of Singapore. *Nature in Singapore*, 2: 125-134.
- Santharam V (1980) Some observations on the nests of Yellow-wattled Lapwing, stone Curlew, Blackbellied Finch-Lark and redwinged Bush-Lark. *Newsletter for birdwatcher*, 20 (6-7): 5-12.
- Sethi VK, Bhatt D and Kumar A (2010) Hatching success in Yellow-wattled Lapwing *Vanellus malabaricus*. *Indian Birds*, 5 (5):139-142.